

# Somatic mutations in tumors and normal tissues

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 @nlbigas

- Identifying Cancer Drivers
- Understanding Neutral Mutagenesis
- Identifying Clonal Hematopoiesis Drivers
- Chemotherapy effect in hematopoiesis
- Cancer Promotion



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Which are the genes that drive tumorigenesis upon mutations?



# Accumulated literature on cancer genes for decades



**BRCA1**

**TP53**

**MYC**

**APC**

**EGFR**

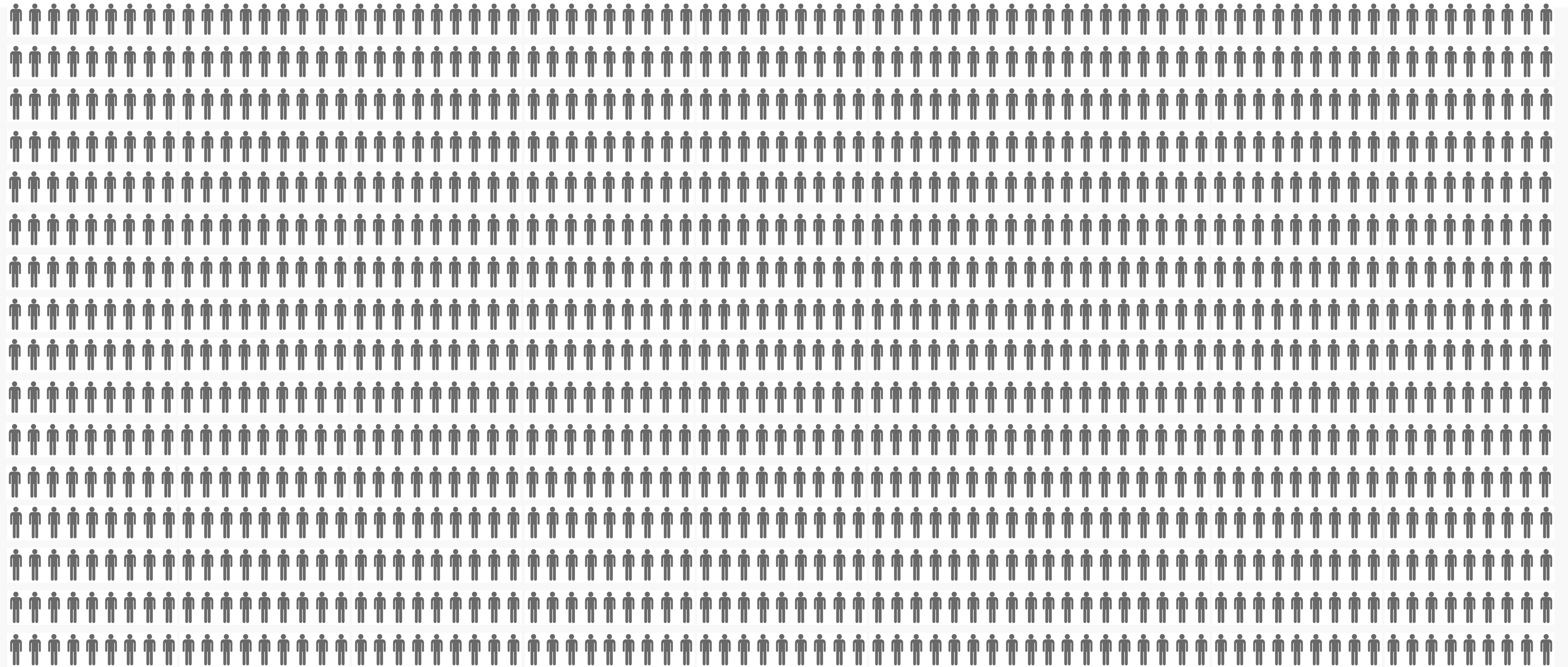
**HRAS**

**RB1**

**KRAS**

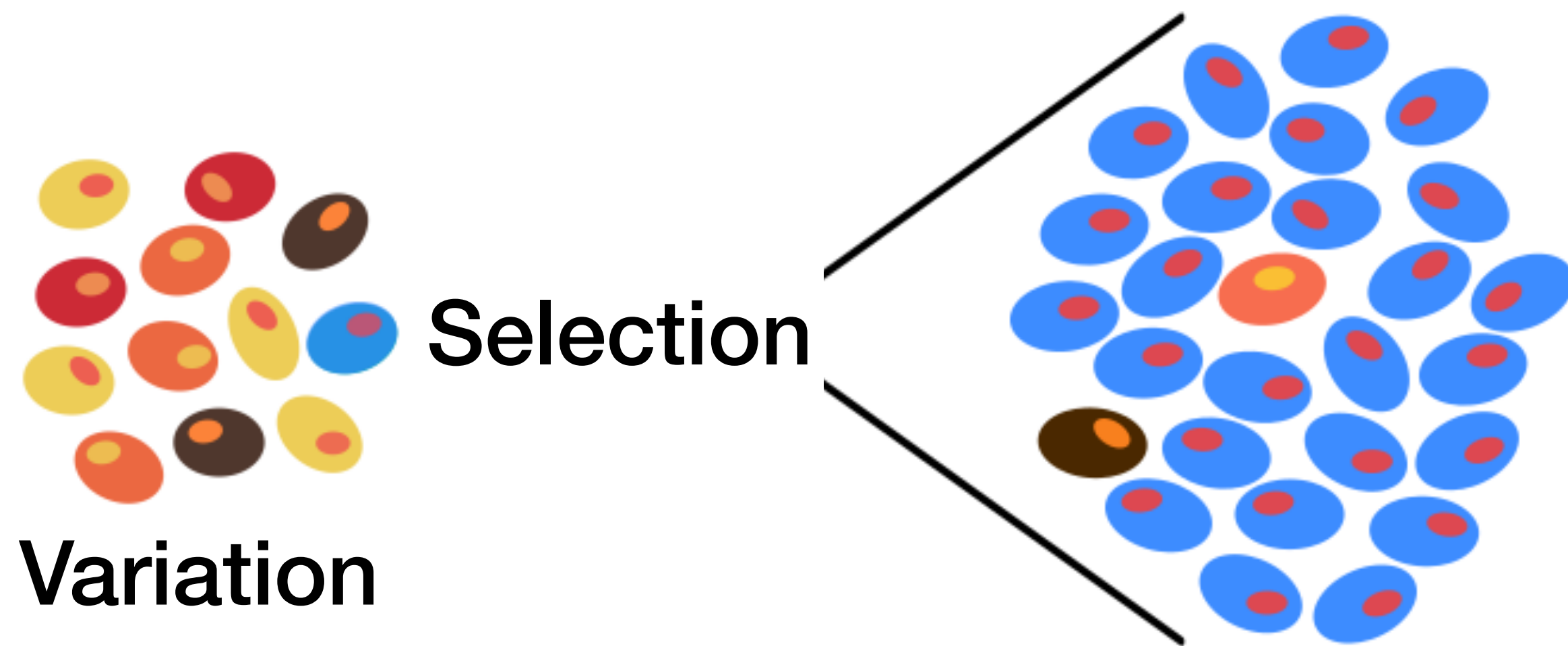


# Can we find cancer genes directly from tumors data?



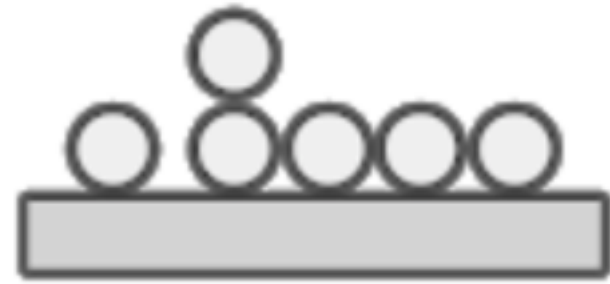
Thousands of tumor genomes sequenced

# Tumor development follows Darwinian Evolution



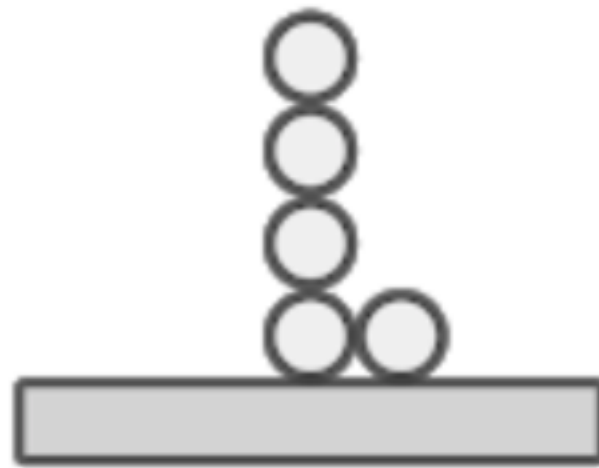
Drivers confer selective advantage to the cell

# Identifying signals of positive selection is an effective way to find cancer drivers



**R**

Identify genes mutated more frequently than the background mutation rate



**C**

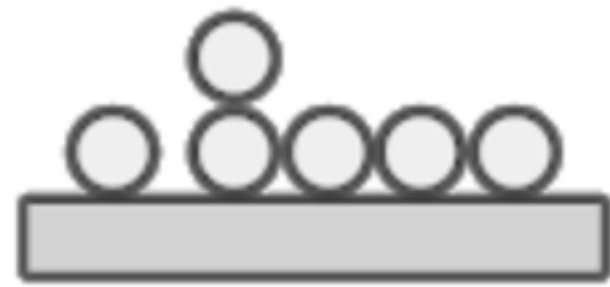
Identify genes with a significant regional clustering of mutations



**FI**

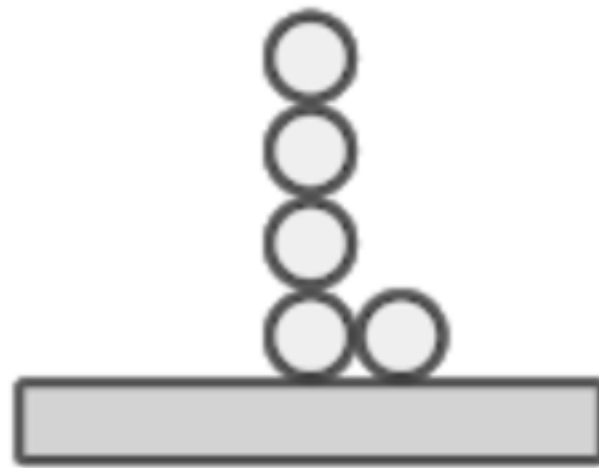
Identify genes with a bias towards high functional mutations (FM bias)

# Identifying signals of positive selection is an effective way to find cancer drivers



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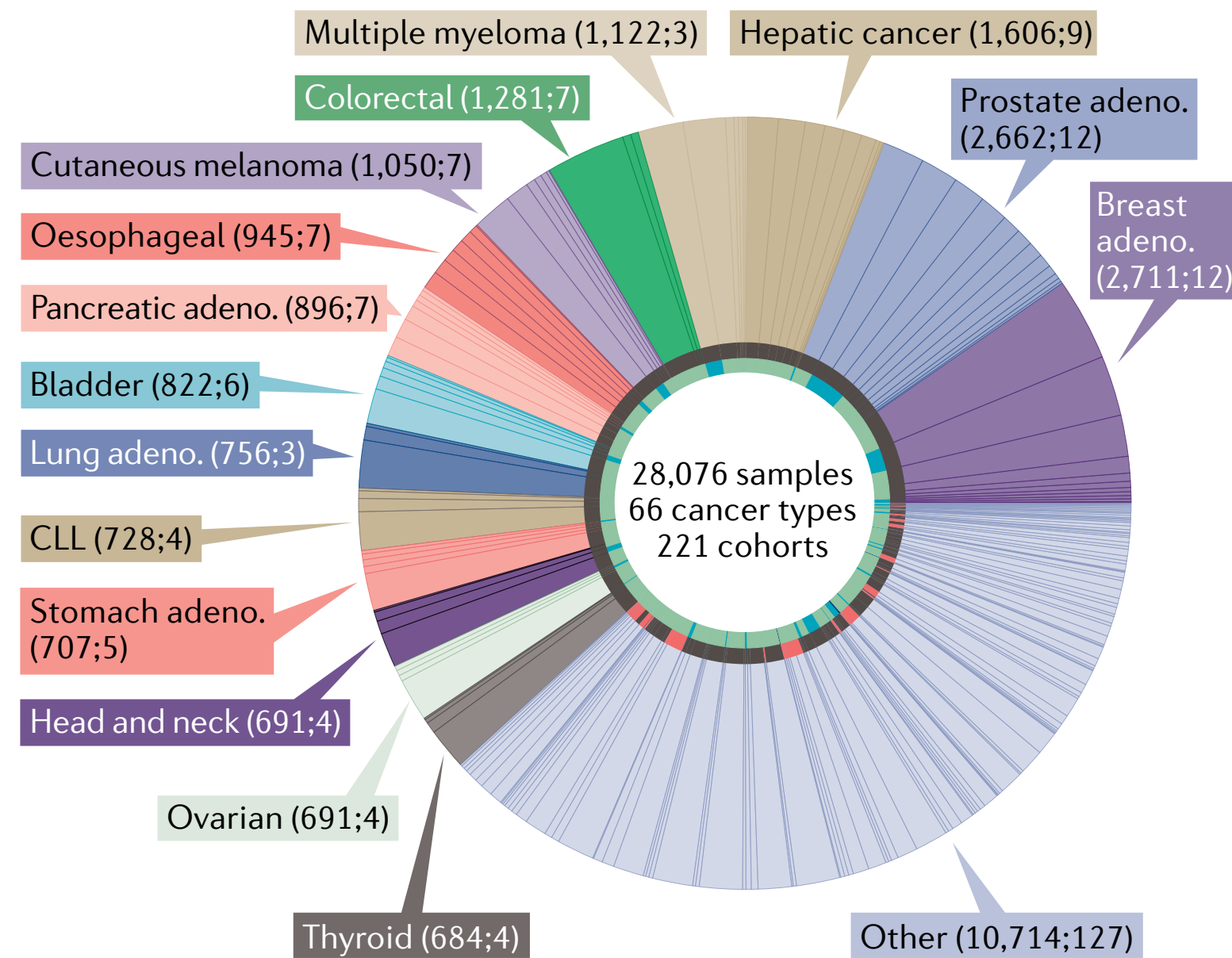


**FI**

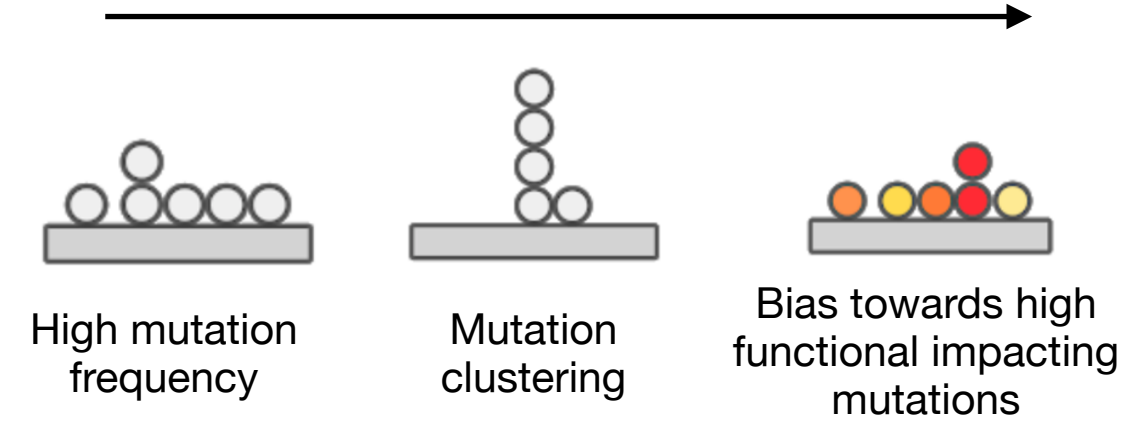
Identify genes with a bias towards high functional mutations (FM bias)



# The Compendium of Mutational Cancer Driver Genes



28,076 Tumors · 221 cohorts · 66 Cancer Types ·  
203,003,747 Mutations



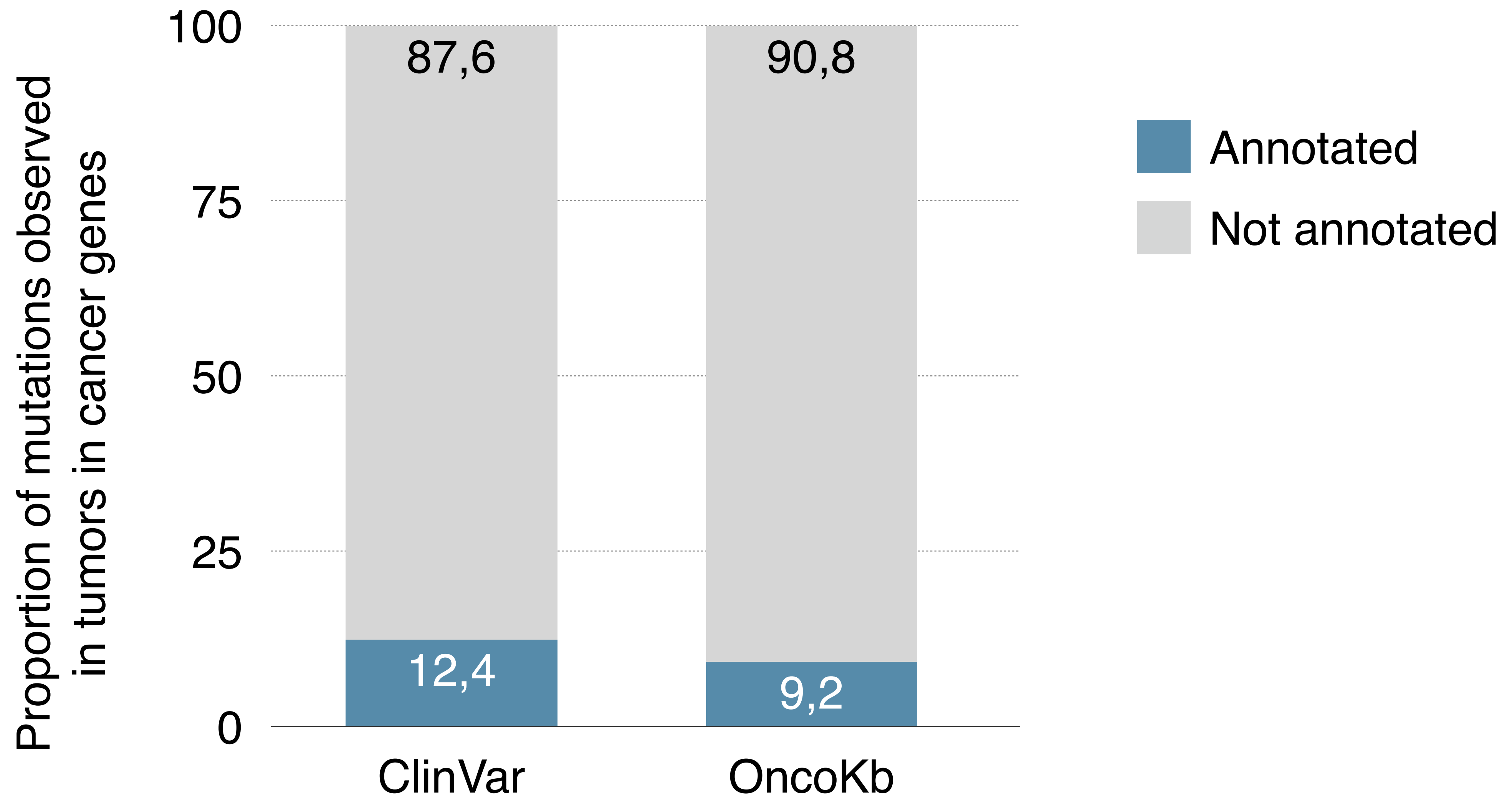
Mutation Pattern Analysis  
to identify Cancer Genes



This approach recovers most known cancer genes and also identify new ones

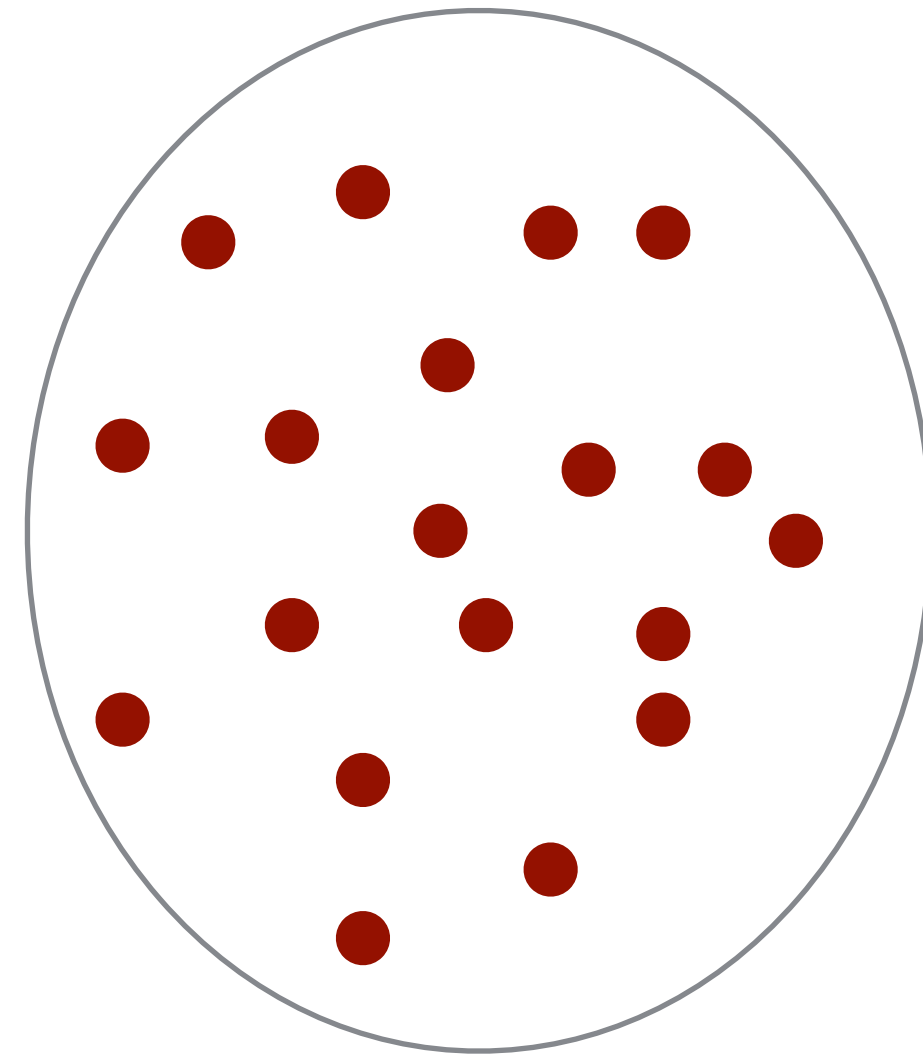


# Most mutations in cancer genes are of uncertain significance

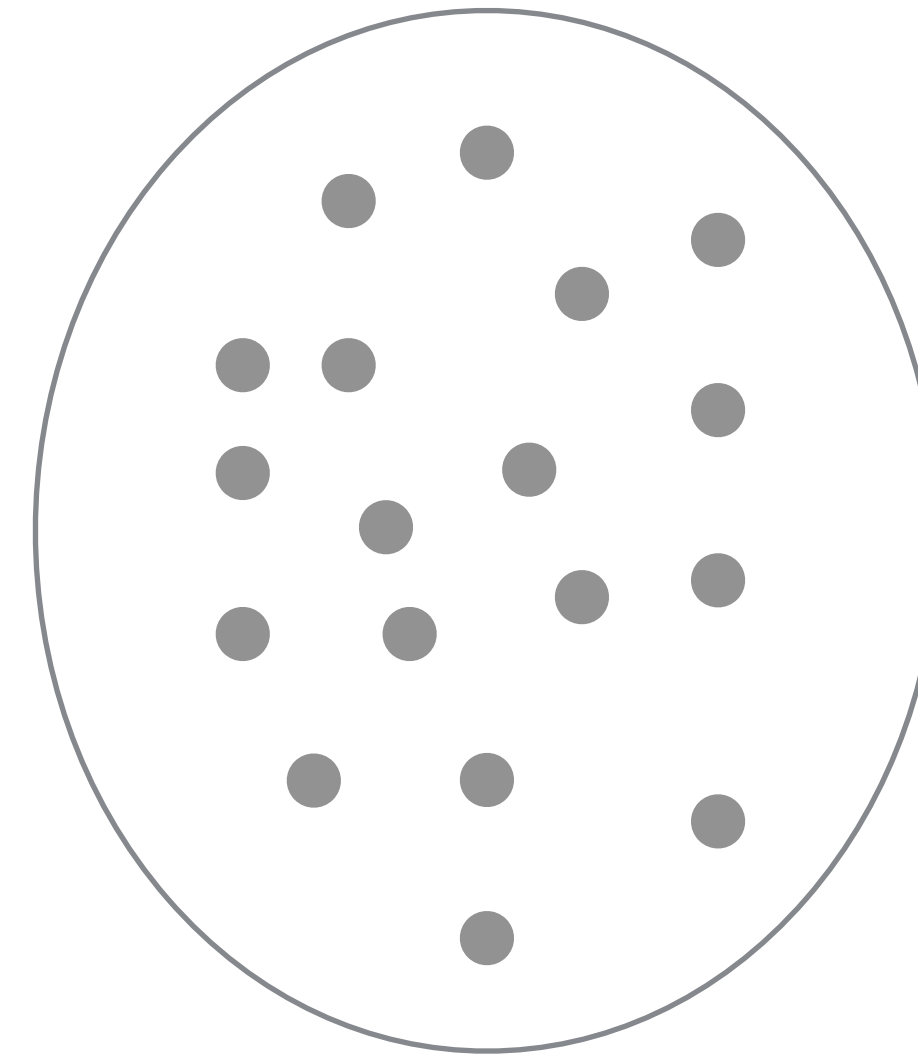


Which mutations in these cancer genes are capable of driving tumorigenesis?

# BoostDM: Learns feature combinations that define driver mutations in each gene/tumor type

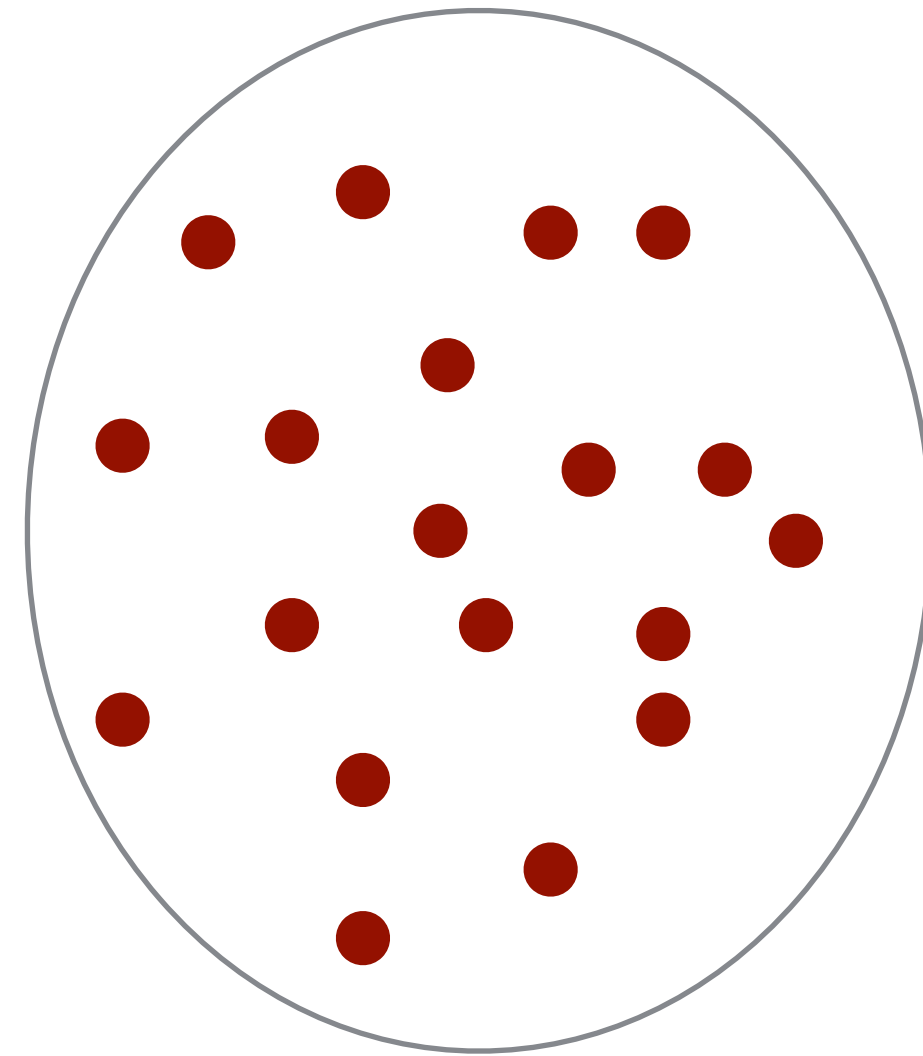


**Driver mutations**



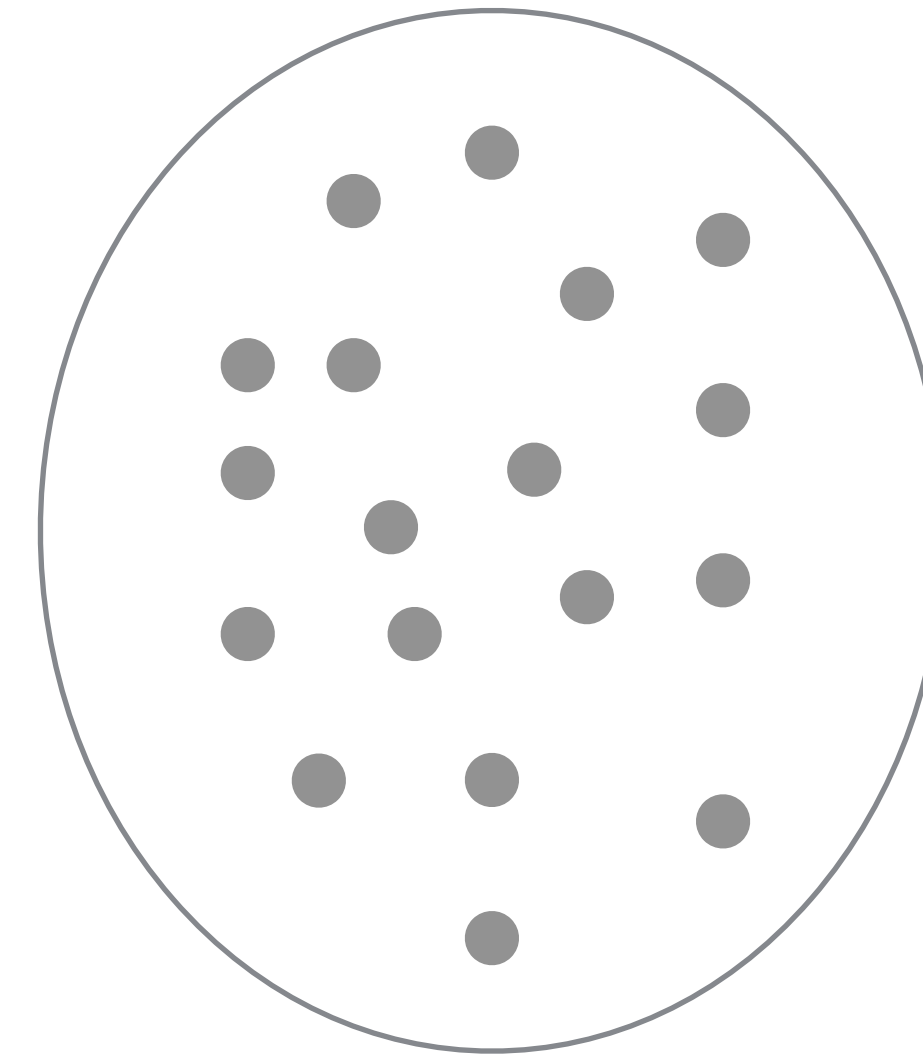
**Passenger mutations**

# BoostDM: Learns feature combinations that define driver mutations in each gene/tumor type



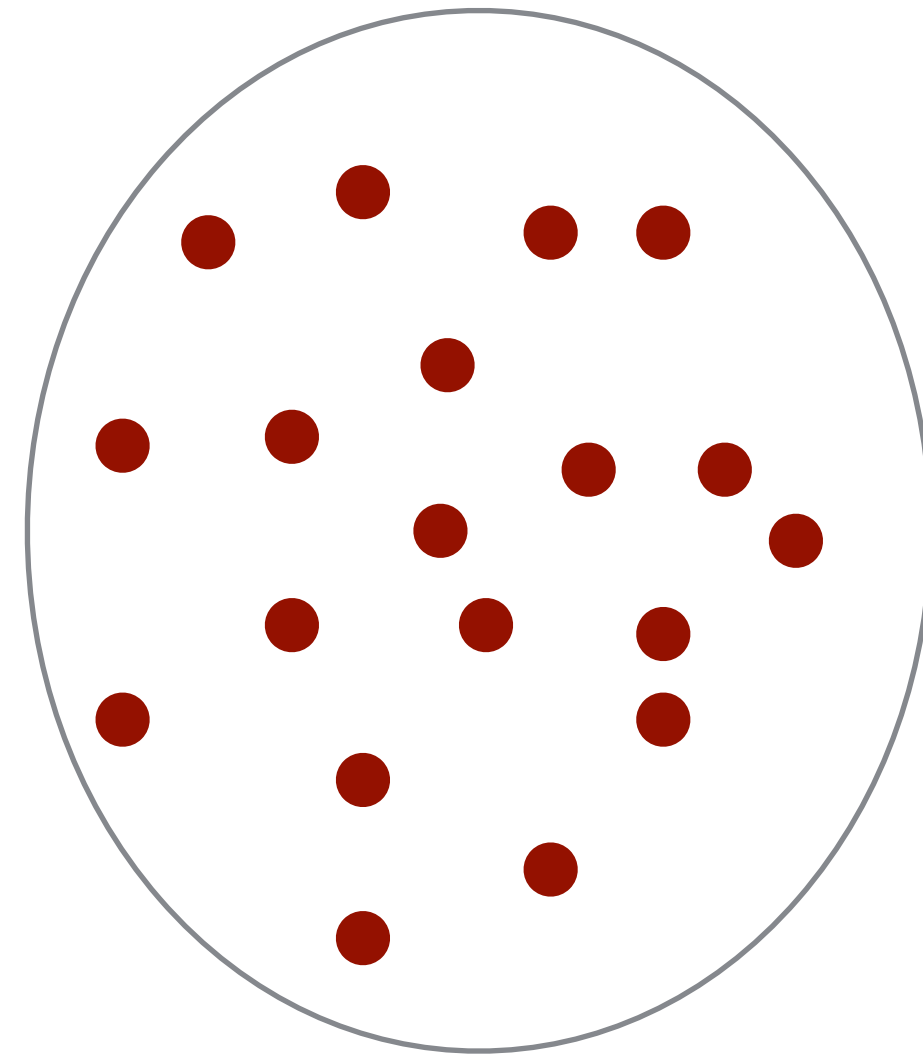
**Driver mutations**

Mutations observed in cancer genes in human tumors are enriched for drivers



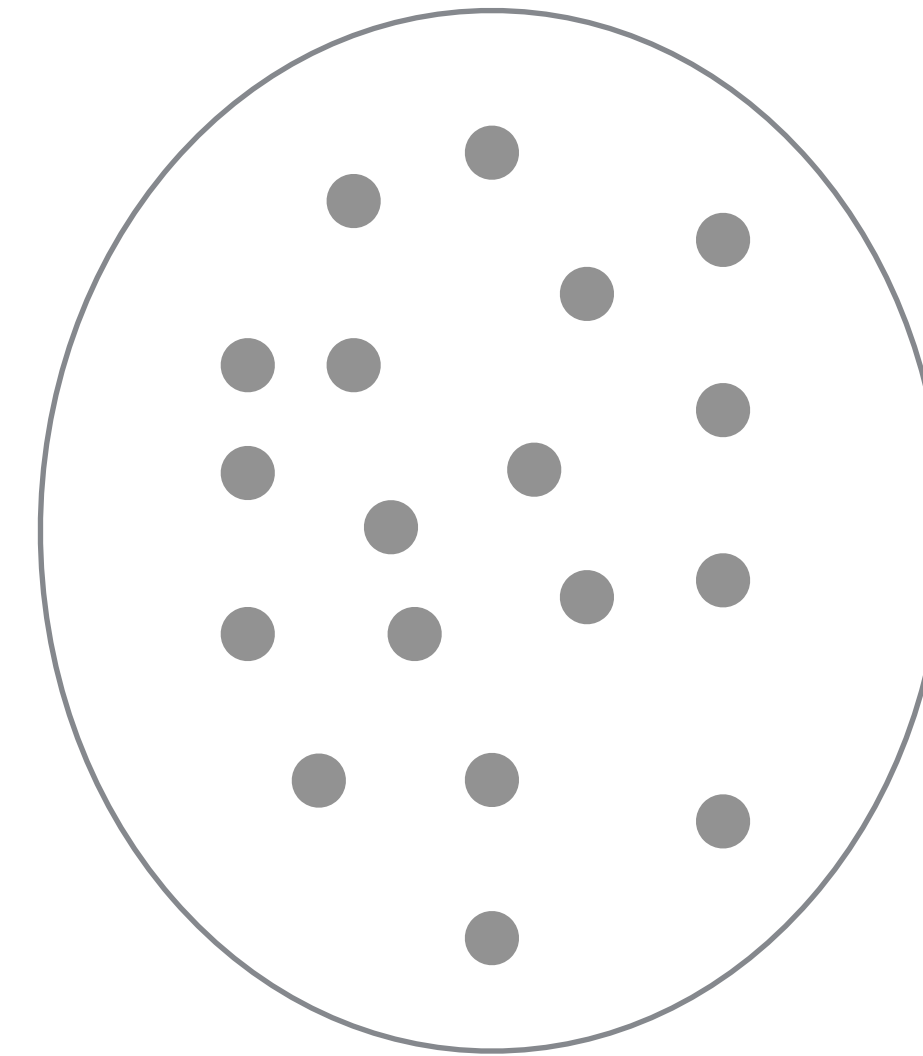
**Passenger mutations**

# BoostDM: Learns feature combinations that define driver mutations in each gene/tumor type



**Driver mutations**

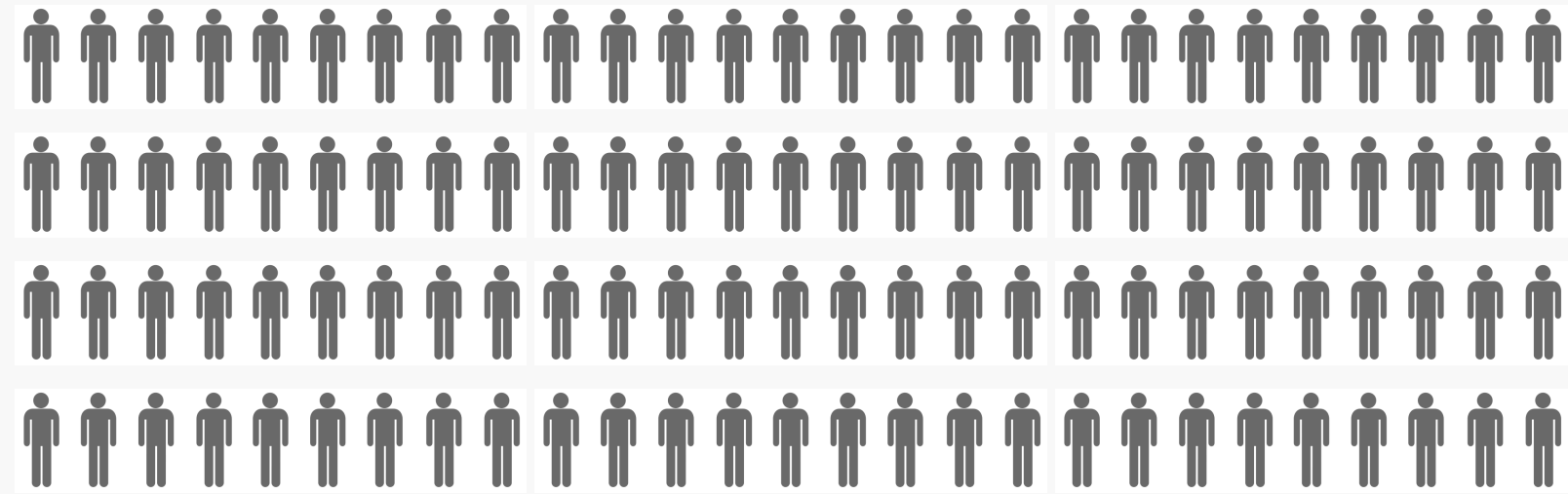
Mutations observed in cancer genes in human tumors are enriched for drivers



**Passenger mutations**

Simulate neutral mutagenesis to create mutations enriched for passengers

# BoostDM: Learns feature combinations that define driver mutations in each gene/tumor type



28,076 Tumors · 221 cohorts · 66 Cancer Types ·  
203,003,747 Somatic Mutations

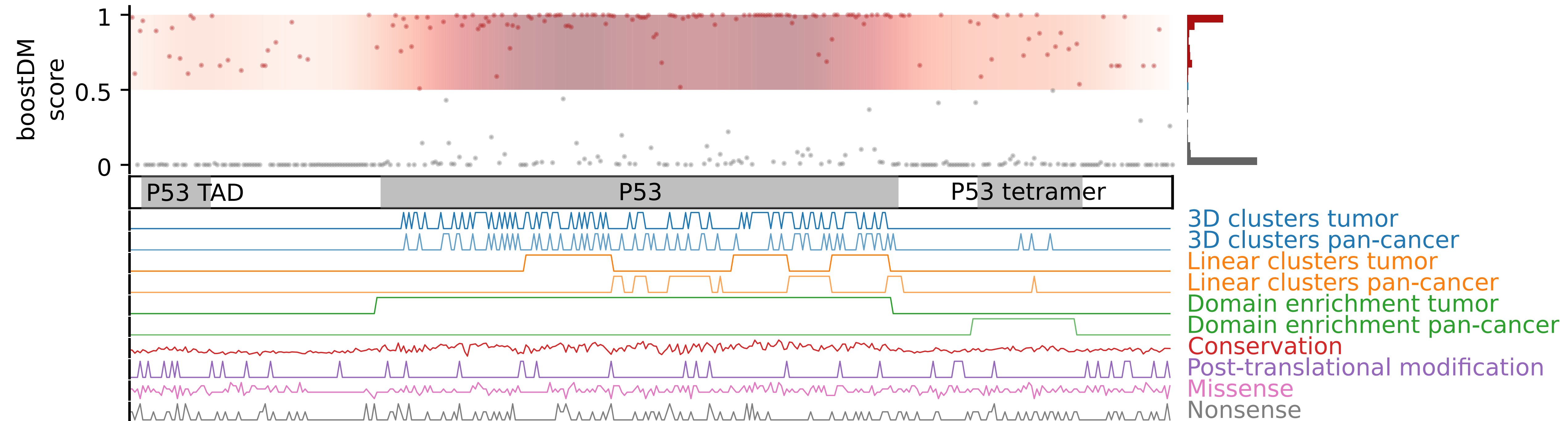
- Gene-tissue specific models
- Interpretable models

185 high quality models



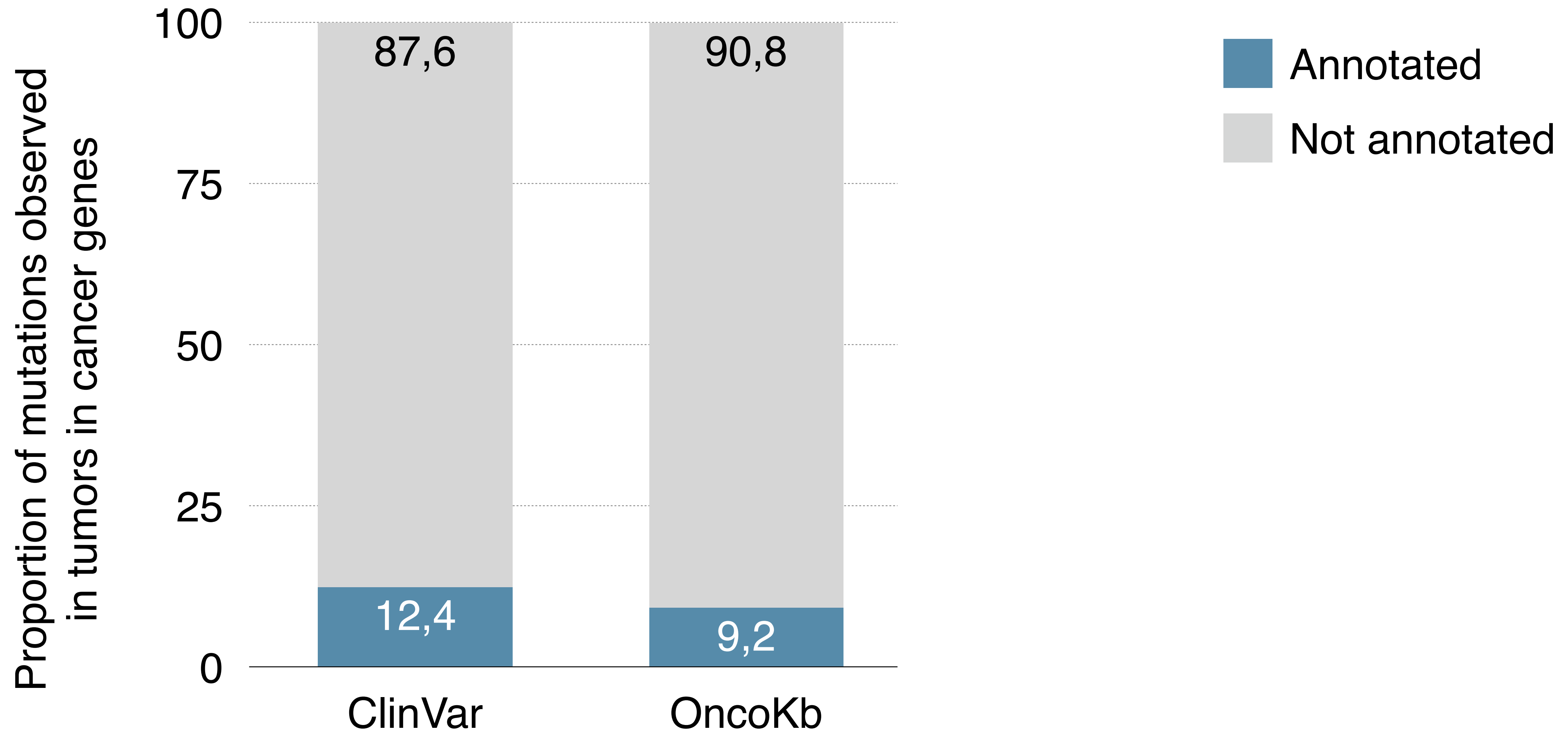
# In Silico Saturation Mutagenesis of Cancer Genes

## TP53 Colorectal Cancer



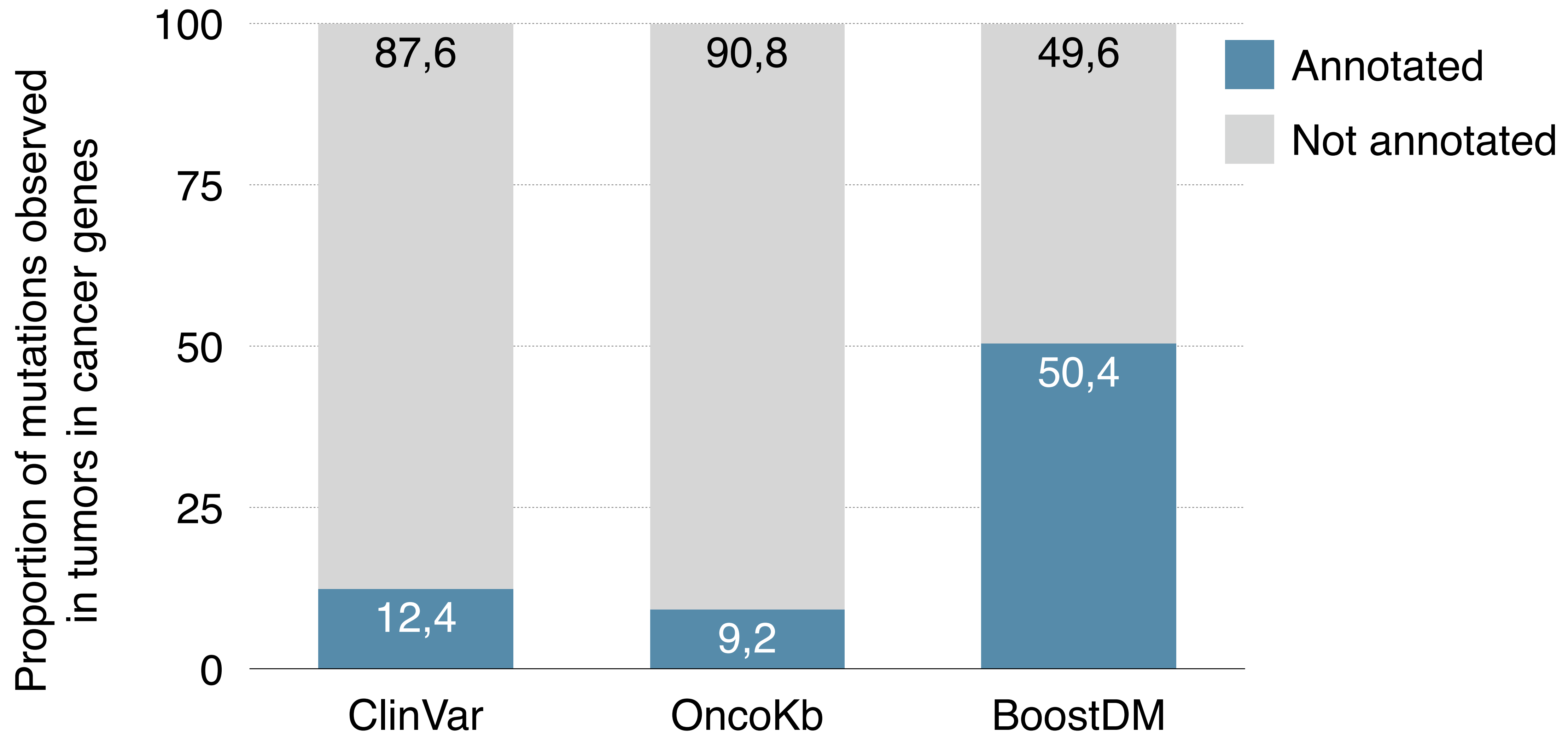
More blueprints at <http://intogen.org/boostdm>

# Most mutations in cancer genes are of uncertain significance





# Most mutations in cancer genes are of uncertain significance





# CANCER GENOME INTERPRETER

<http://www.cancergenomeinterpreter.org>



- ✓ Identifies potentially oncogenomic alterations
- ✓ Flags genomic biomarkers of drug response with different levels of clinical relevance

ALTERATIONS

PRESCRIPTIONS

## Mutations

Show entries with:  Mutations identified as **drivers**  Mutations with oncogenic **annotations**  **Other** mutations

Sample ID	Gene	Protein Change	Oncogenicity <span>?</span>	Mutation	Consequence	Oncog <span>≡</span>
Search here...						
TCGA-49-4494-01A-01D-	<u>EGFR</u>	T790M	<b>driver</b>	chr7:55249071 C>T	missense variant	
TCGA-49-4494-01A-01D-	<u>EGFR</u>	L858R	<b>driver</b>	chr7:55259515 T>G	missense variant	
TCGA-49-4494-01A-01D-	<u>MGA</u>	E2115*	<b>driver</b>	chr15:42042148 G>T	stop gained	
TCGA-49-4494-01A-01D-	<u>LRP1B</u>	R851P	<b>driver</b>	chr2:141751656 C>G	missense variant	
TCGA-49-4494-01A-01D-	<u>LRPPRC</u>	splice acceptor variant	<b>driver</b>	chr2:44204416 C>A	splice acceptor variant	
TCGA-49-4494-01A-01D-	<u>RBM10</u>	splice donor variant	<b>driver</b>	chrX:47034492 G>T	splice donor variant	
TCGA-49-4494-01A-01D-	<u>ARHGAP21</u>	E724*	<b>passenger</b>	chr10:24908654 C>A	stop gained	

# Clinical Implementation



**CGI-Clinics**  
Cancer Genome Interpreter

Data-driven cancer genome interpretation  
for personalized cancer treatment

<https://www.cgiclinics.eu/>

HORIZON-HLTH-2021-CARE-05  
5 years EU project, started November 2022

**CGI-Clinics** is an international multidisciplinary project with 17 partners involving biologists, bioinformaticians, oncologists, patients,...



4 EU Countries  
5 years  
10 M€

**Clinical partners and Healthcare payers**

9 Hospitals + 3 collaborators

30 hospitals in implementation phase (from 2026)

**Technical Development and Data Sharing**

**Health economics, Regulatory, Patient engagement**

**Patient associations and Policy makers**

**Communication and Dissemination**



- Identifying Cancer Drivers
- Understanding Neutral Mutagenesis
- Identifying Clonal Hematopoiesis Drivers
- Chemotherapy effect in hematopoiesis
- Cancer Promotion

# Modeling neutral mutagenesis

# Modeling neutral mutagenesis

- Variable mutation rate along the genome

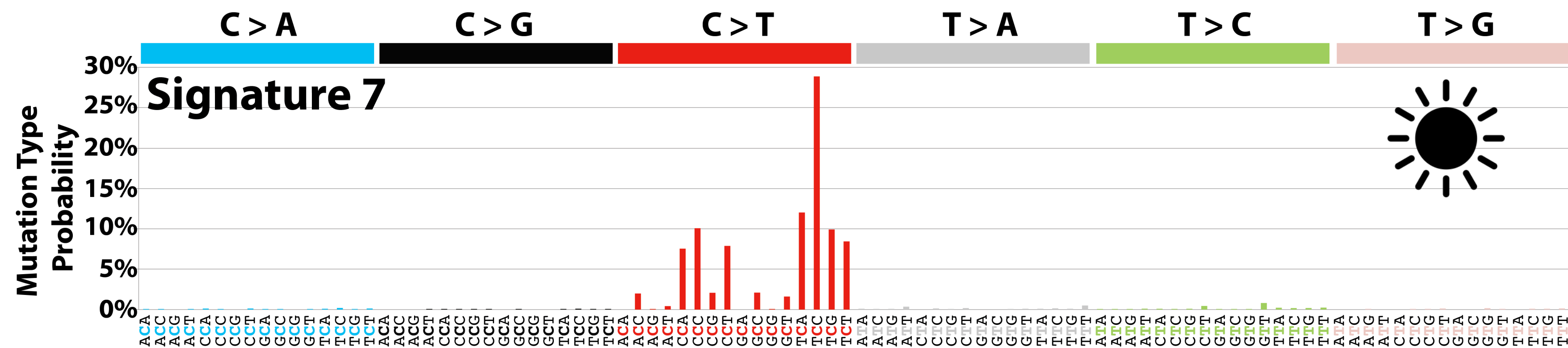


# Modeling neutral mutagenesis

- Variable mutation rate along the genome



- Different probability for different sequence context (mutational signatures)





# Mutational signatures of cancer treatments

## DNA damaging agent

Chemotherapy



e.g. alkylating agents



# Mutational signatures of cancer treatments

**DNA damaging agent**

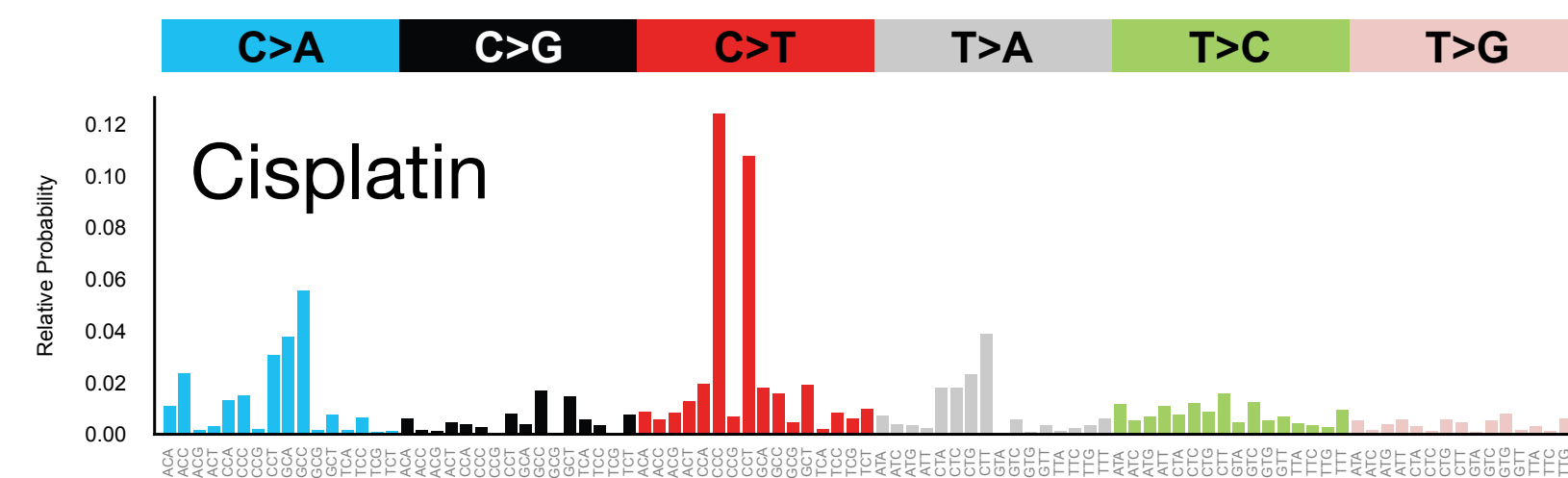
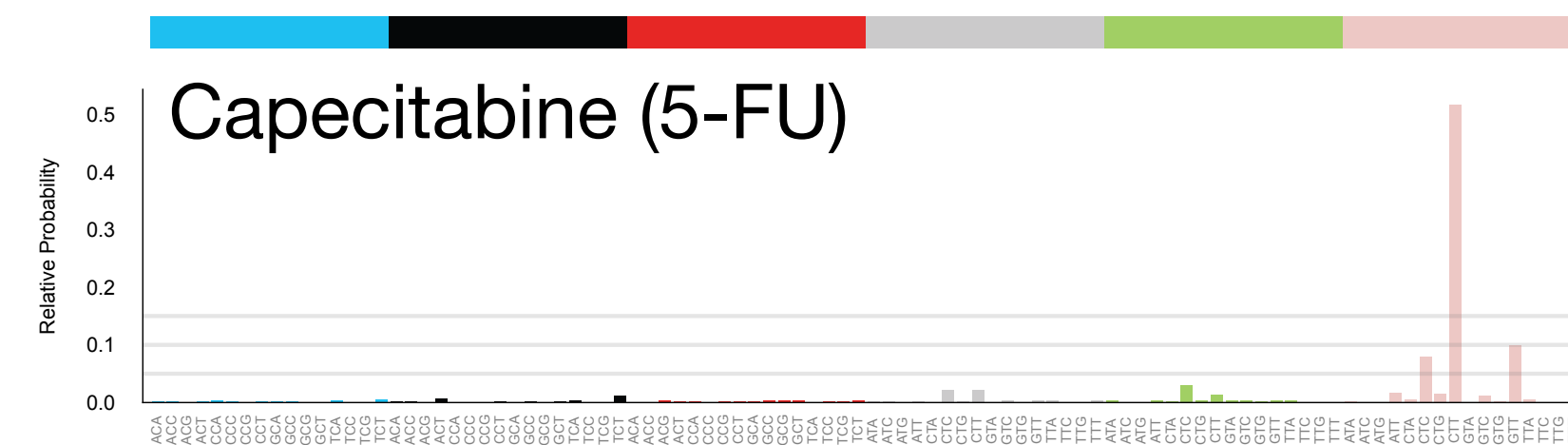
Chemotherapy



e.g. alkylating agents



## Mutational signatures of cancer treatments



Pich et al., Nature Genetics 2019



Oriol Pich



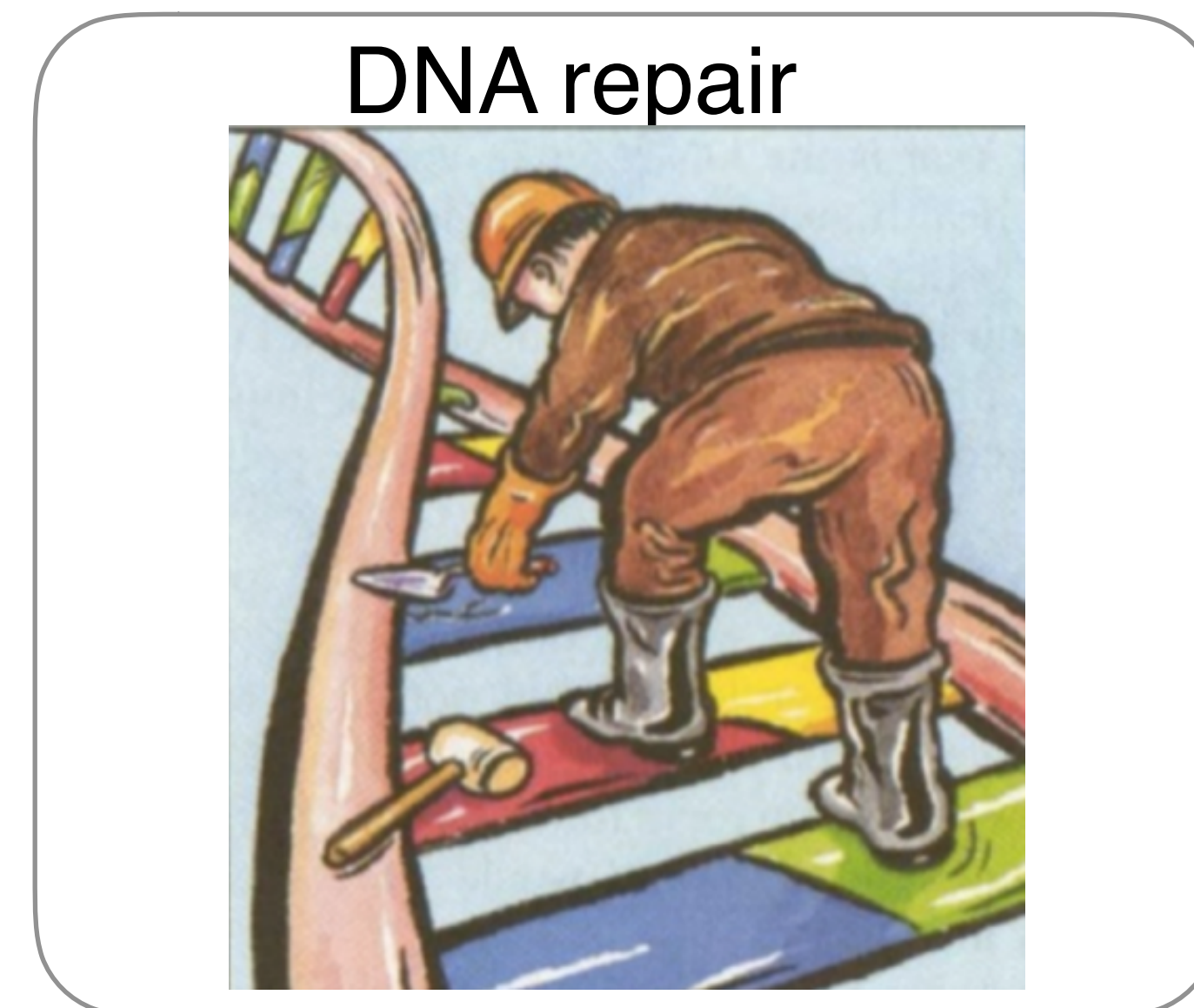
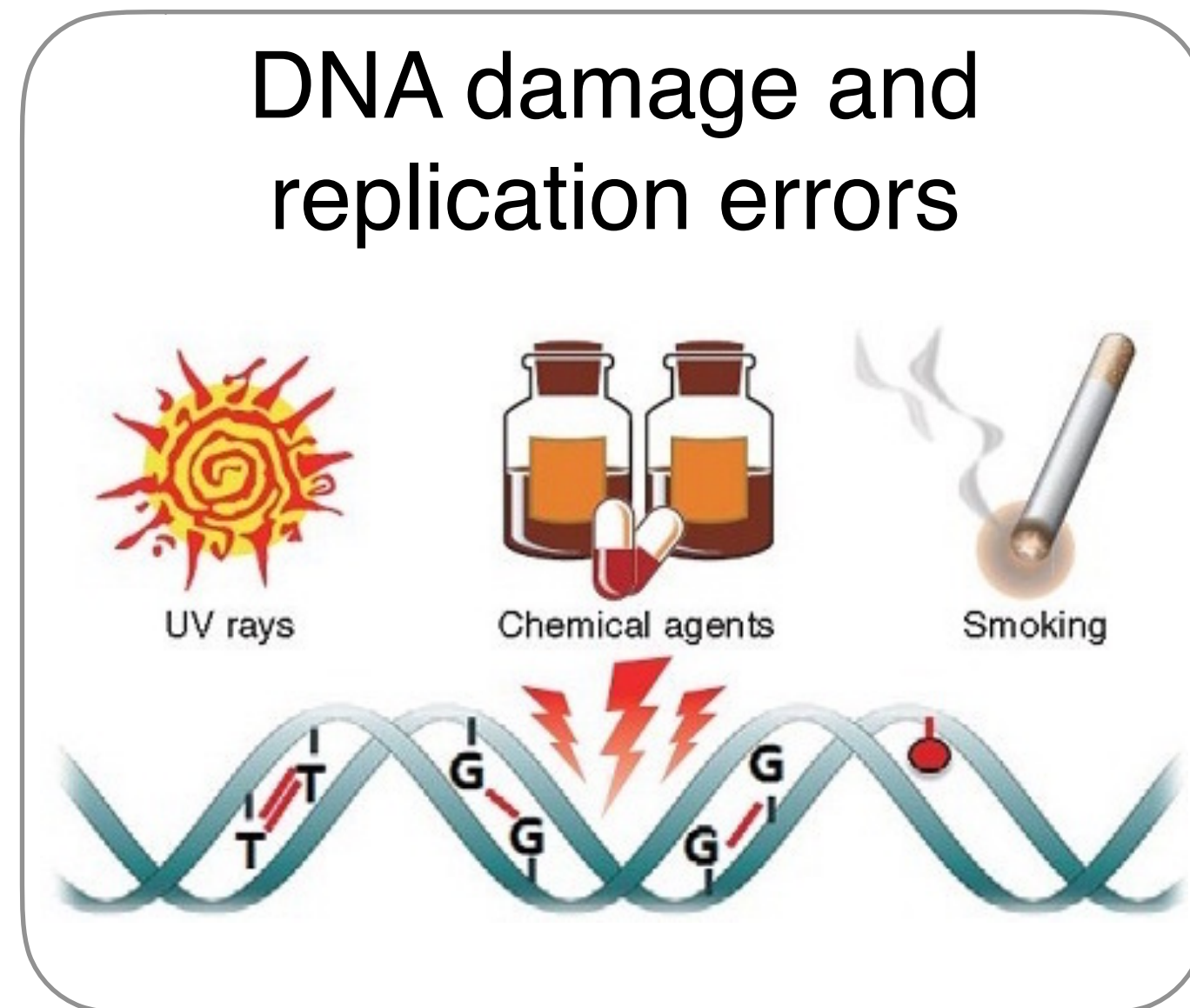
Abel Gonzalez-Perez



Ferran Muiños

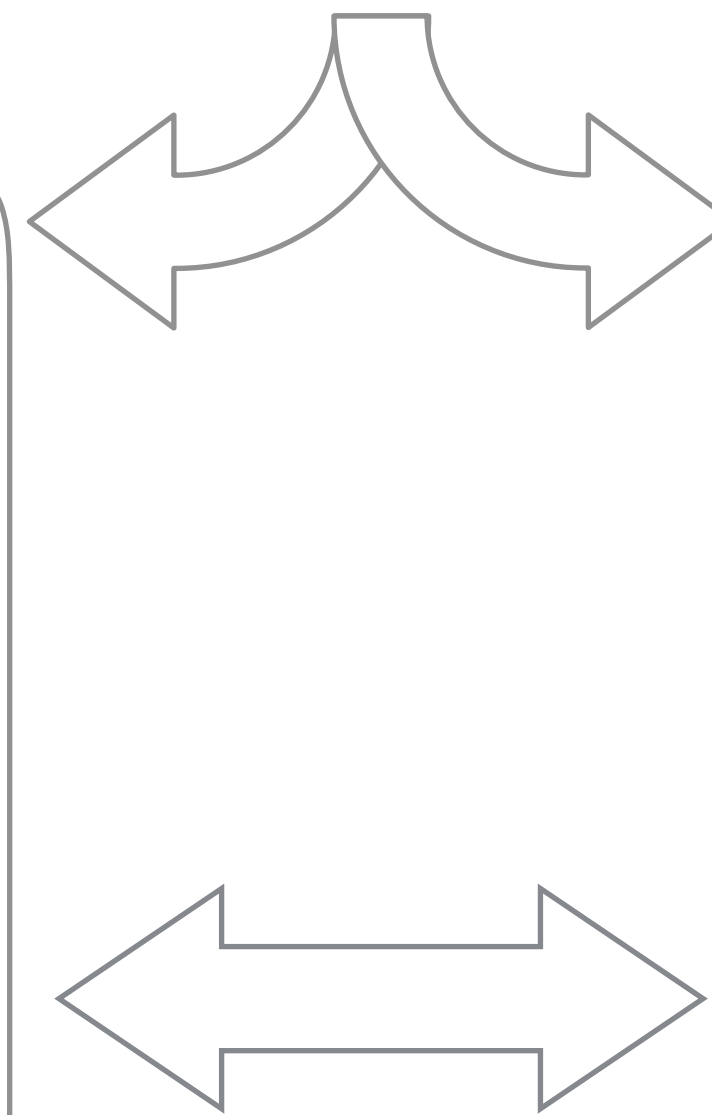
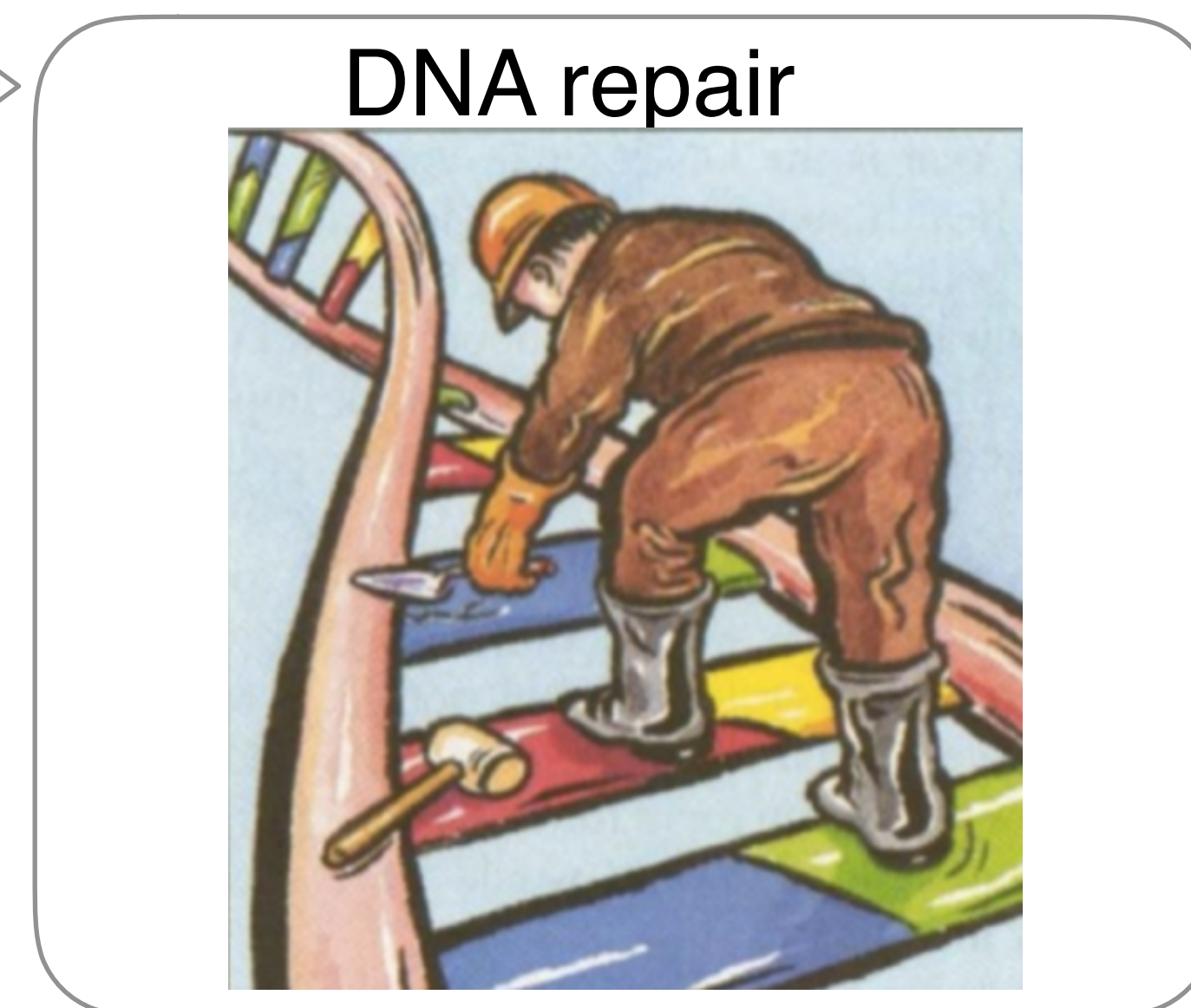
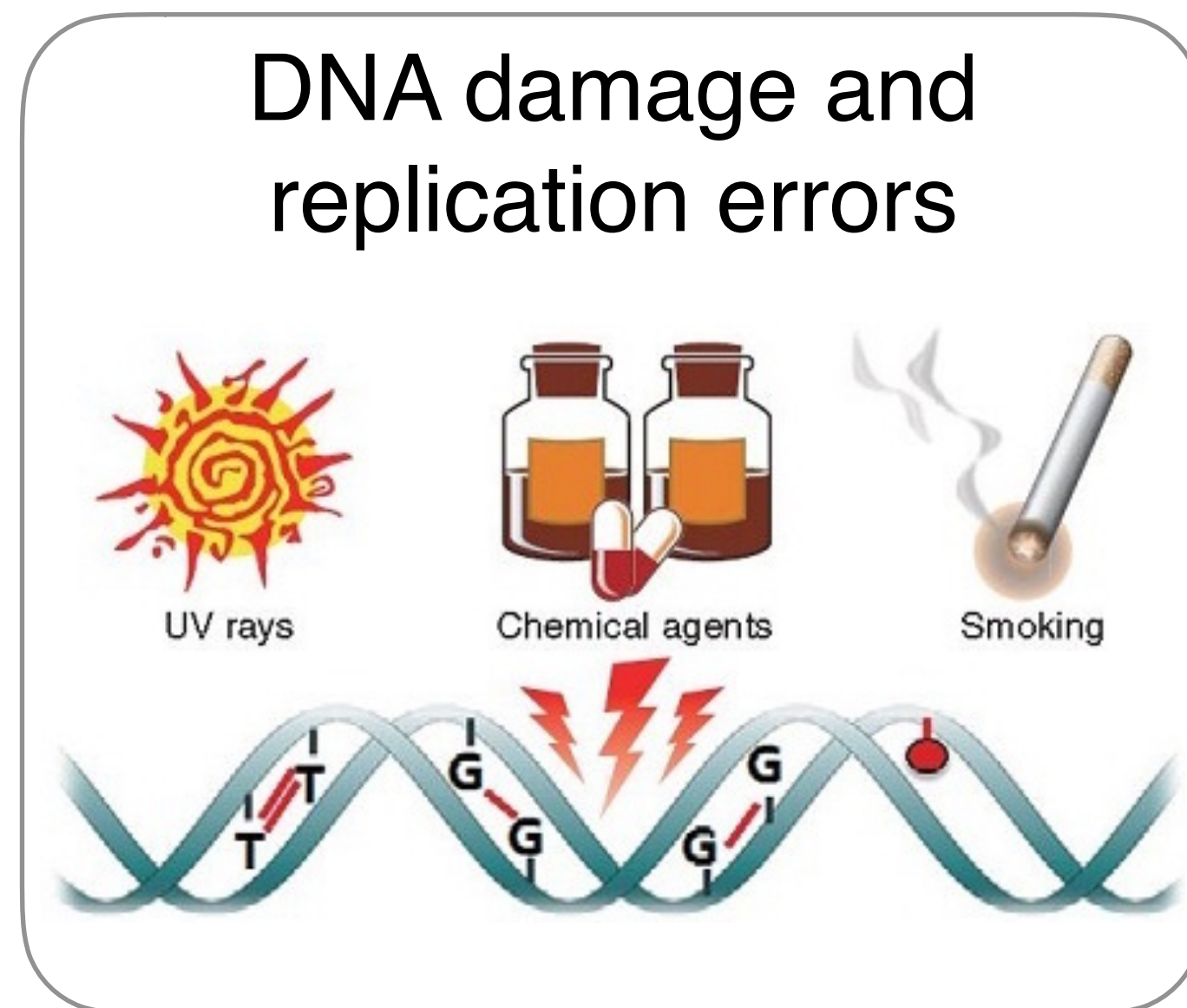
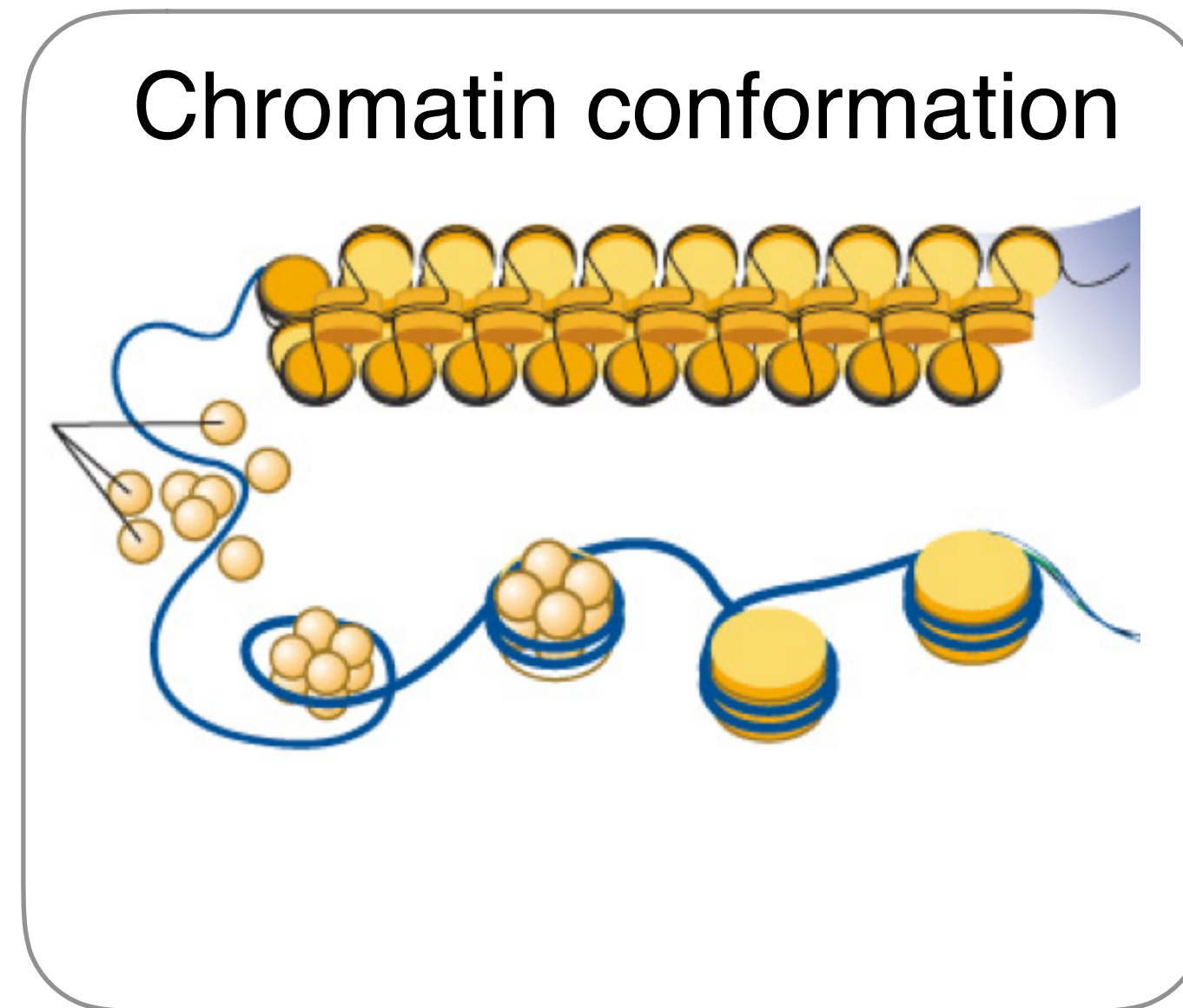


# Interplay between DNA damage and DNA repair

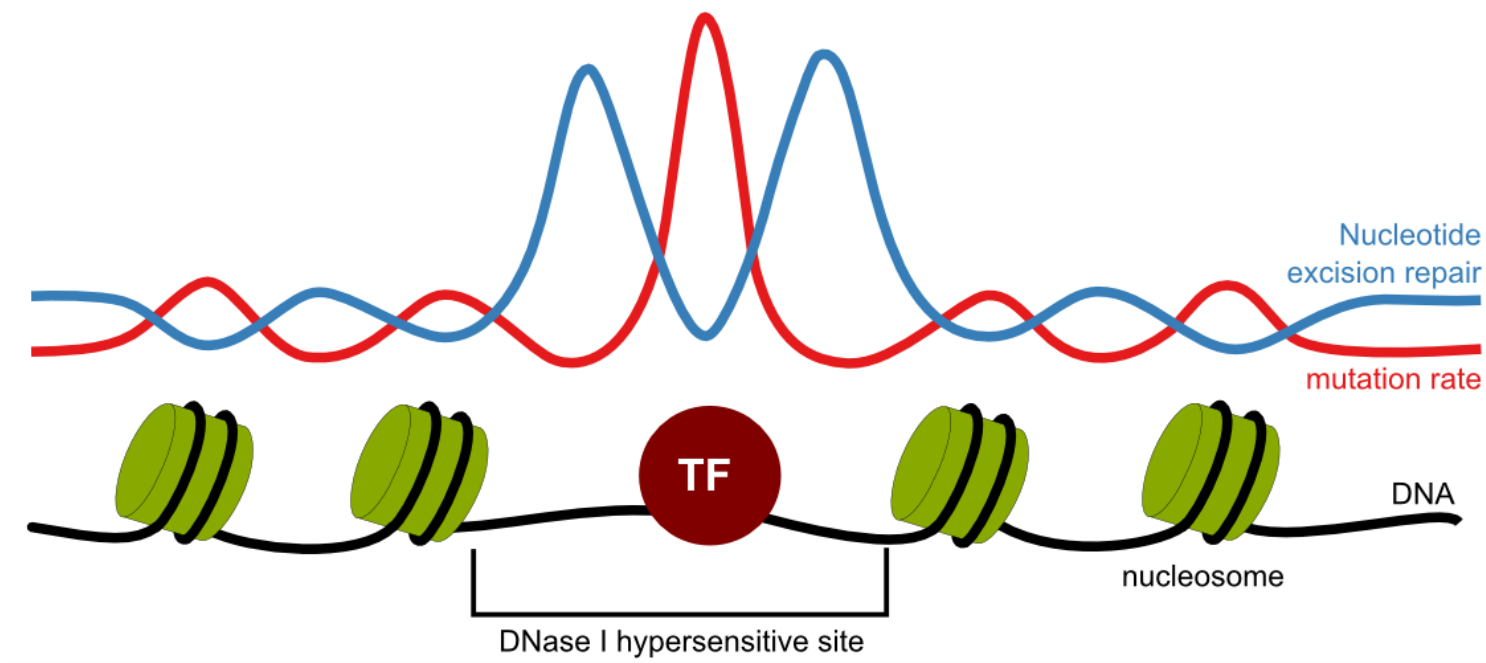




# Interplay between DNA damage and DNA repair and chromatin

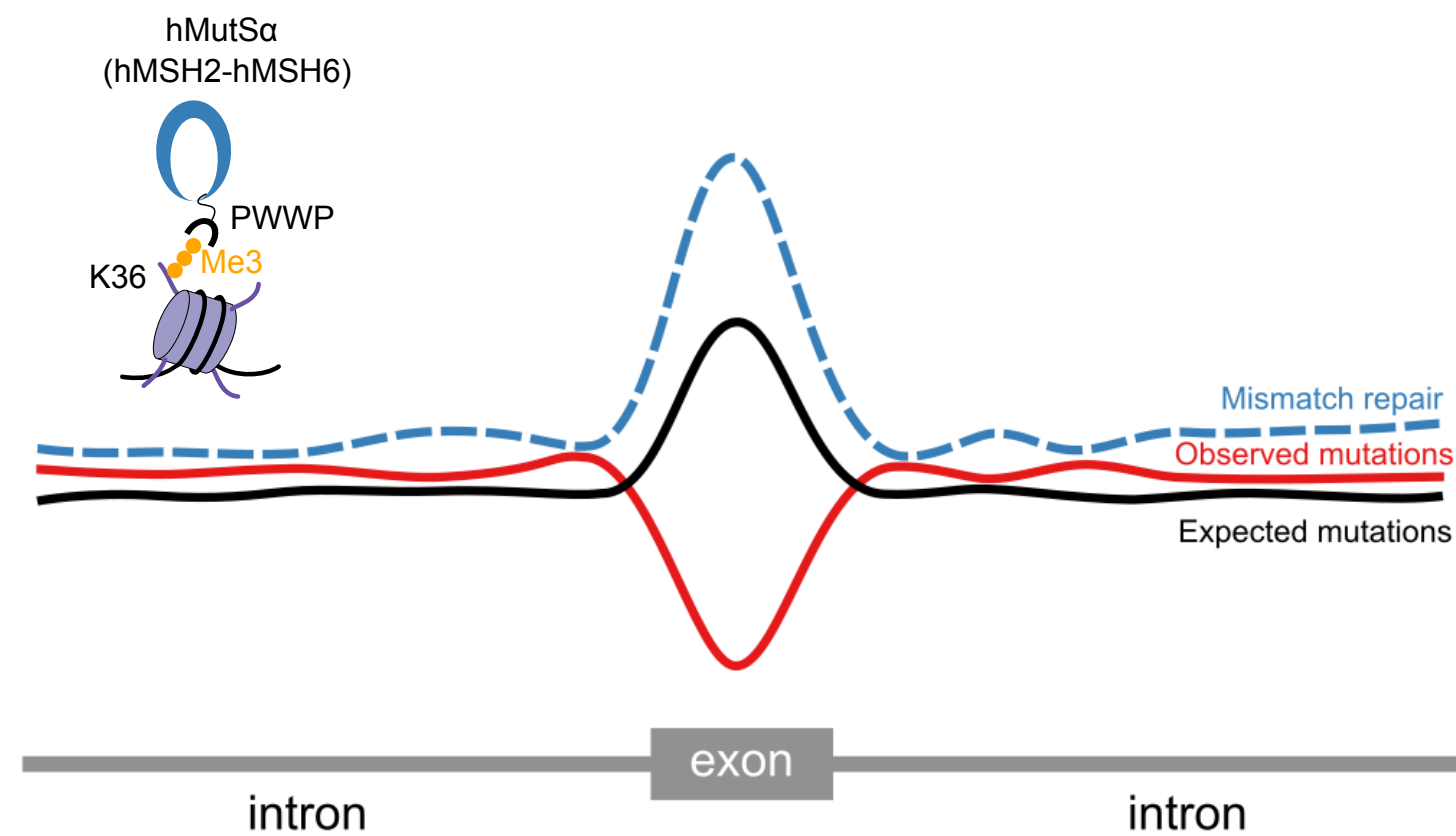


# Mutation rate variability at local scale



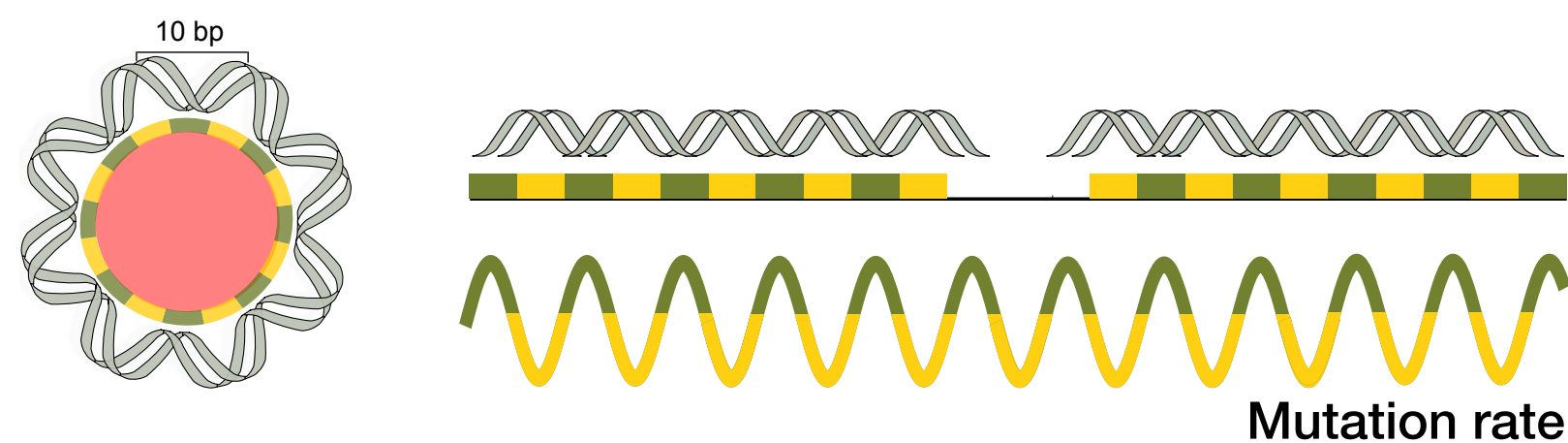
High mutation rate in TFBS due to impaired Nucleotide Excision Repair

Sabarinathan et al., Nature 2016



Differential mismatch repair leads to reduced mutation rate in exons

Frigola et al., Nature Genetics 2017



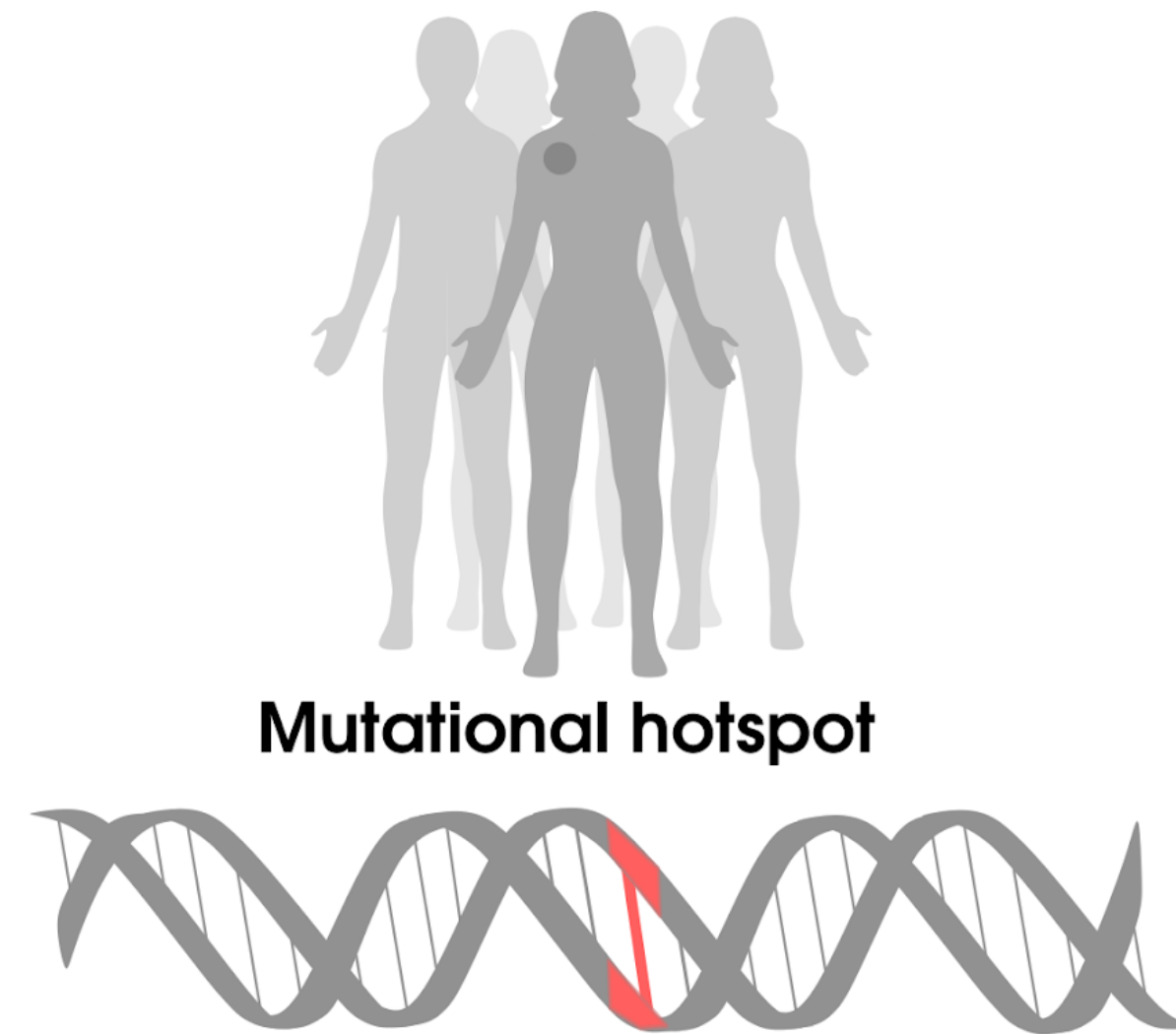
10-bp mutation rate periodicity in nucleosome covered DNA

Pich et al., Cell 2018





# How well do we estimate mutation rate at single-nucleotide resolution?



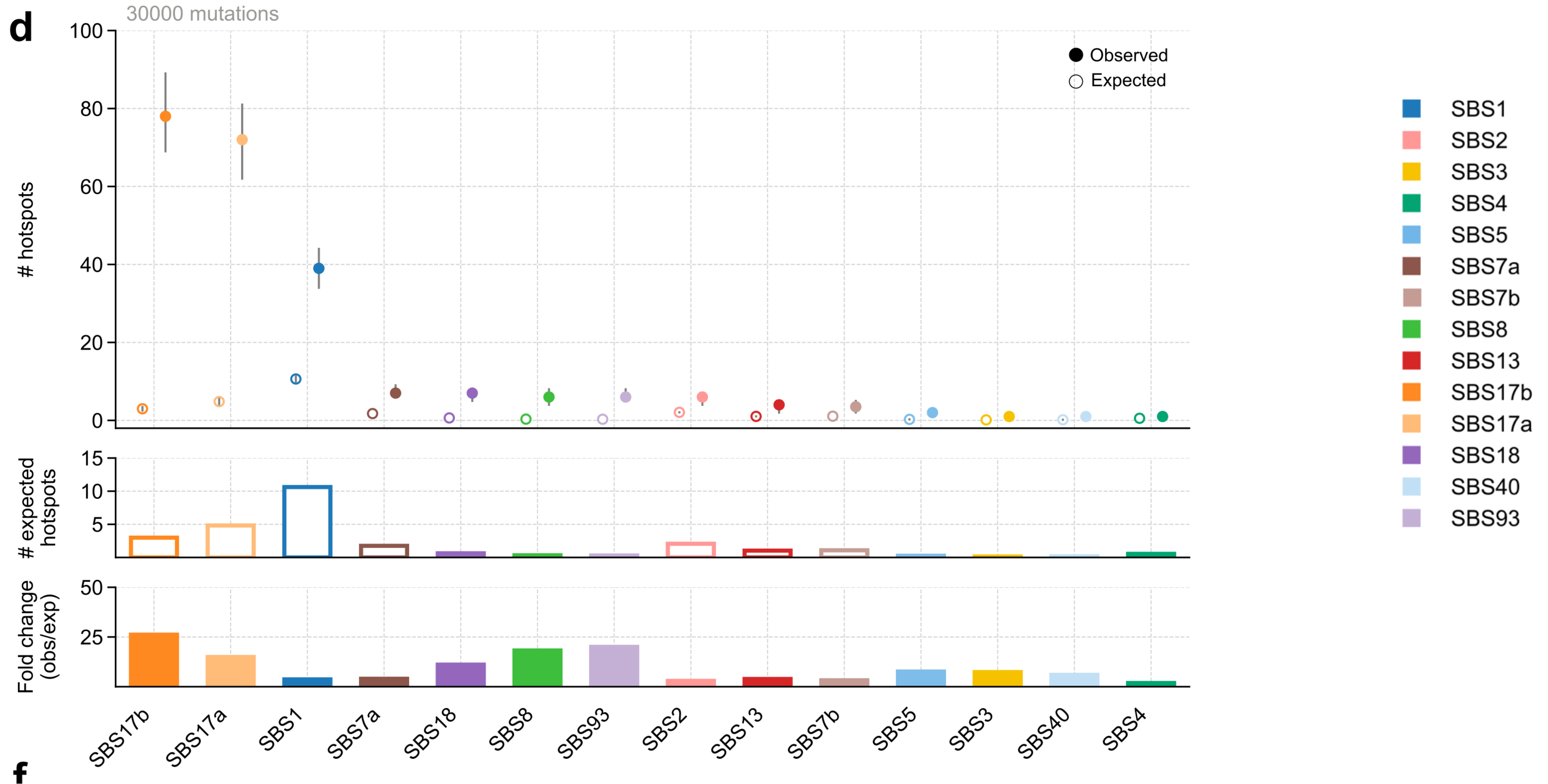
Identification of somatic mutation hotspots across 7,507 whole genomes

Which processes create hotspots? Can we predict those with current models?



Claudia Arnedo

# SBS1 and SBS17a, b have the highest propensity to hotspot formation

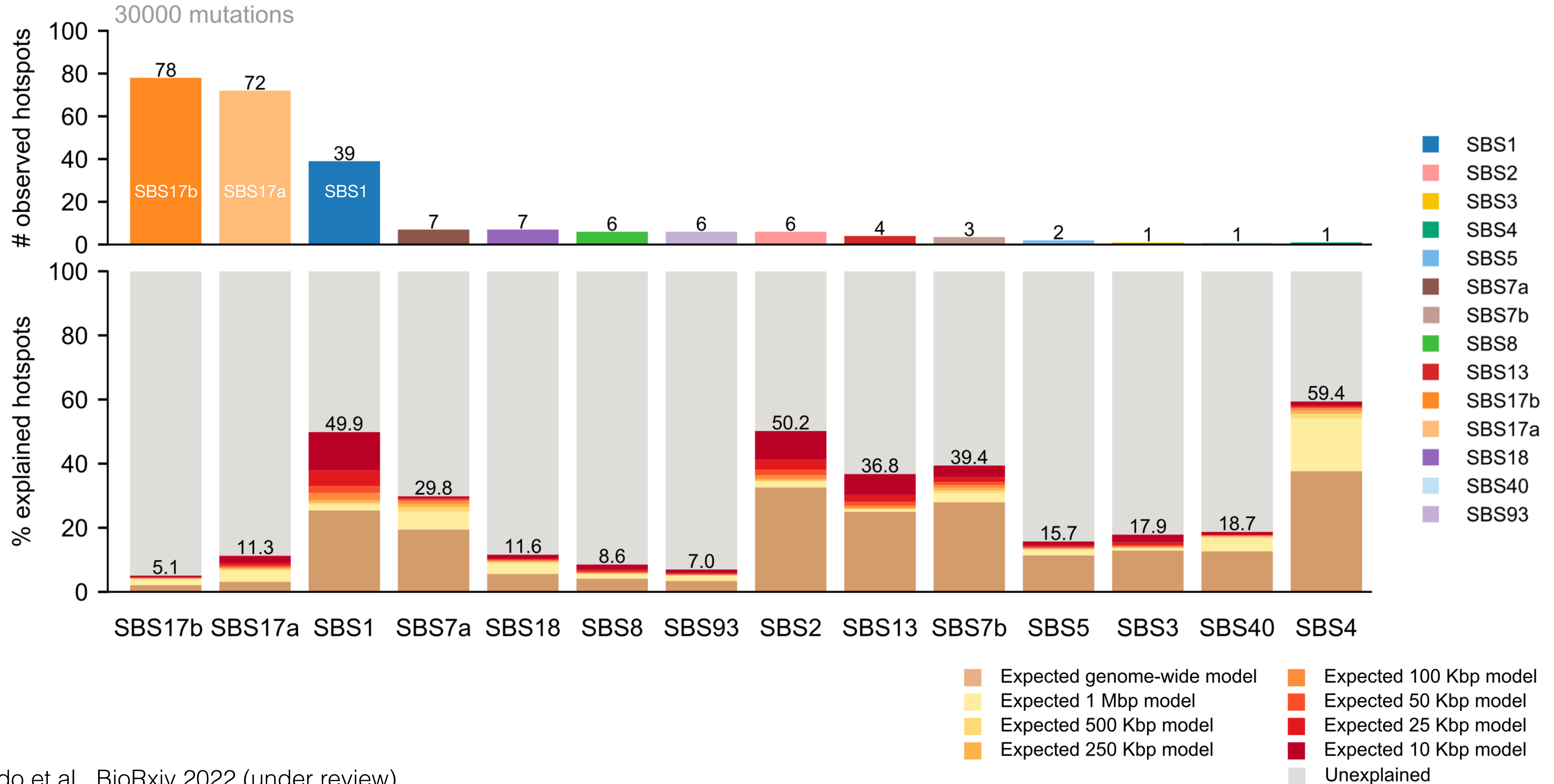


**f**

Hotspot = 2 or more samples with the same mutation

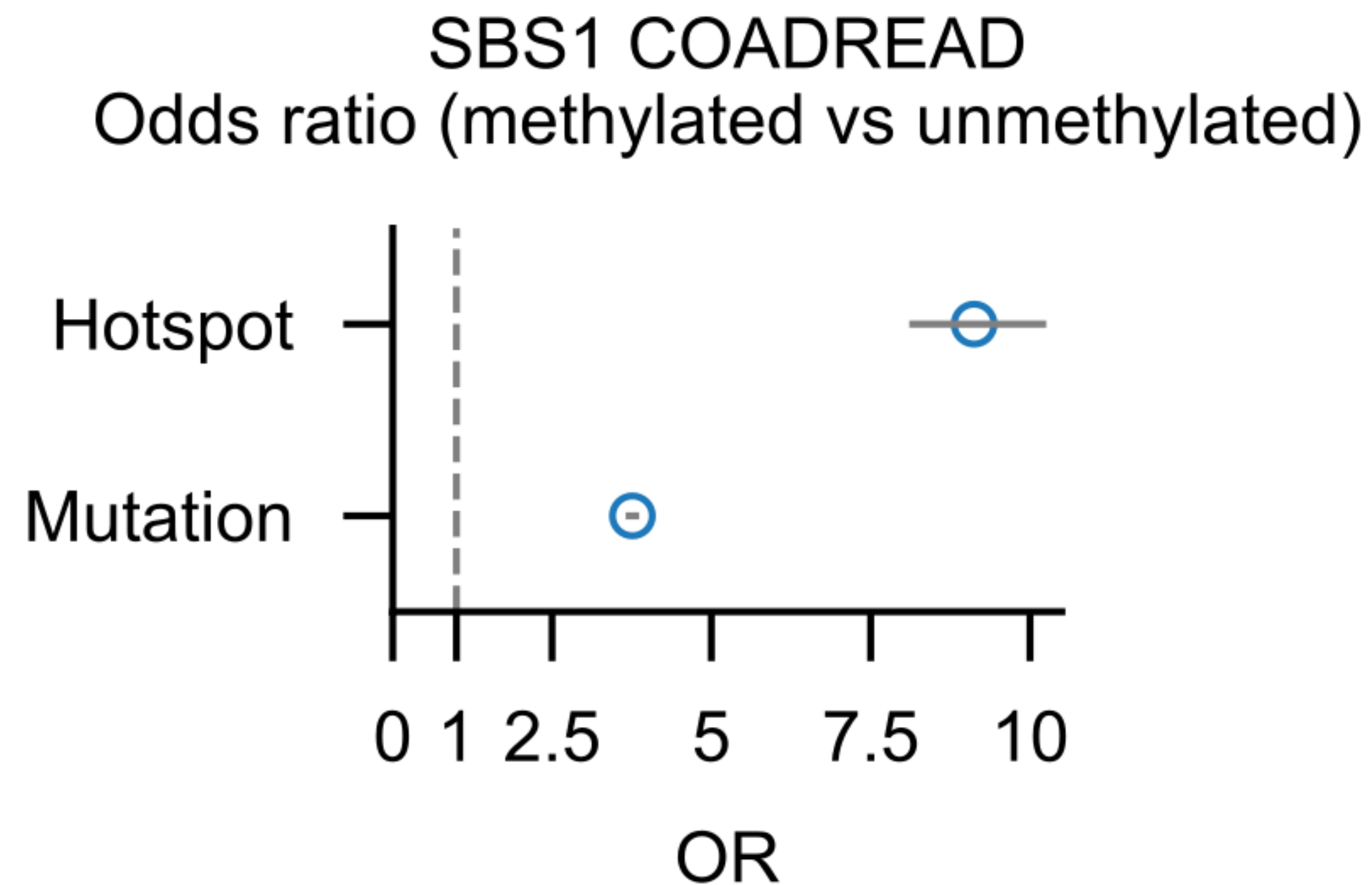
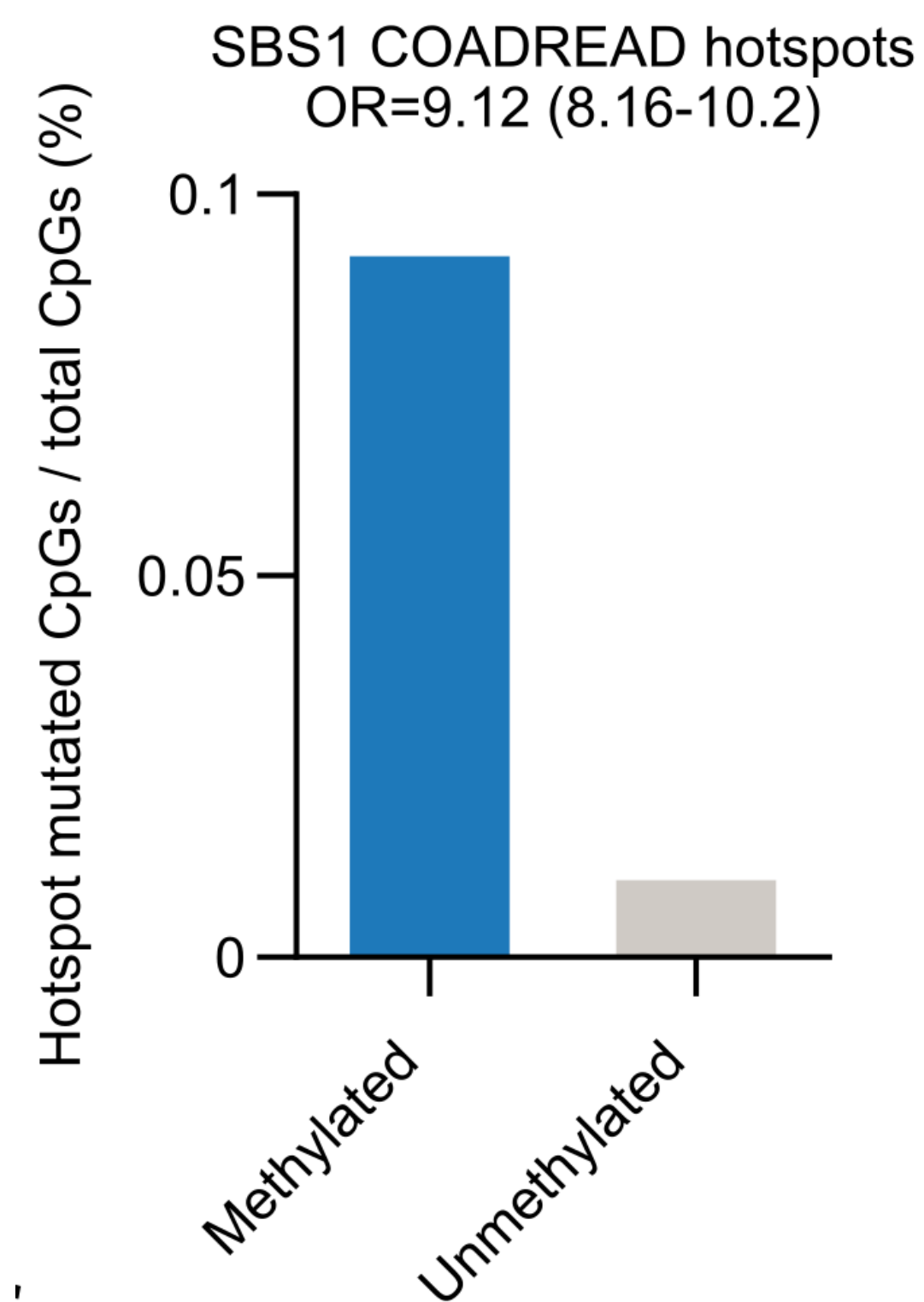
Arnedo et al., BioRxiv 2022 (under review)

# Large proportion of hotspots remain unexplained

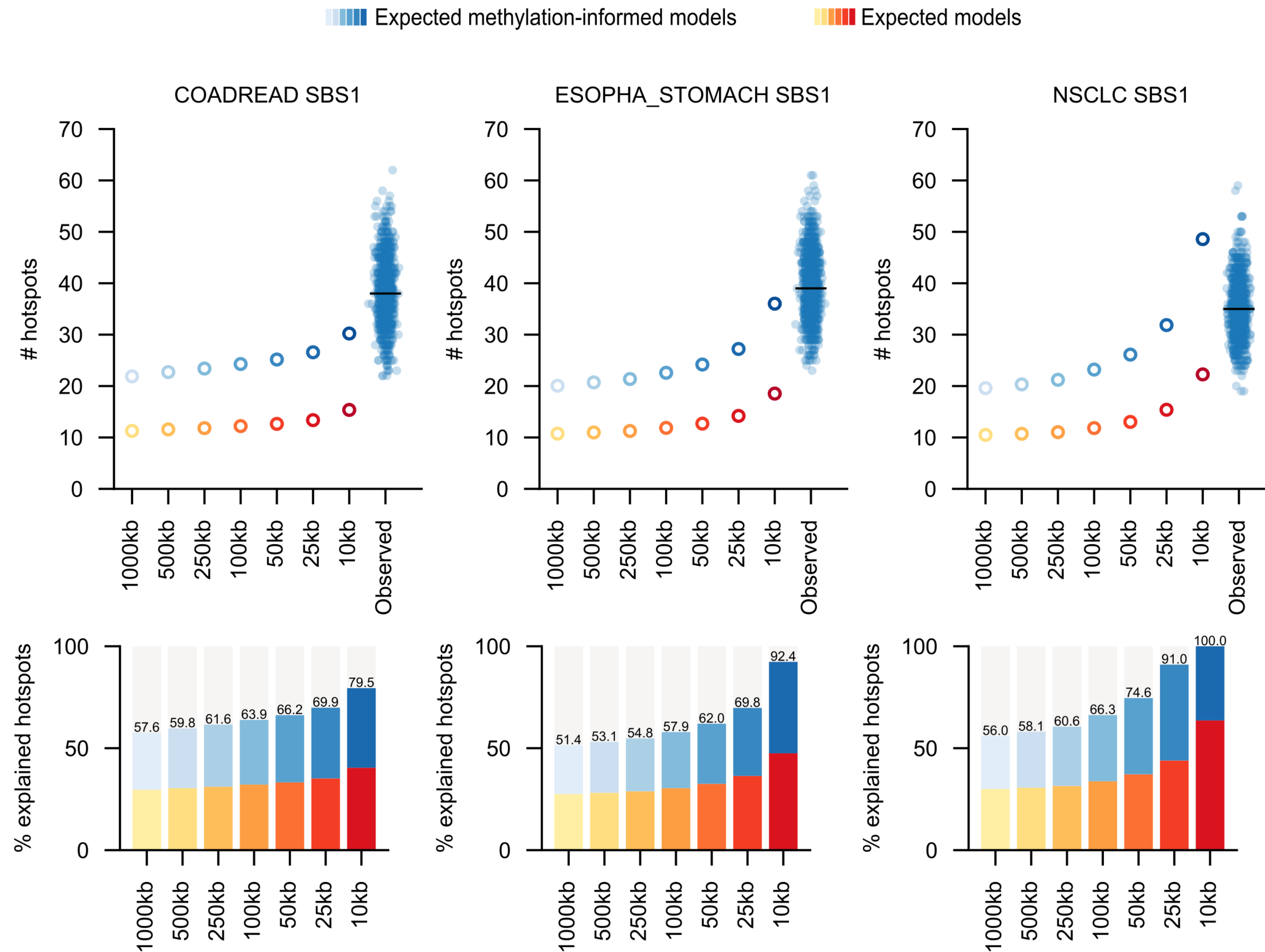




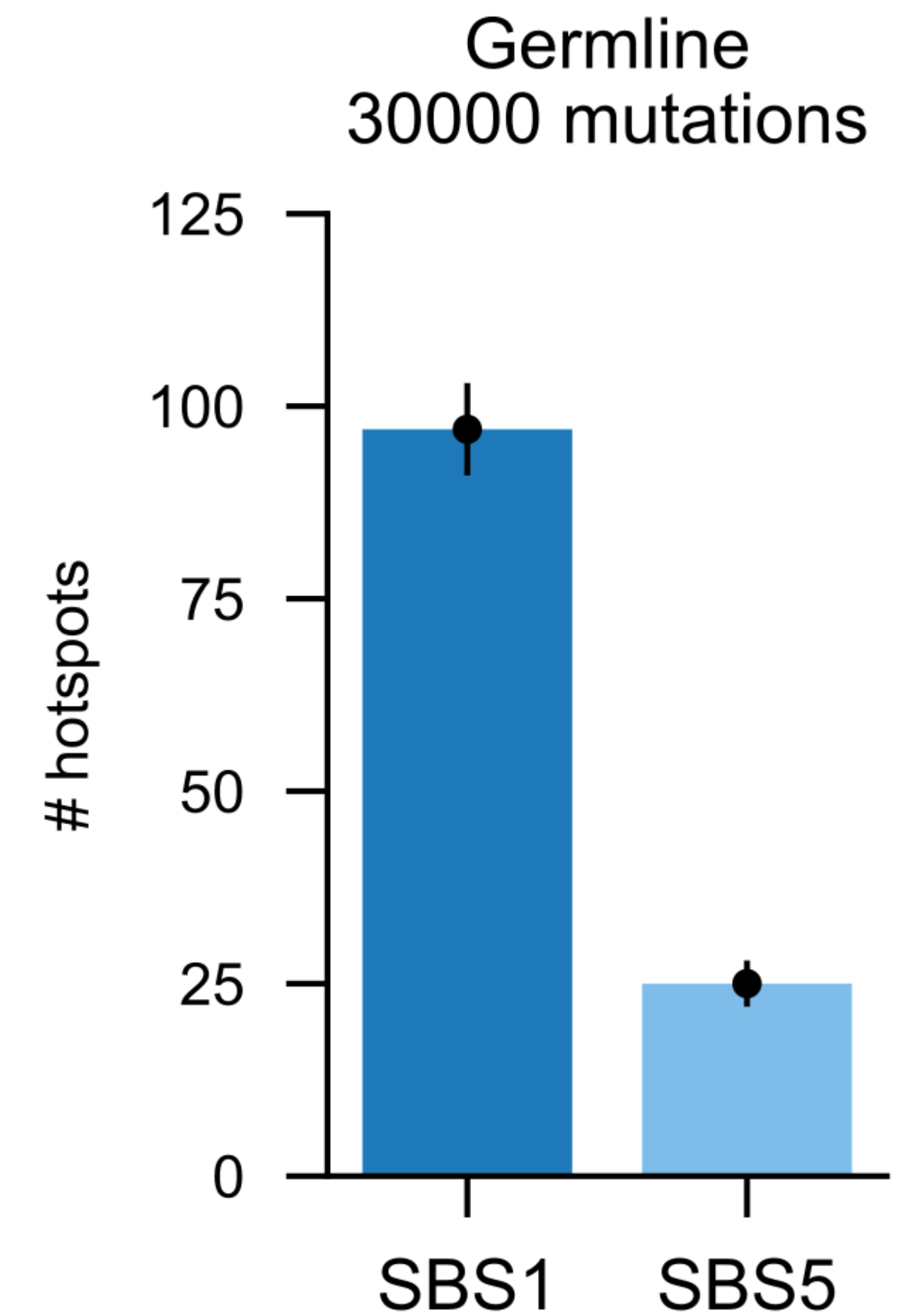
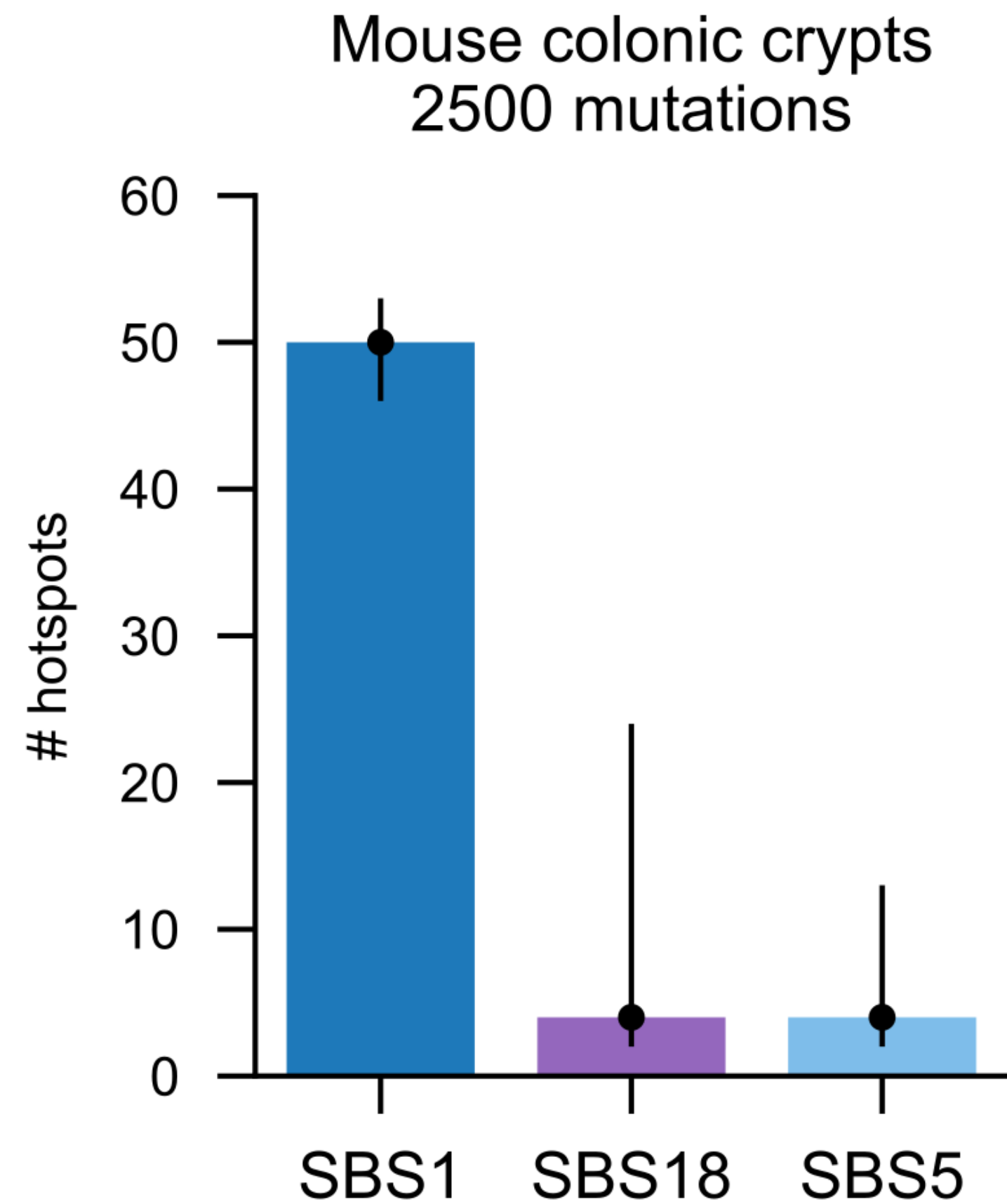
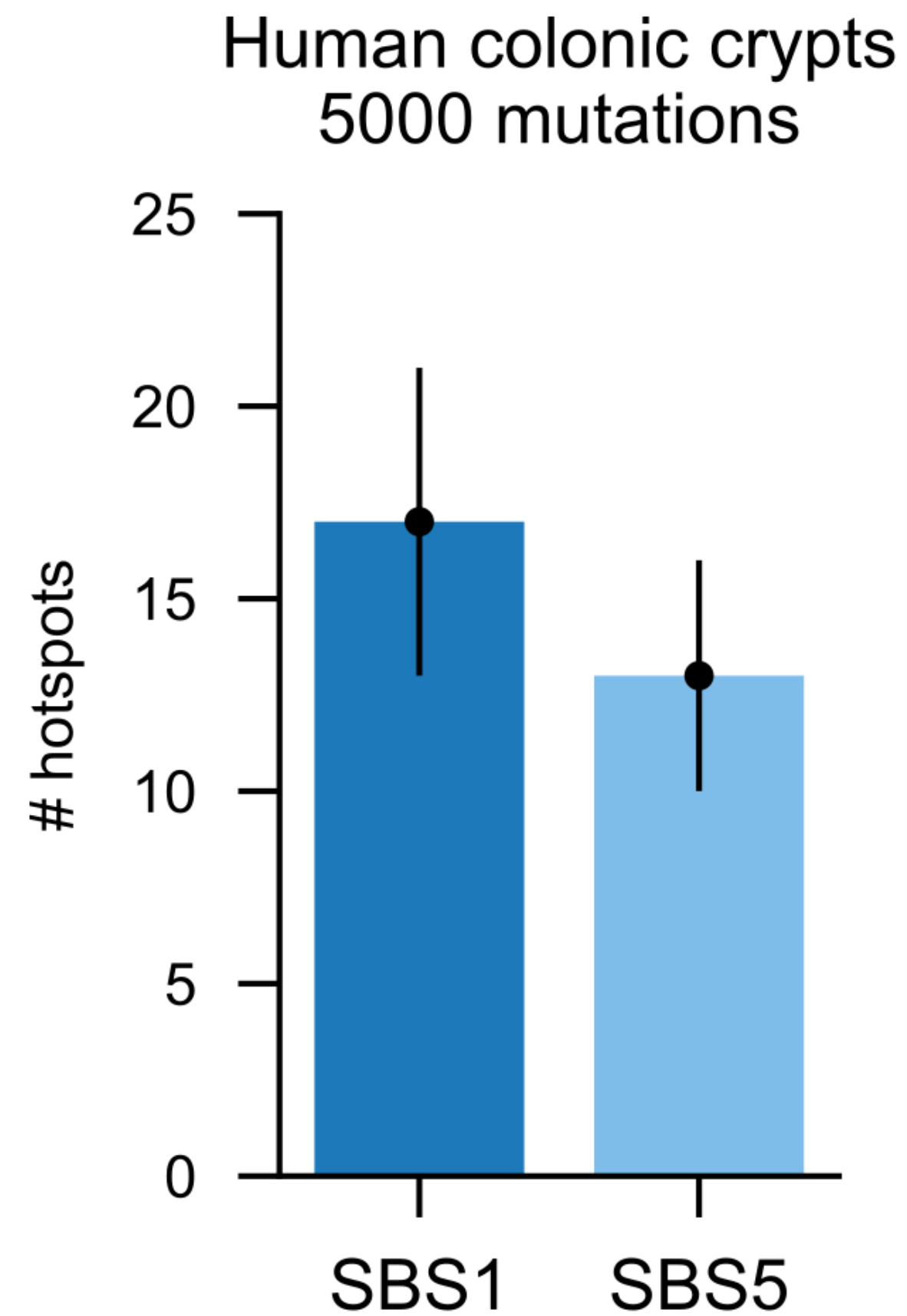
# High proportion of hotspots in methylated CpGs



# Genome-wide distribution of methylated CpGs sites can explain most of SBS1 hotspot propensity



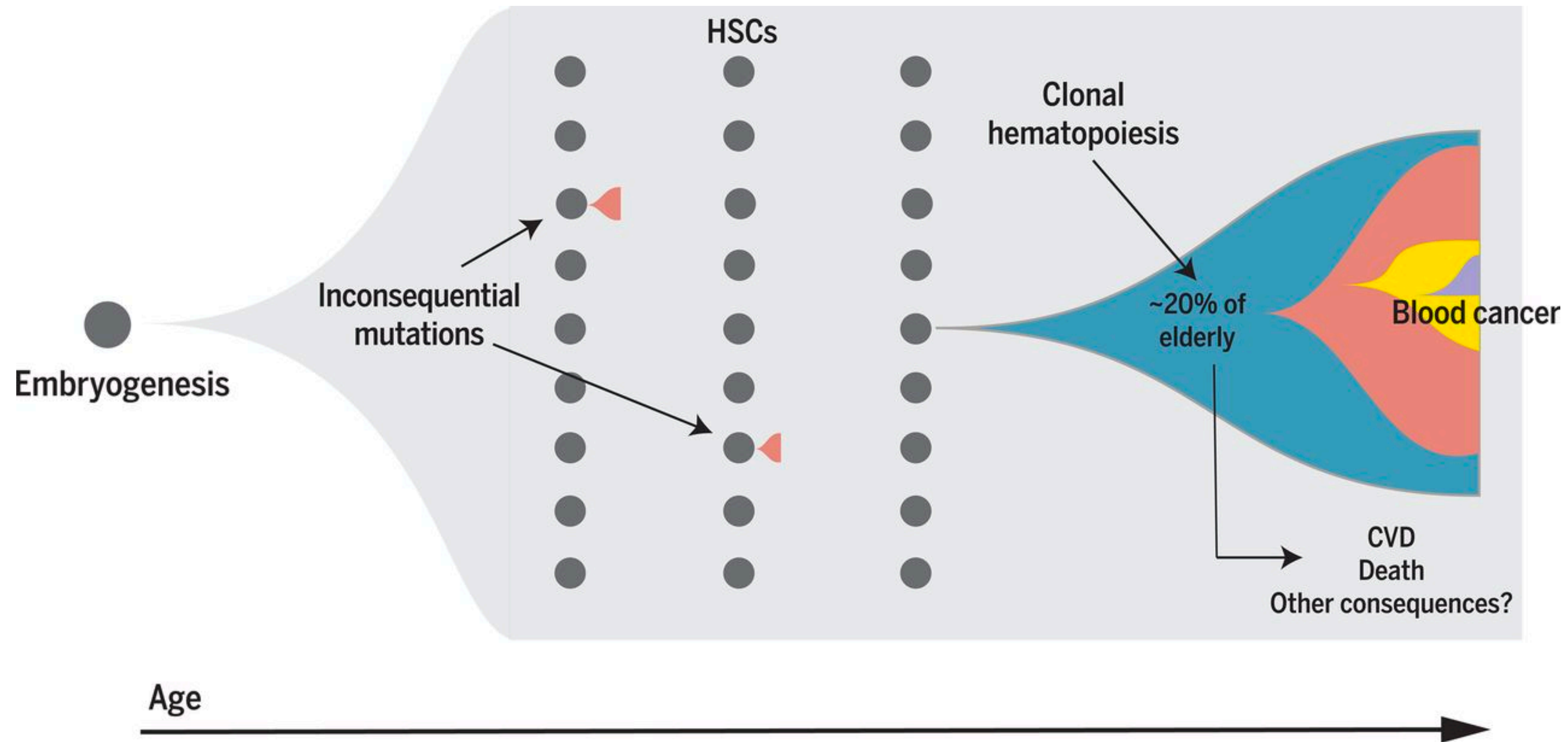
# Beyond cancer: high SBS1 hotspot propensity in normal tissues and germline variants



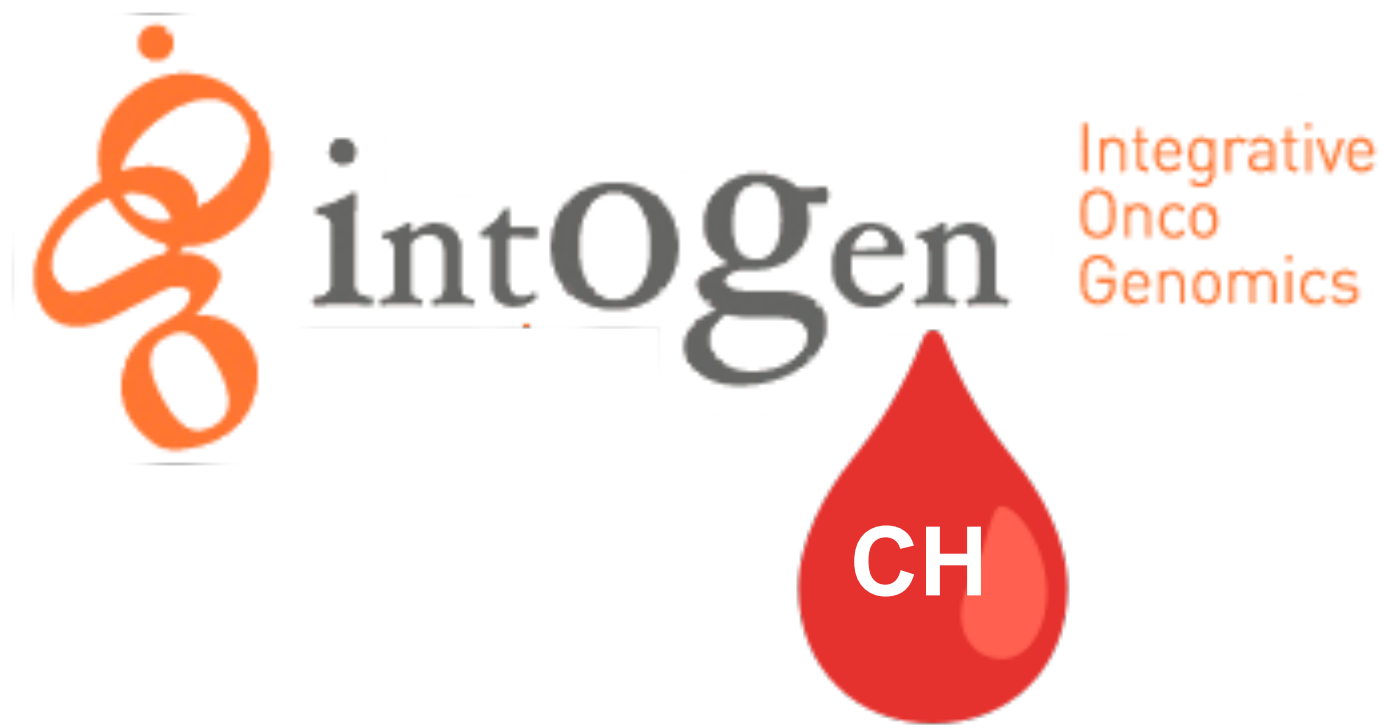


- Identifying Cancer Drivers
- Understanding Neutral Mutagenesis
- Identifying Clonal Hematopoiesis Drivers
- Chemotherapy effect in hematopoiesis
- Cancer Promotion

# Clonal expansions in normal tissue

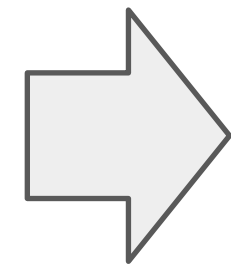


# Identification of clonal hematopoiesis driver genes and driver mutations

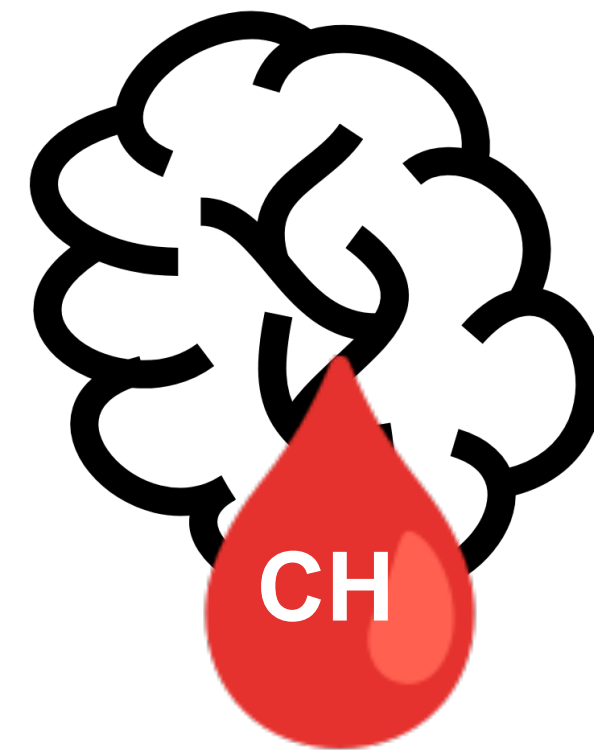


<https://www.intogen.org/ch/>

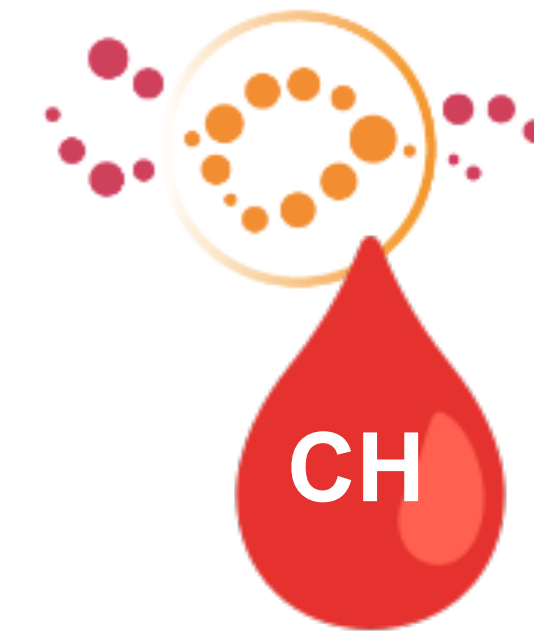
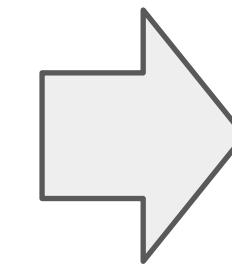
CH driver genes



BoostDM



CH driver mutations



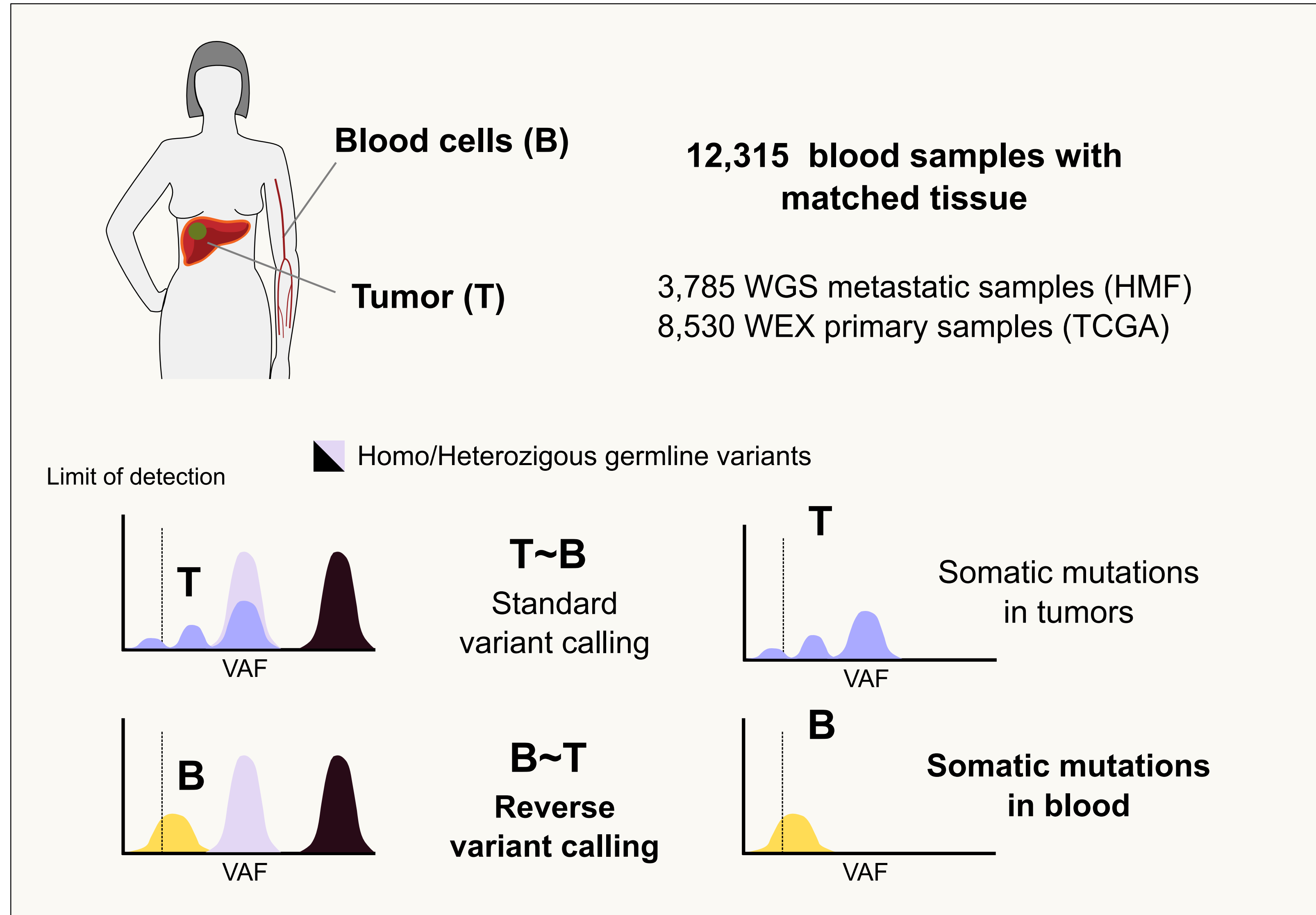
Clinical interpretation of CH mutations

CANCER GENOME INTERPRETER

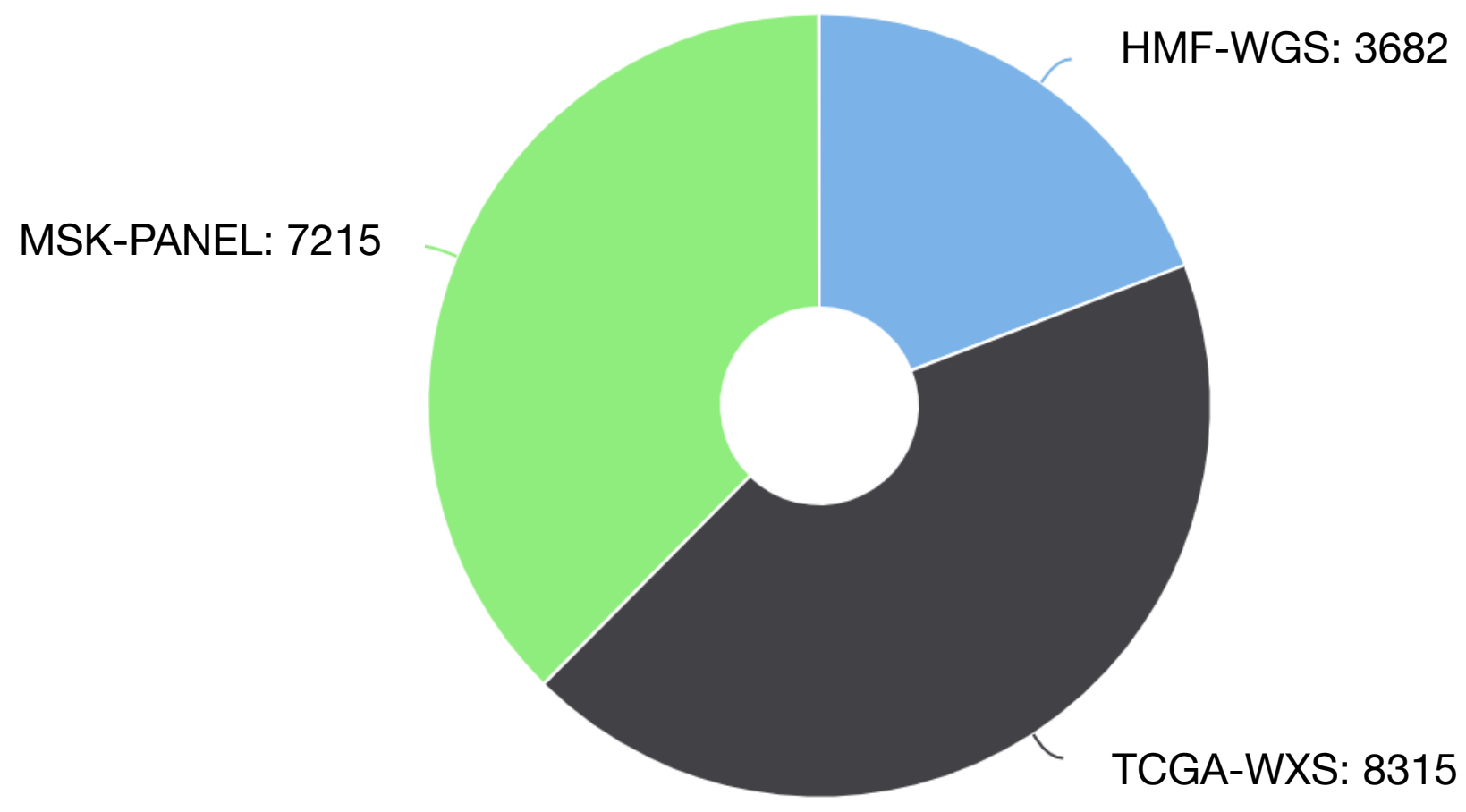




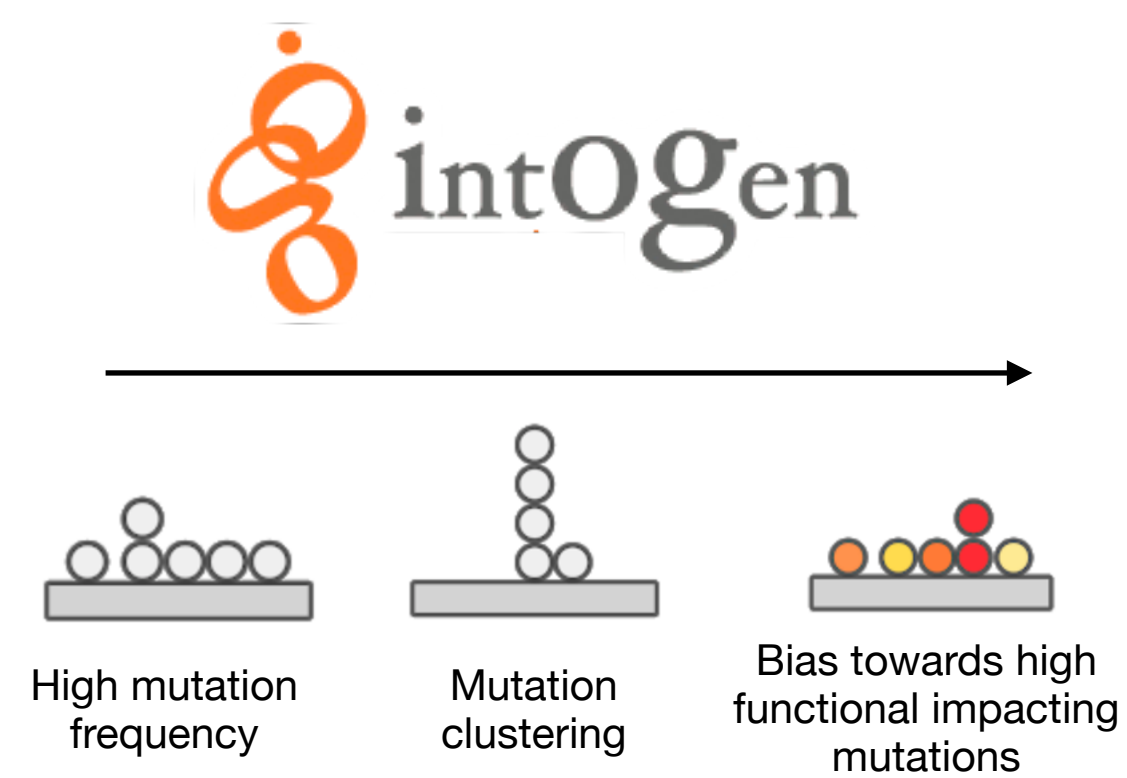
# Exploiting cancer genomics data to identify blood somatic mutations



# Discovering Clonal Hematopoiesis Driver Genes



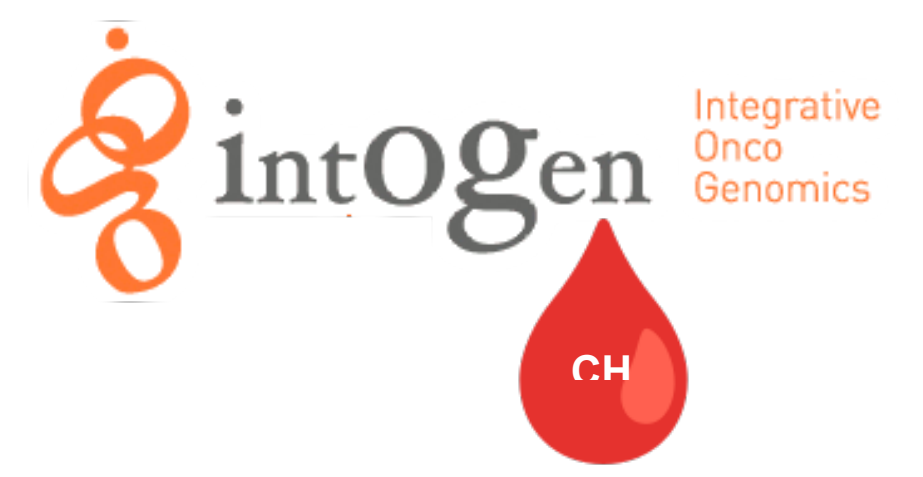
19,202 Tumors · 3 cohorts



Mutation Pattern Analysis to identify Cancer Genes

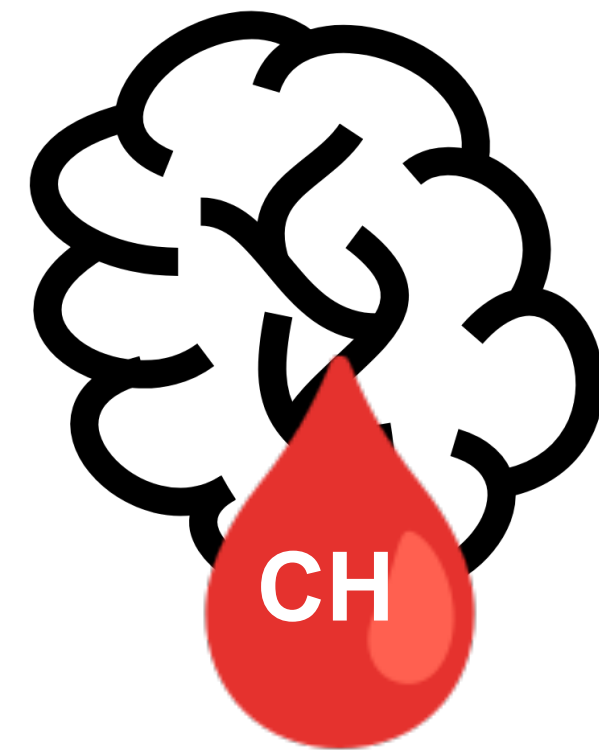


64 Clonal Hematopoiesis Driver Genes (All known + new candidates)



# Identification of Clonal Hematopoiesis Driver Mutations through In Silico Saturation Mutagenesis

BoostDM



Santi Demajo



Joan Enric Ramis



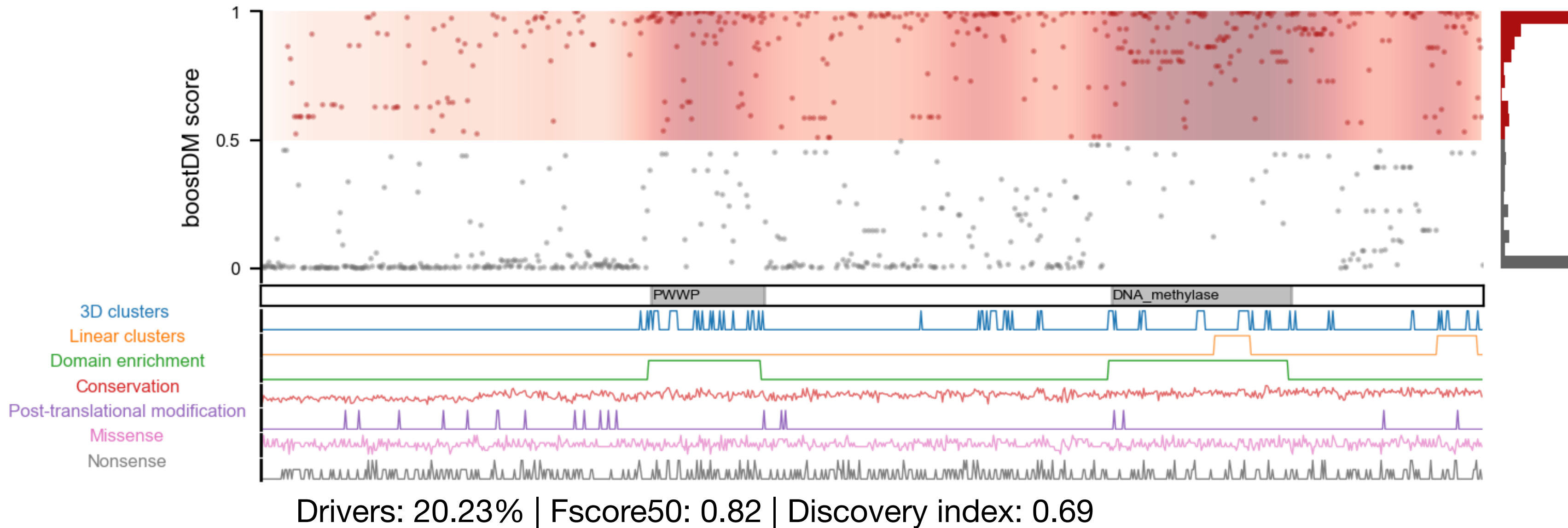
Abel Gonzalez-Perez



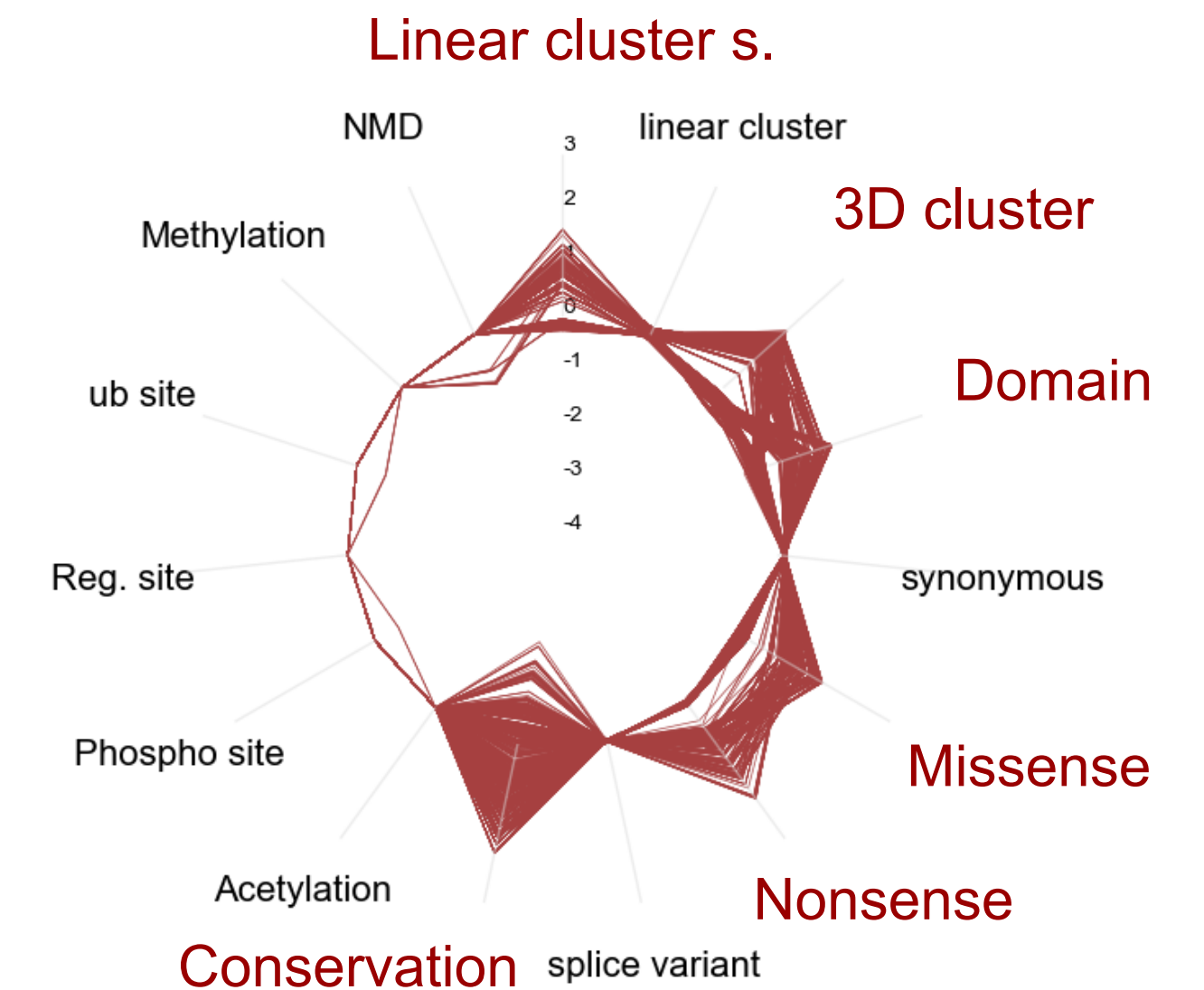
Ferran Muiños



# In silico saturation mutagenesis of DNMT3A



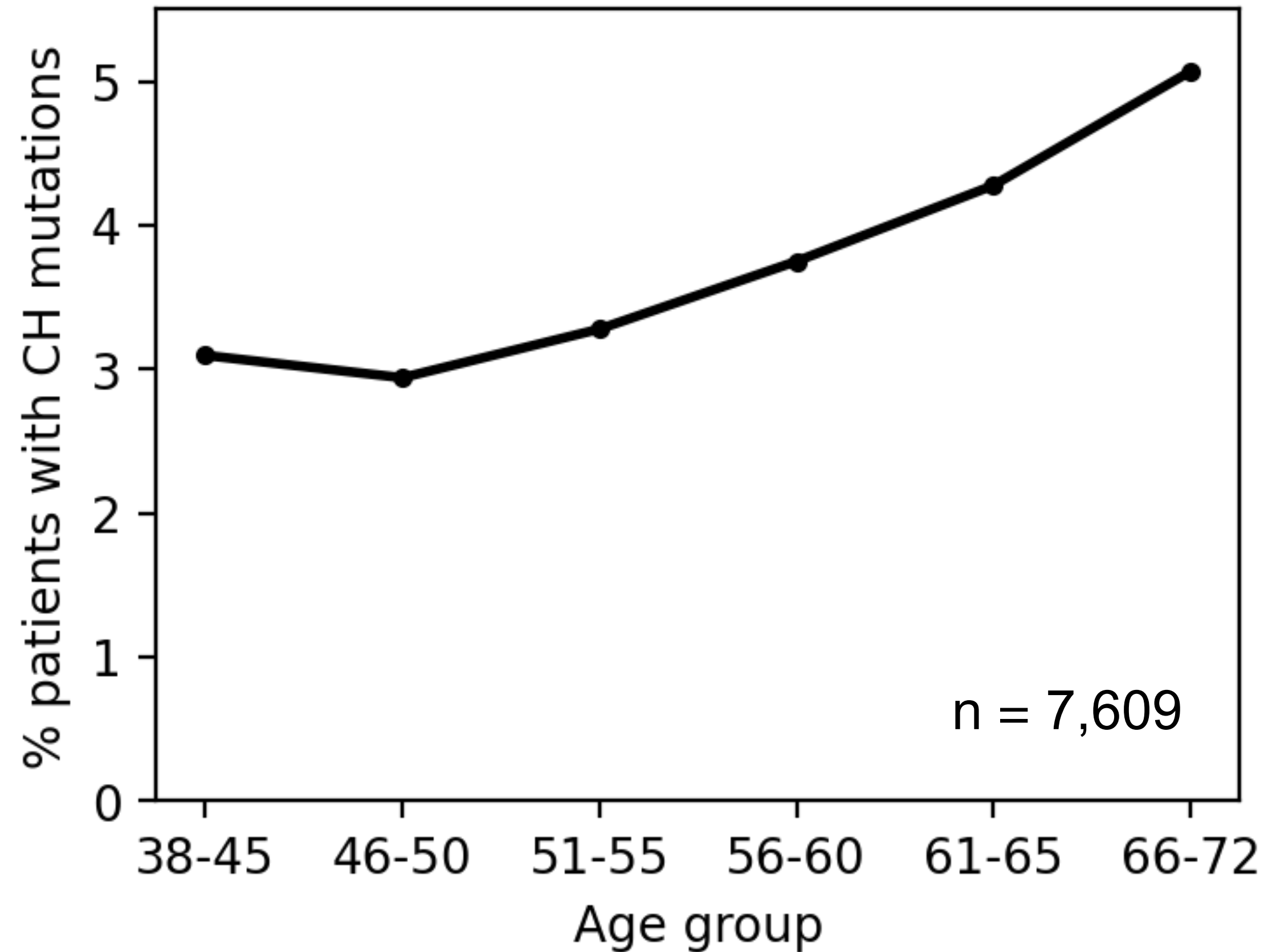
## Explanation Observed DNMT3A drivers



**All predicted DNMT3A drivers:**  
missense & nonsense mutations  
in two functional domains and two clusters

# Finding CH mutations in 200,000 individuals (UK Biobank)

Potential CH mutations in 12 CH genes



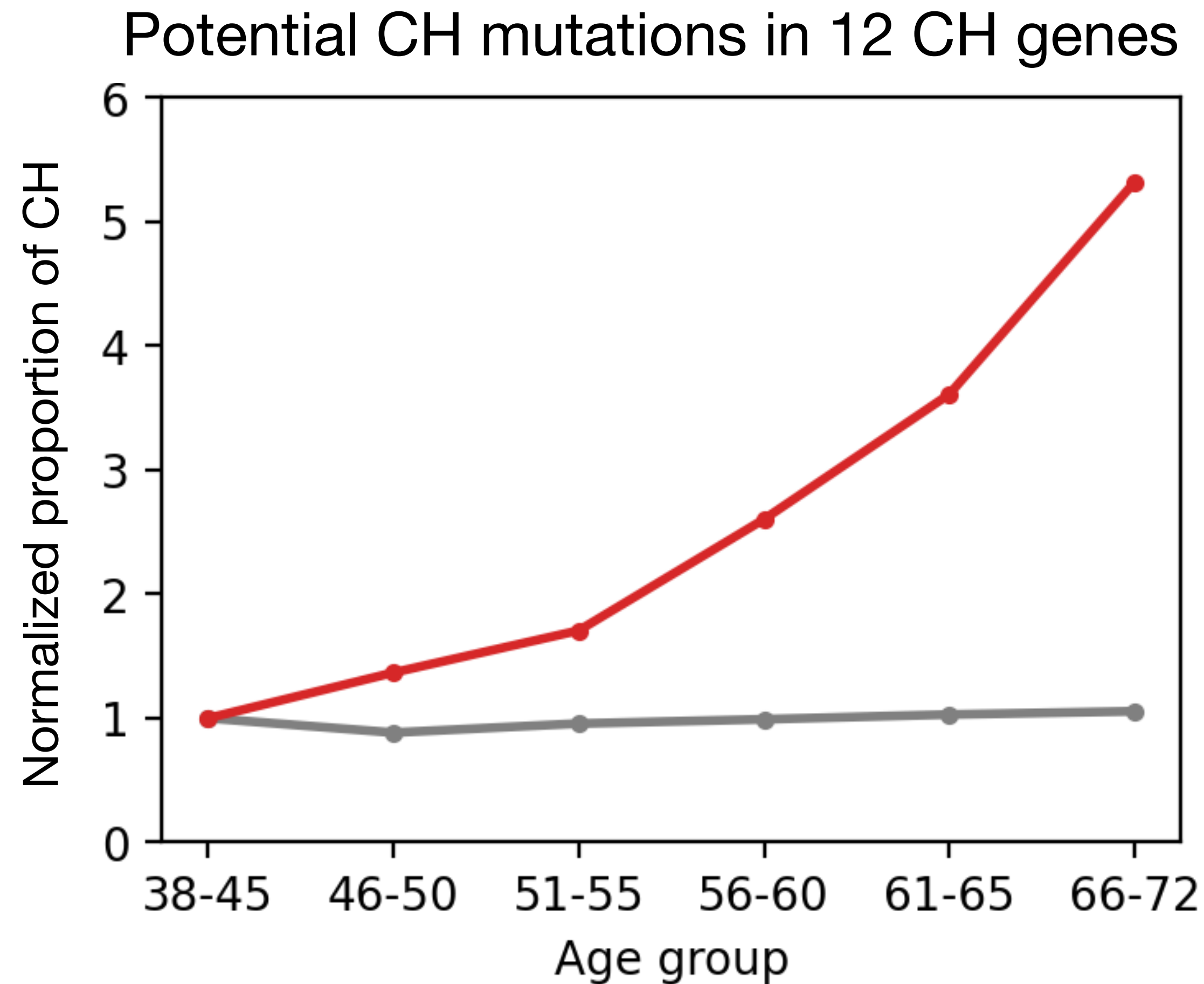
Potential CH mutations are associated with age but show a high rate of false positives

Some method to select specific mutations is needed to eliminate false positive mutations



# Only CH mutations predicted as drivers by BoostDM-CH highly correlate with age

Identification of driver mutations from the potential CH mutations by BoostDM-CH



BoostDM-CH drivers  
n = 2,298 (~30%)

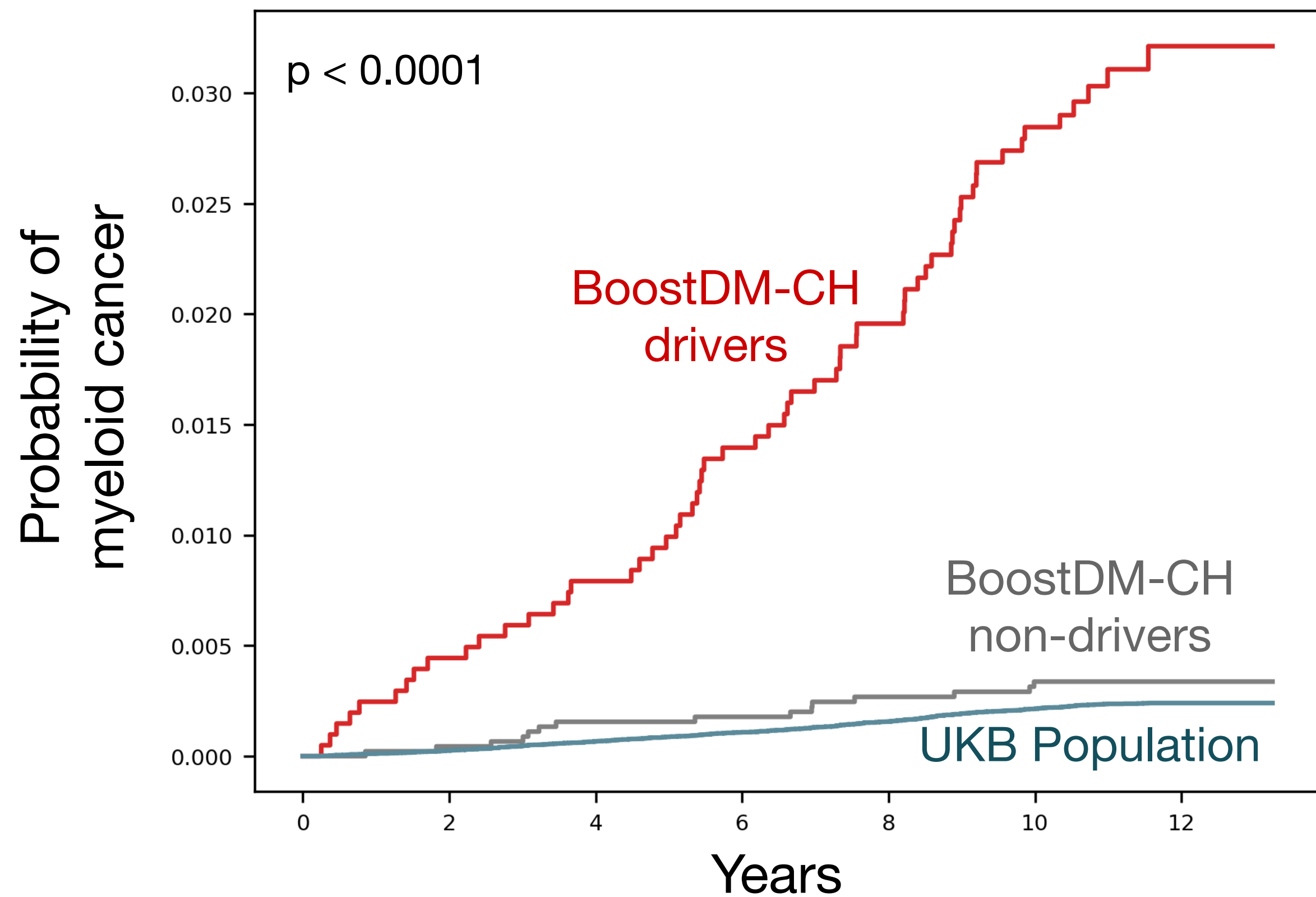
BoostDM-CH non-drivers  
n = 5,311 (~70%)

Logistic regressions:  
Drivers p-val =  $6e^{-72}$   
Non-drivers p-val = *ns*

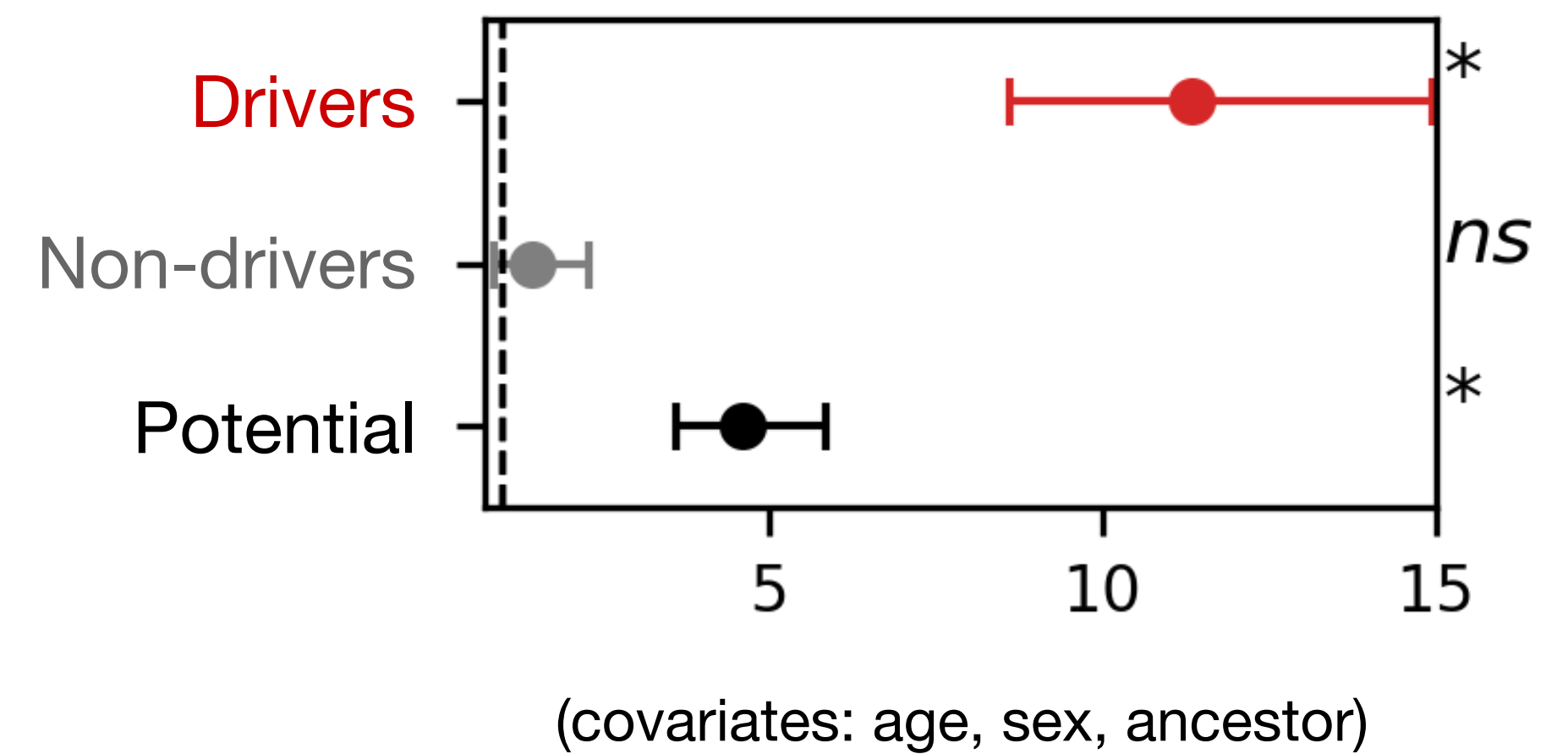


# Only CH mutations predicted as drivers by BoostDM-CH are associated with an increased risk of **hematological cancer**

Myeloid cancer risk  
(Kaplan-meier curves)

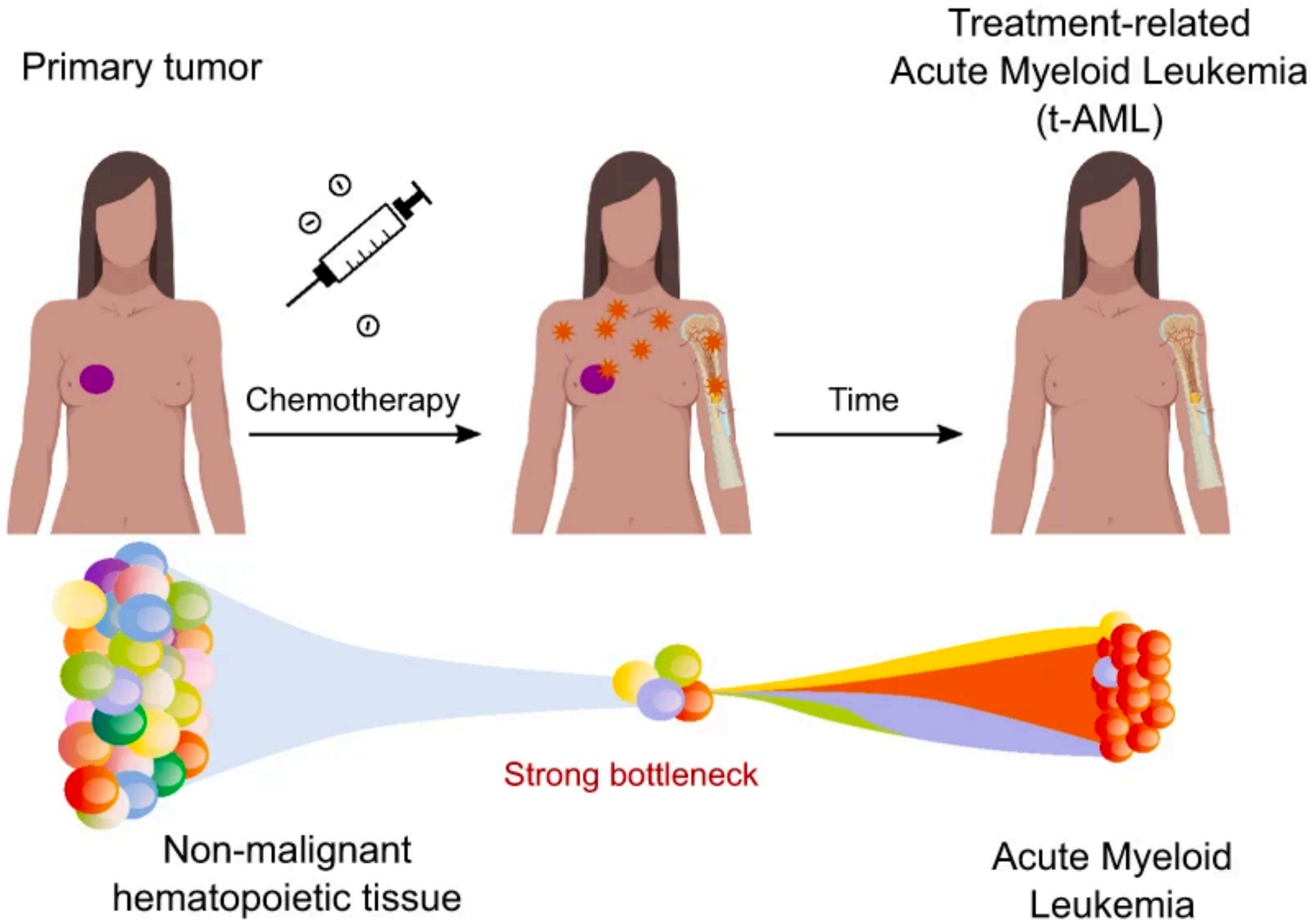


COX regression



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# Treatment-related Acute Myeloid Leukemia (tAML)



Oriol Pich



Abel Gonzalez-Perez

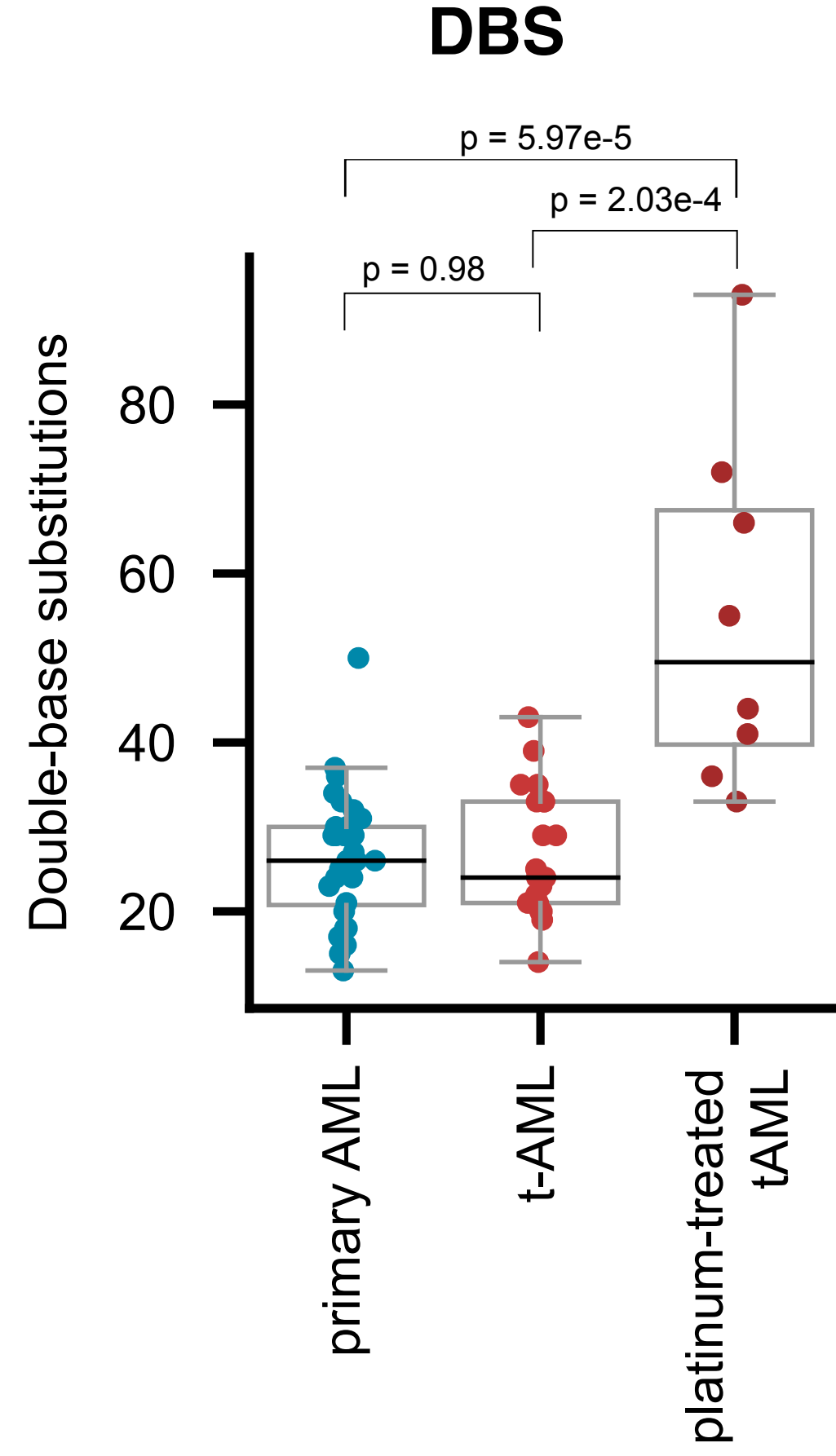
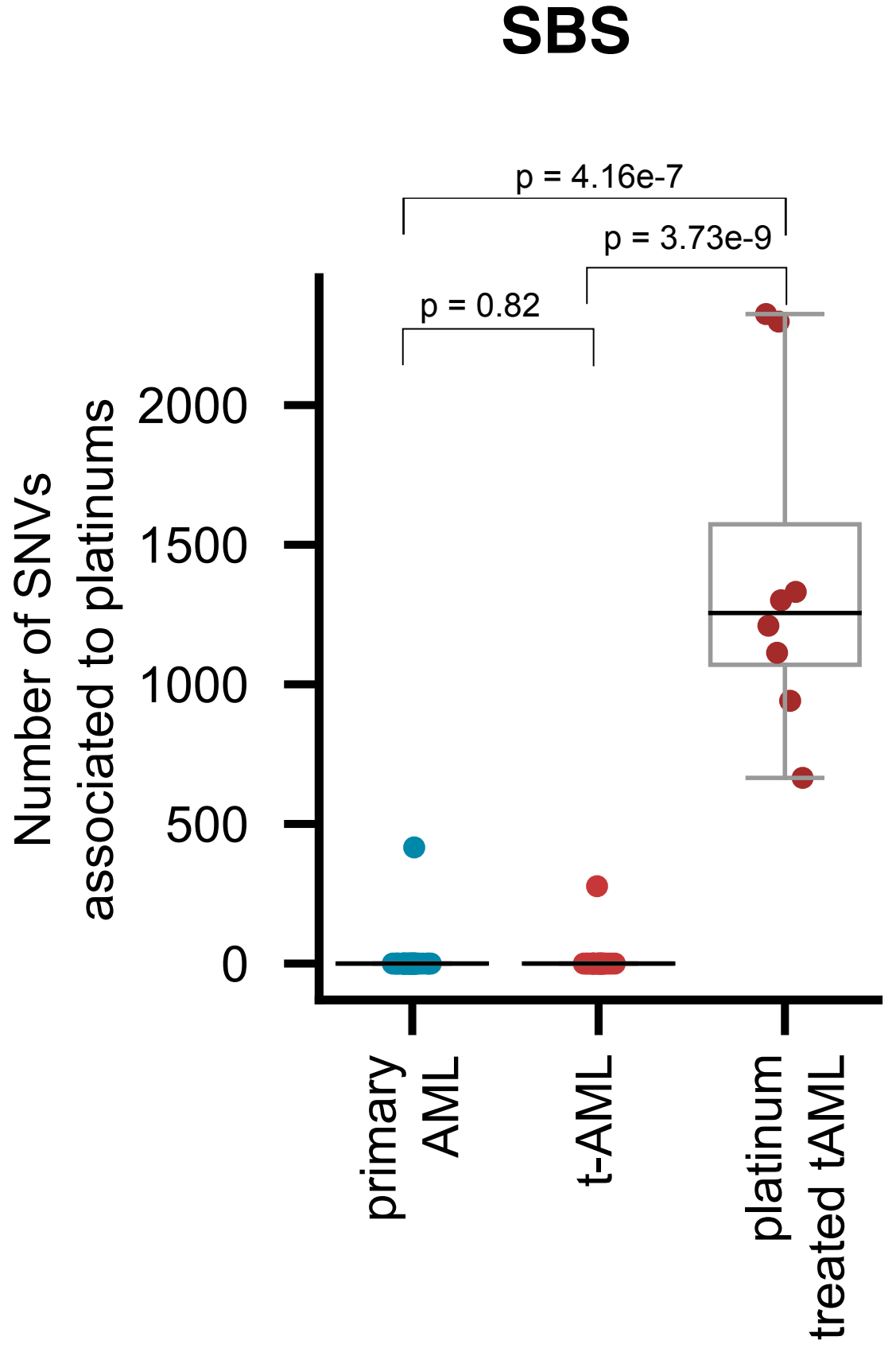
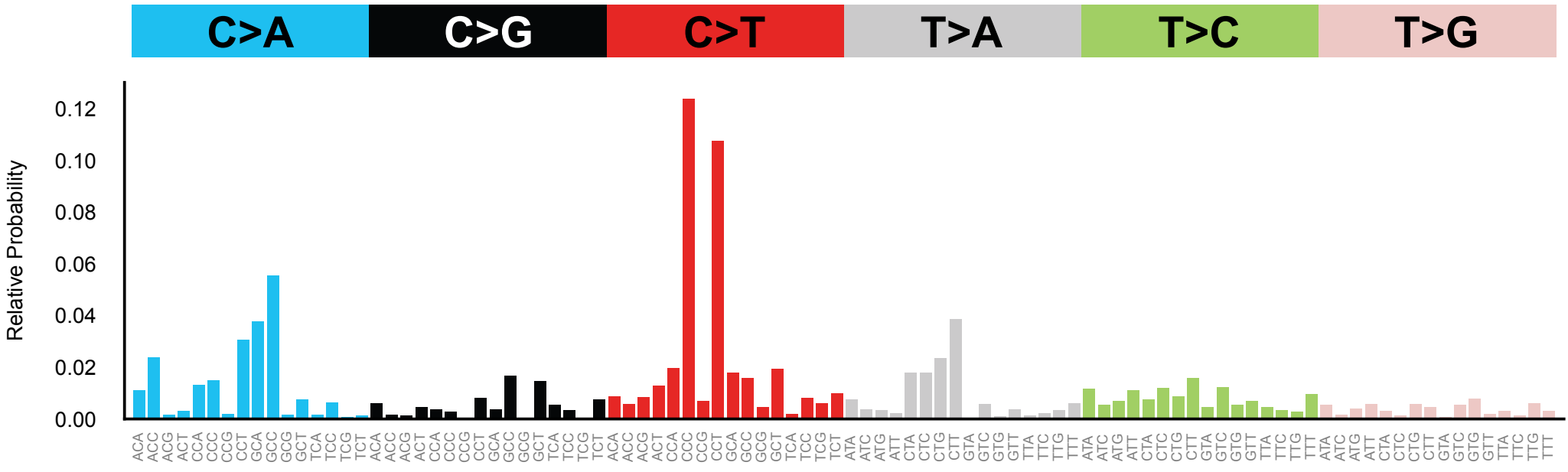


Albert Cortés  
Marta Pratcorona

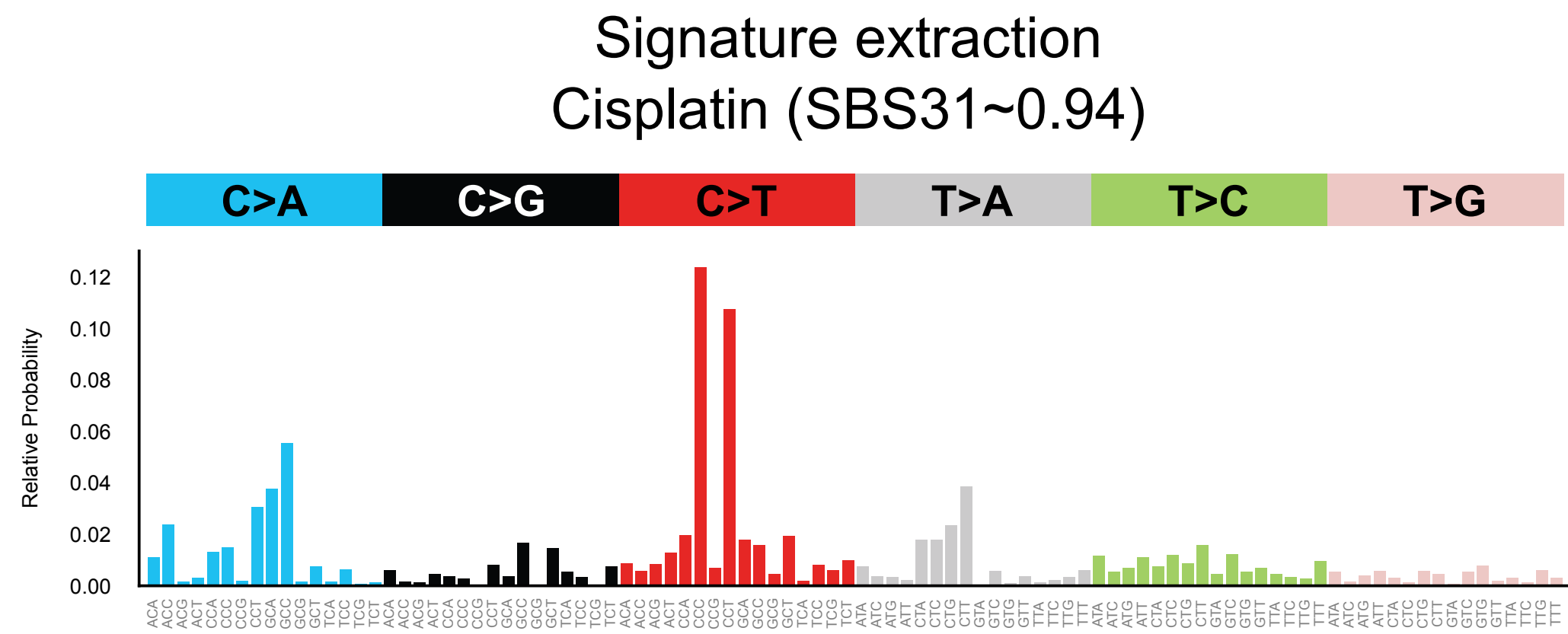


# Platinum mutations detected as clonal in tAML

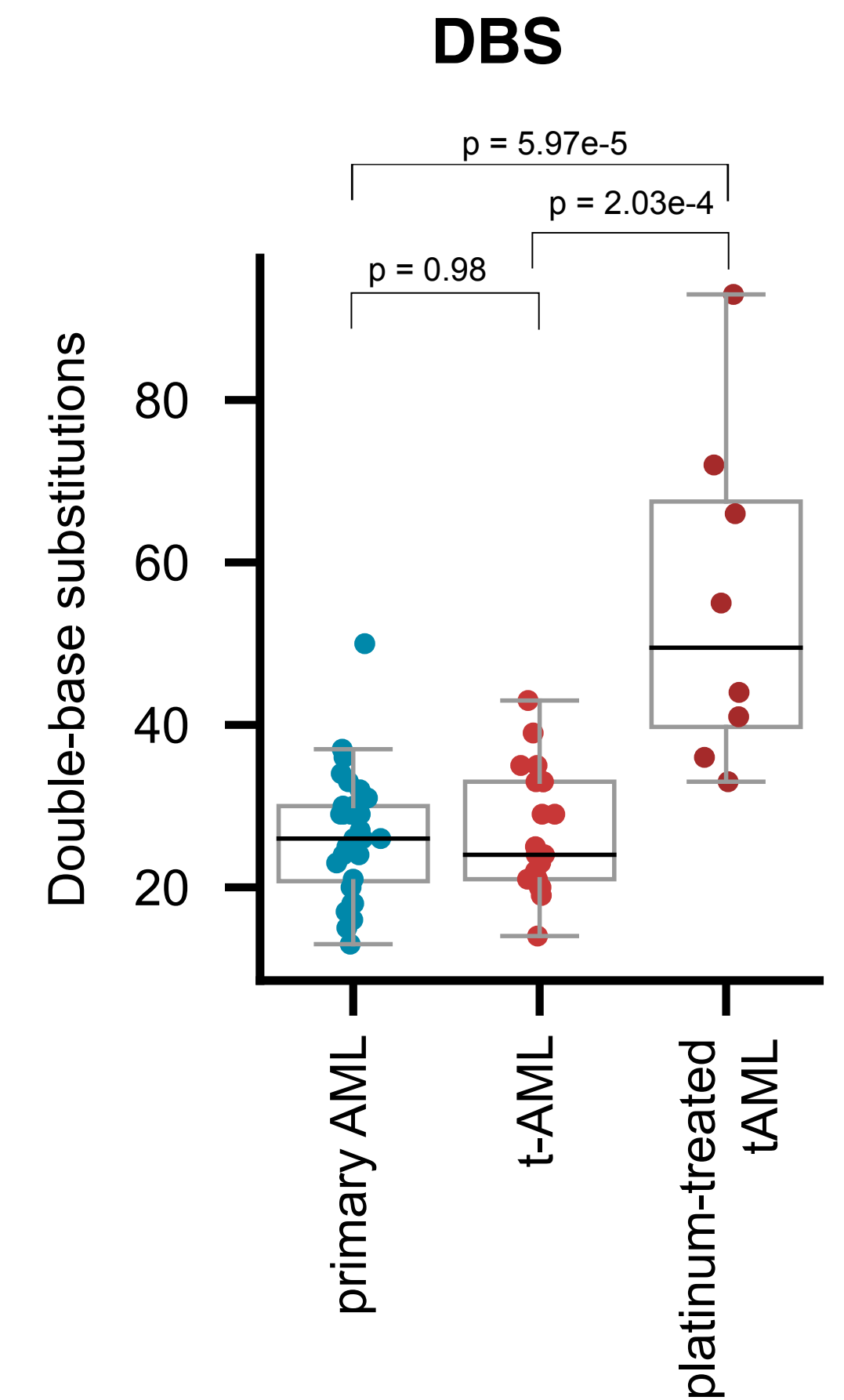
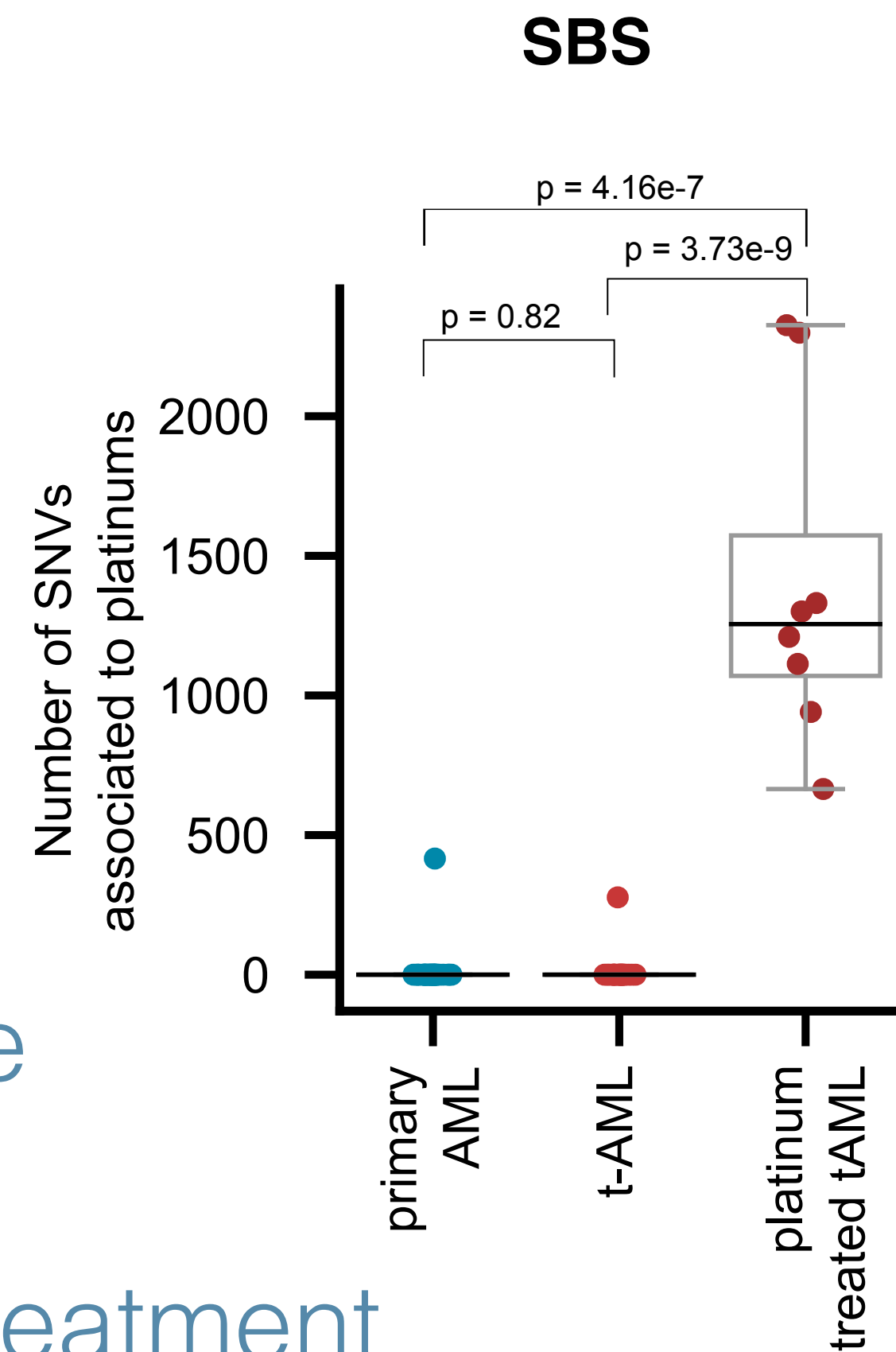
Signature extraction  
Cisplatin (SBS31~0.94)



# Platinum mutations detected as clonal in tAML

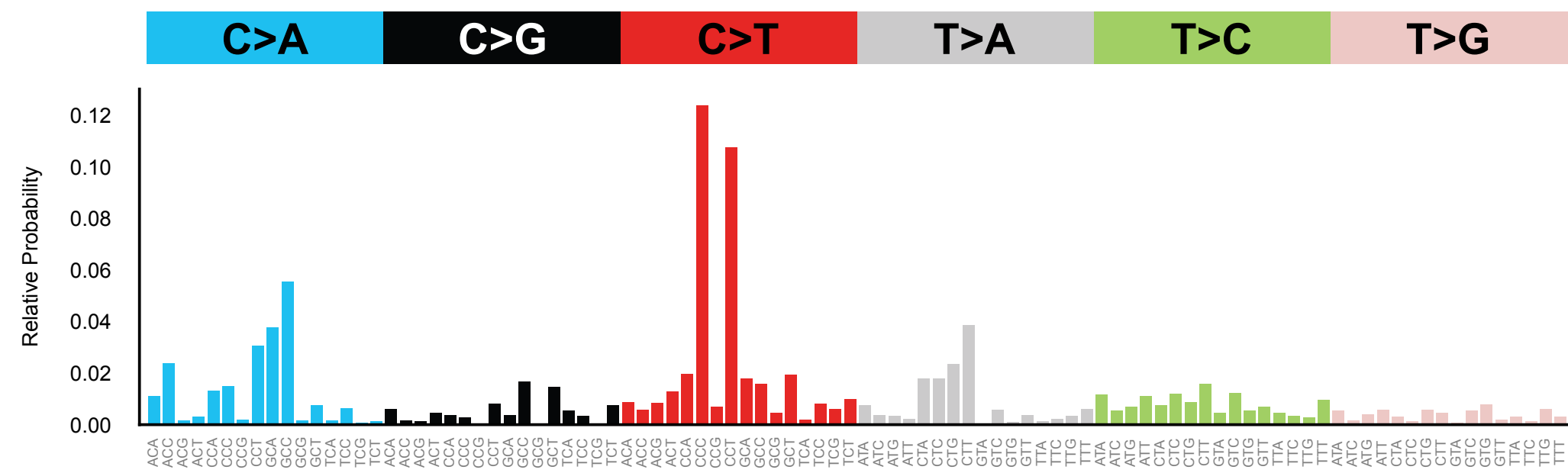


1. Healthy hematopoietic cells receive platinum mutations
2. Full clonal expansion posterior to treatment

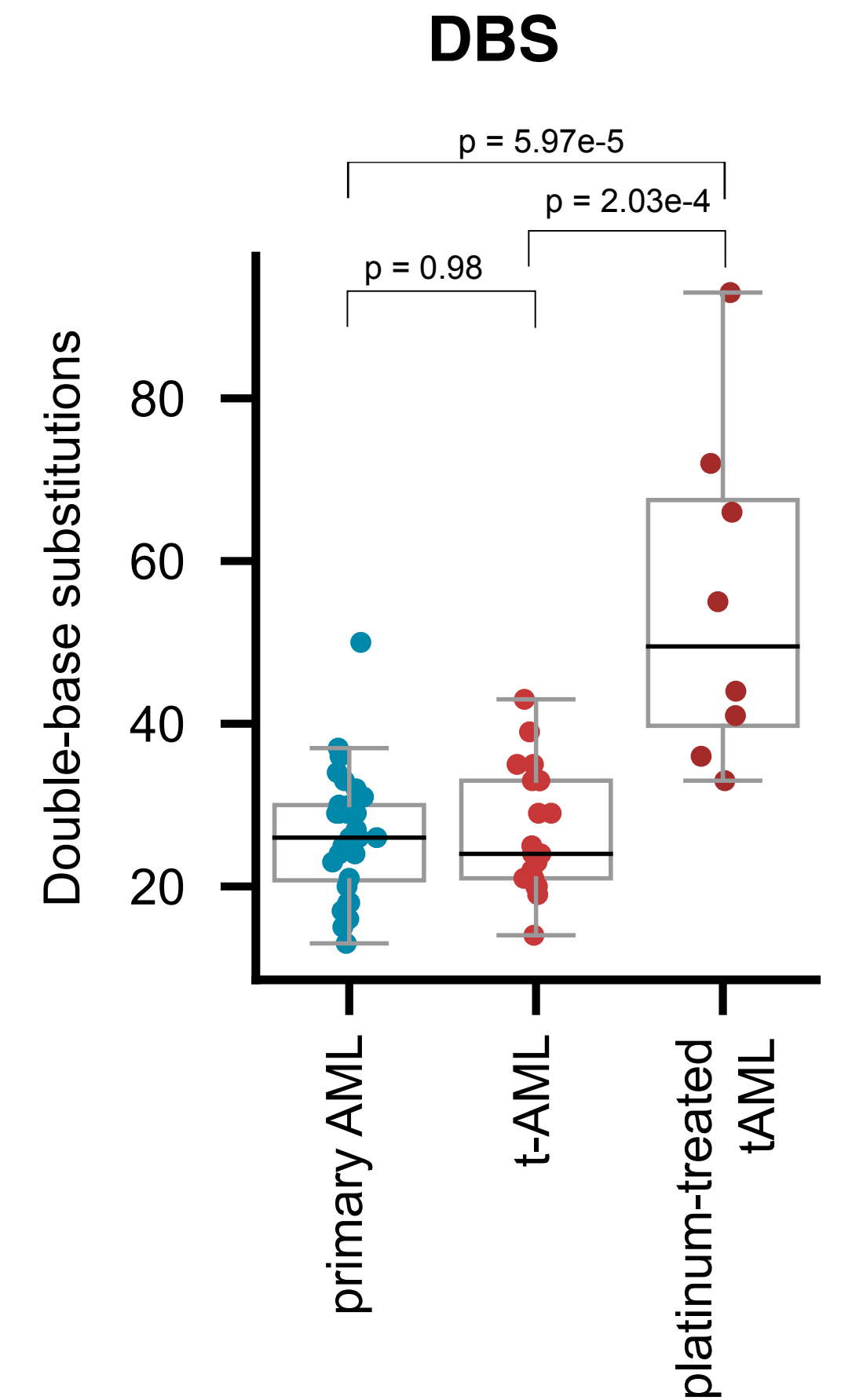
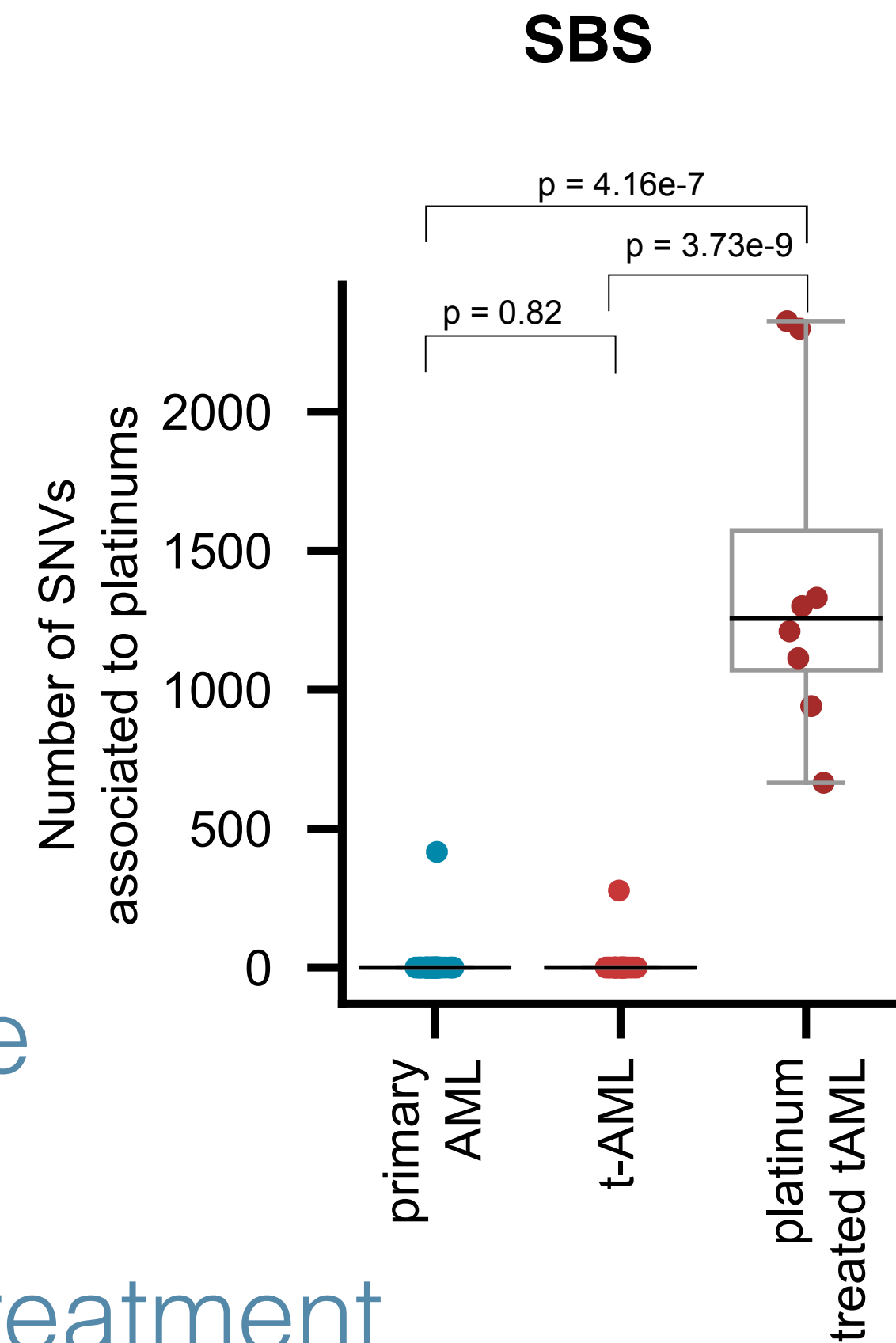


# Platinum mutations detected as clonal in tAML

Signature extraction  
Cisplatin (SBS31~0.94)



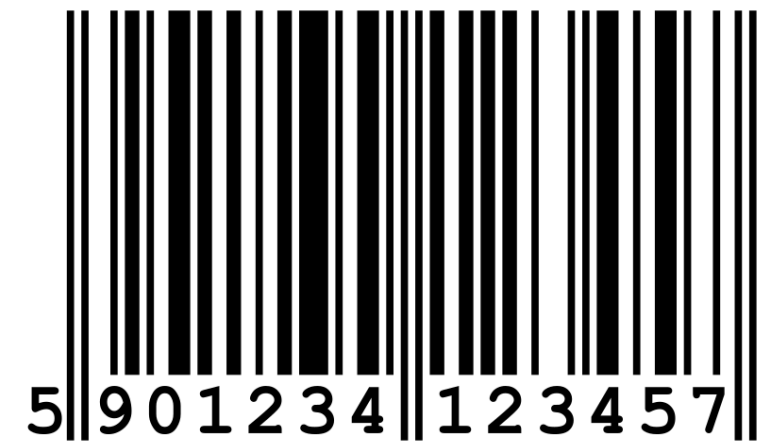
1. Healthy hematopoietic cells receive platinum mutations
2. Full clonal expansion posterior to treatment



**Mutational signature of treatment as a barcode of clonal expansion**





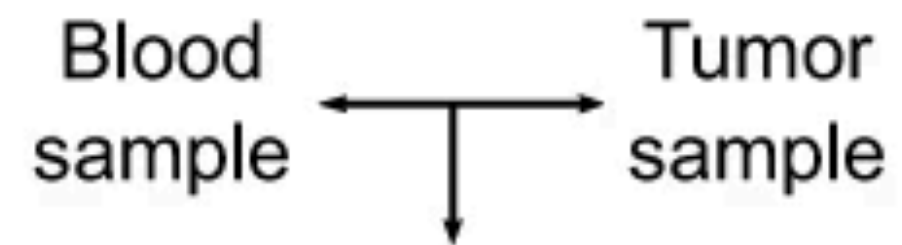


Chemotherapy mutational signatures as  
barcode to time clonal expansion before  
or after treatment

# Identifying blood somatic mutations by Reverse Mutation Calling

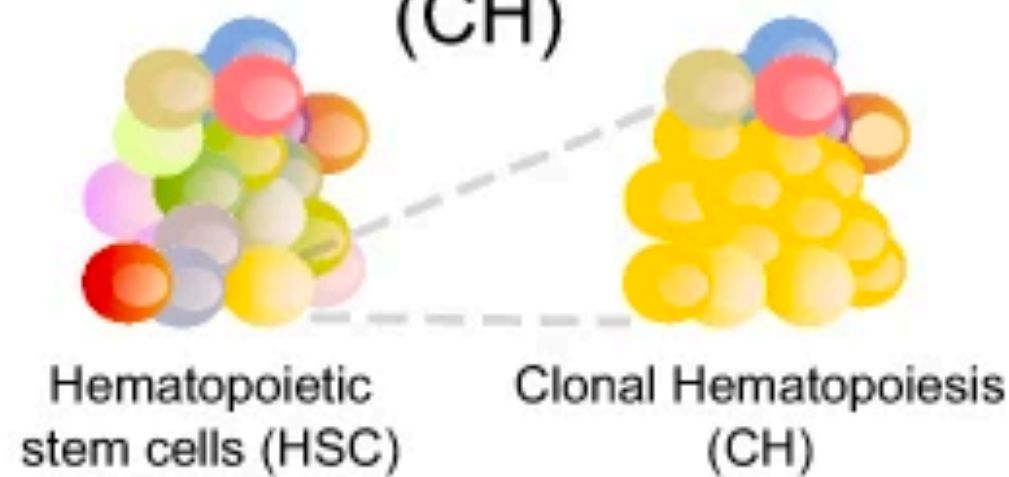


3,785 patients (metastasis cohort)  
(1,766 treated with cytotoxic therapies)



1,429,110 blood somatic mutations

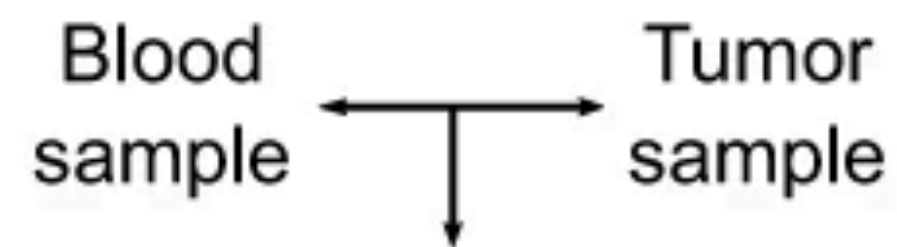
Clonal Hematopoiesis  
(CH)



# Identifying blood somatic mutations by Reverse Mutation Calling

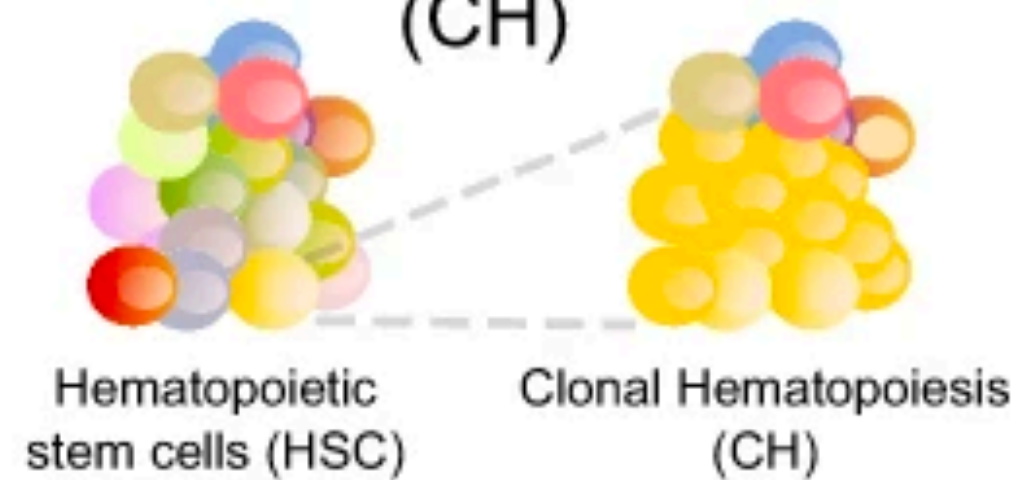


3,785 patients (metastasis cohort)  
(1,766 treated with cytotoxic therapies)



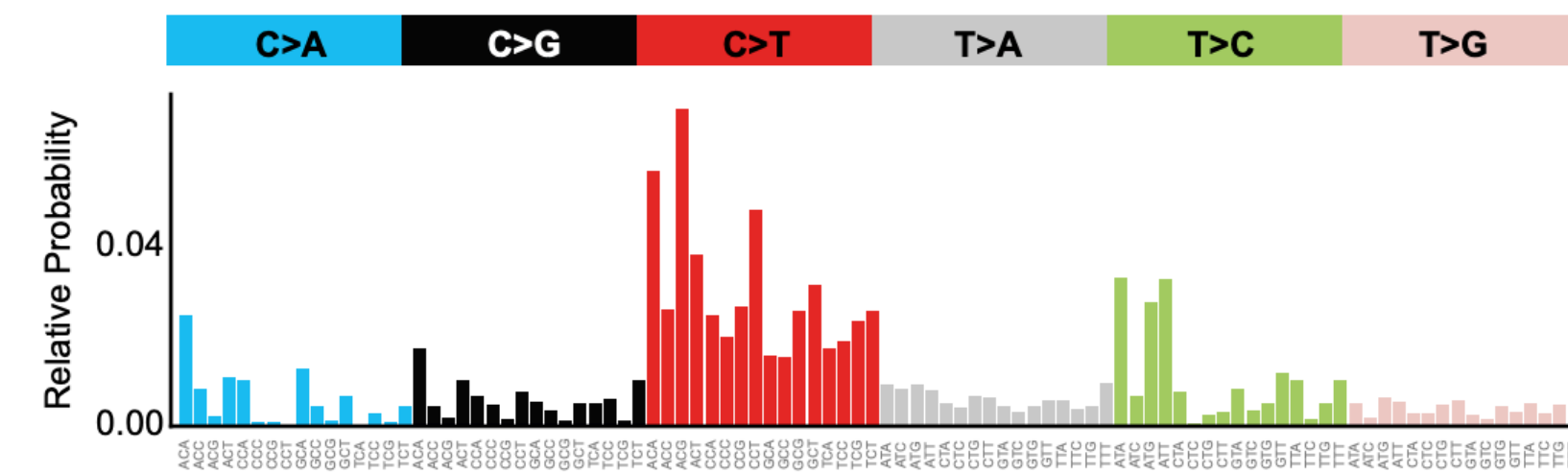
1,429,110 blood somatic mutations

Clonal Hematopoiesis (CH)



## We find HSC signature

Hematopoietic stem cell (HSC) signature in healthy blood (cos similarity = 0.96)

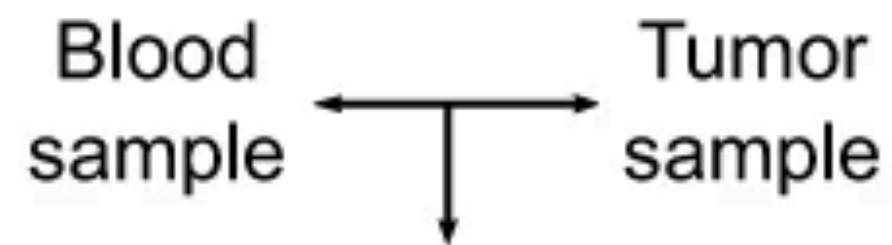




# Identifying blood somatic mutations by Reverse Mutation Calling

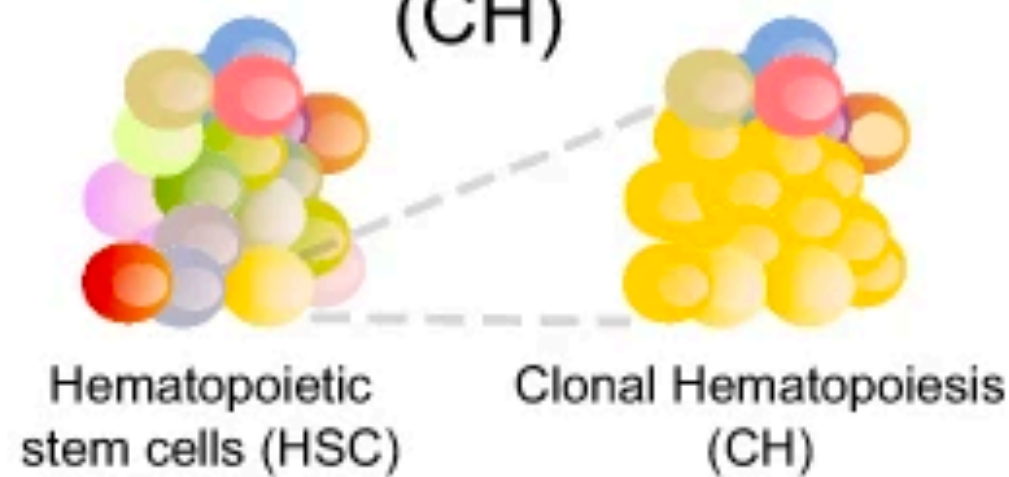


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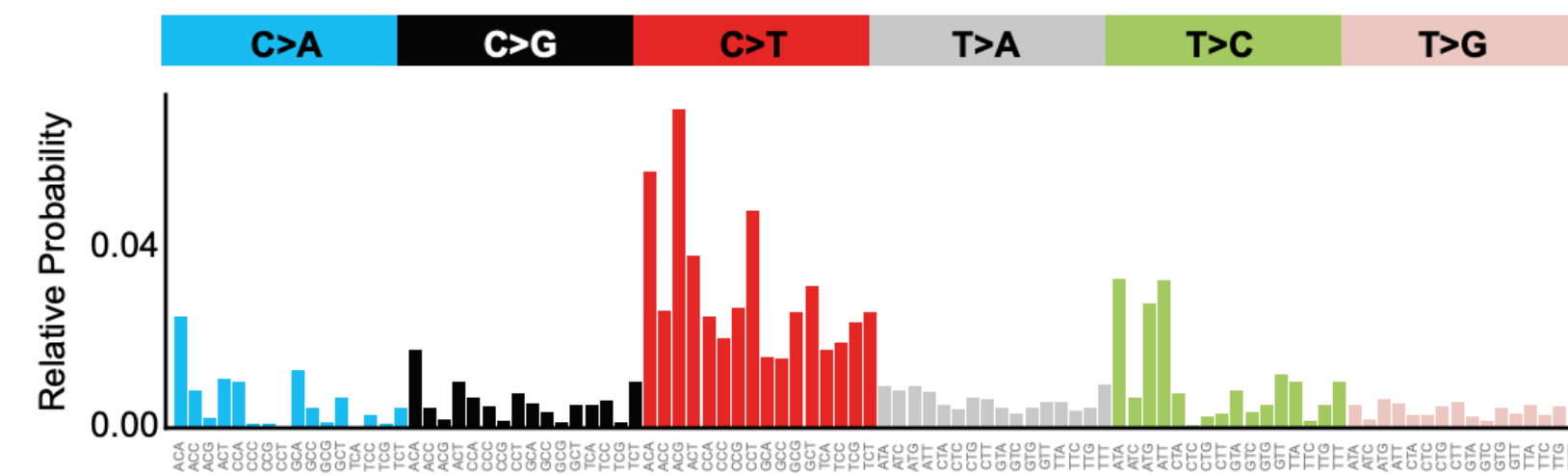
1,429,110 blood somatic mutations

Clonal Hematopoiesis (CH)



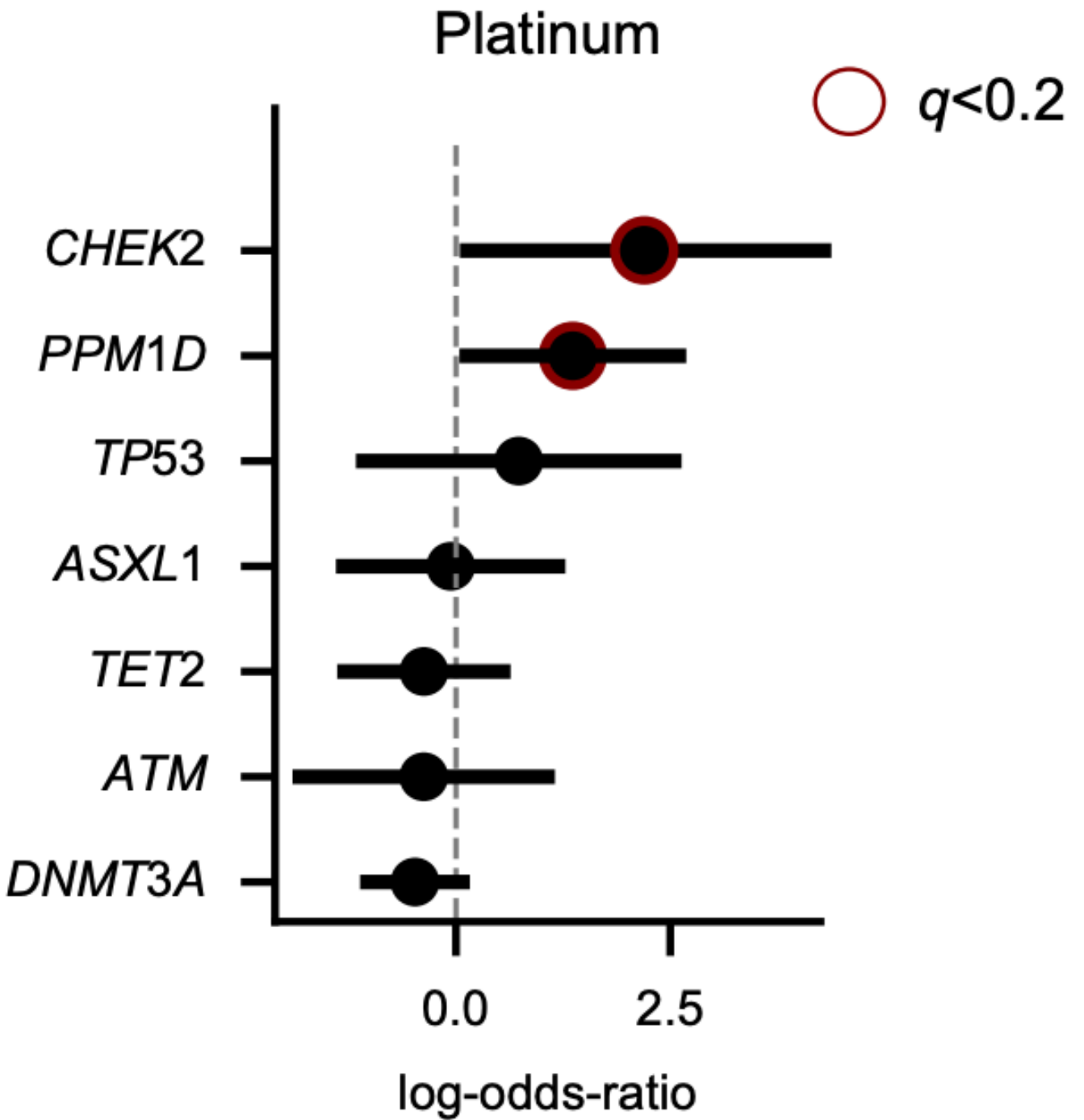
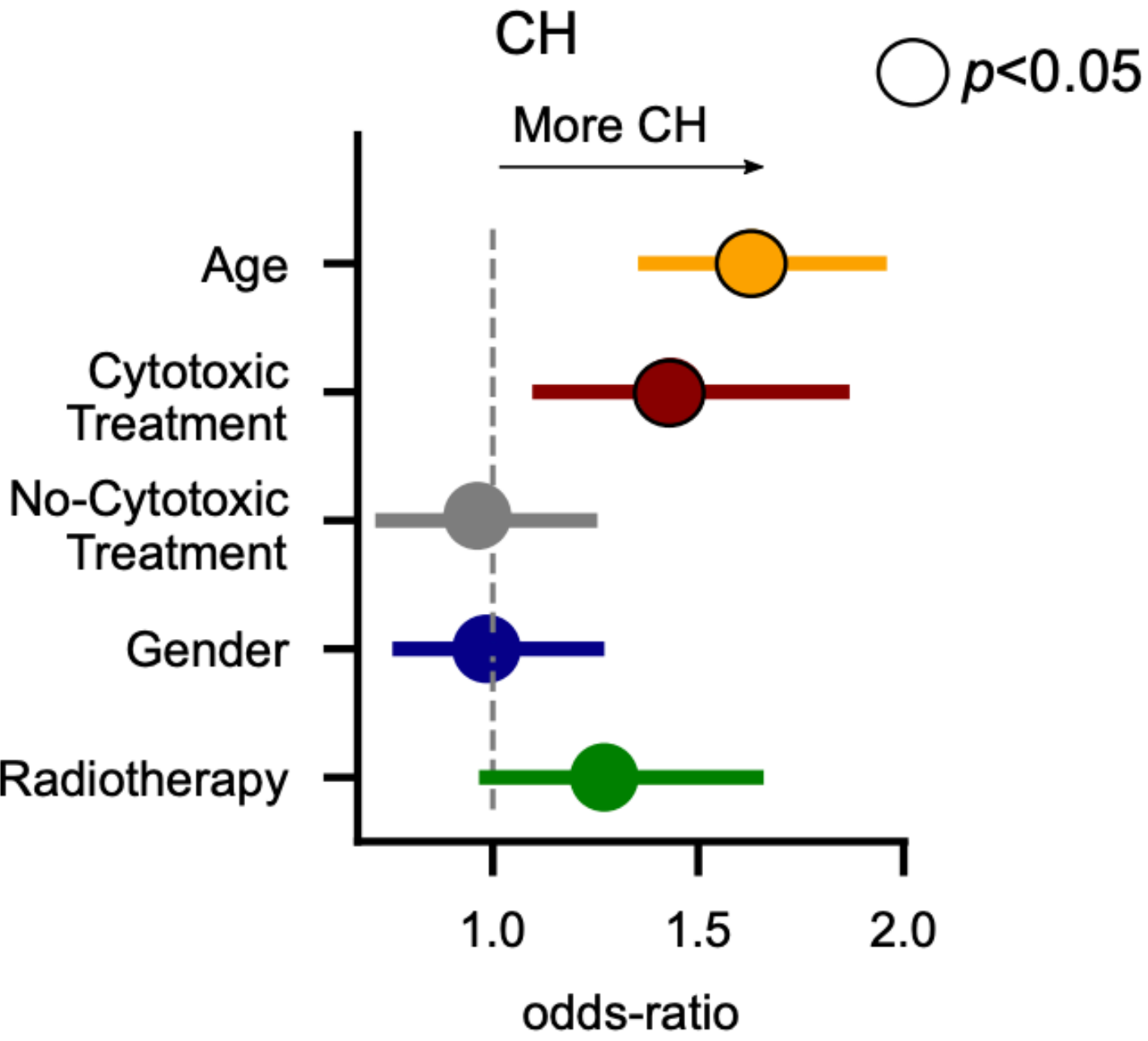
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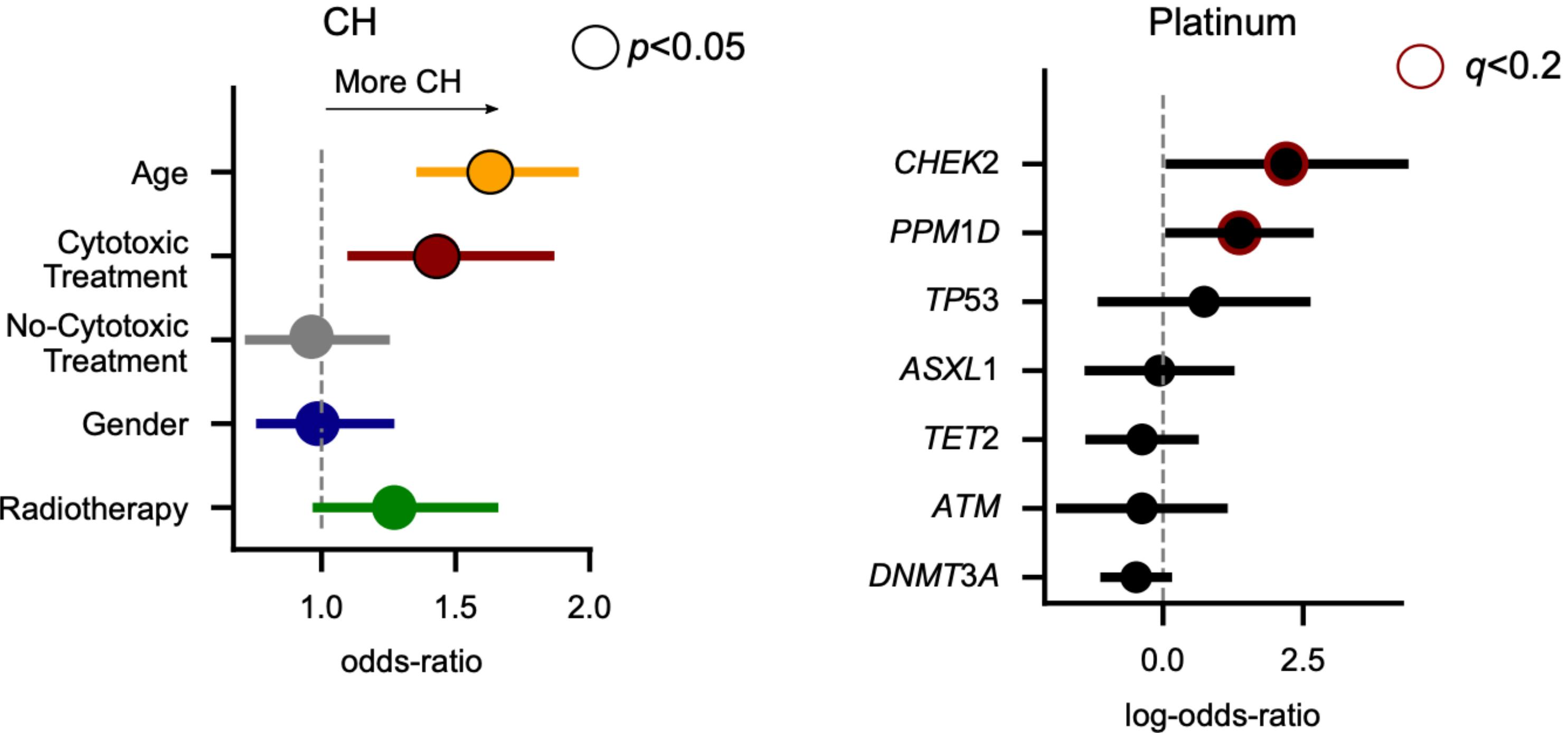


## In contrast we do not find the chemotherapy mutational signatures

# Chemotherapy is associated with clonal hematopoiesis with preference for mutations in certain genes



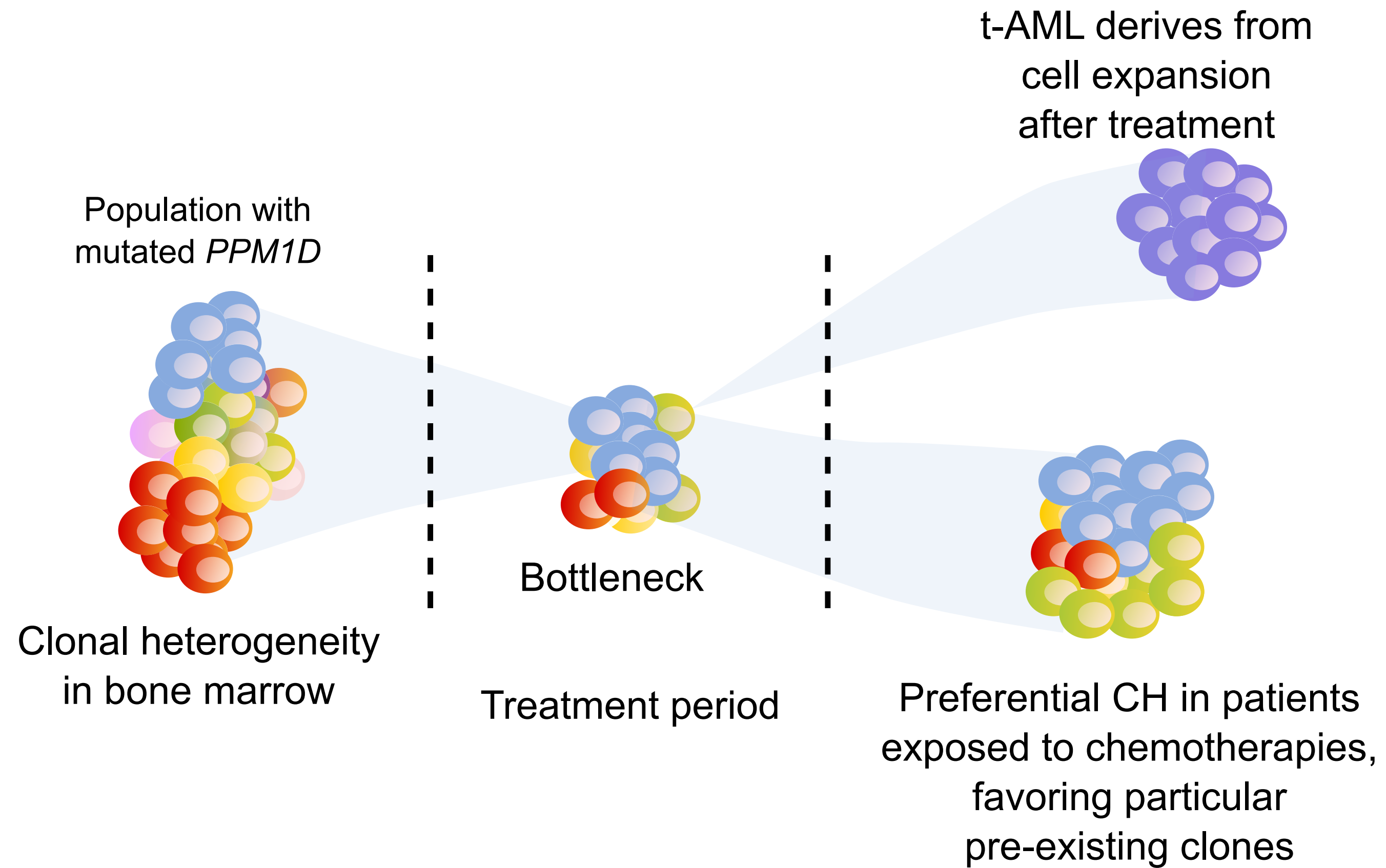
# Chemotherapy is associated with clonal hematopoiesis with preference for mutations in certain genes



Chemotherapy selects preexisting clones with specific mutations



# Evolution of hematopoietic cells under cancer therapy



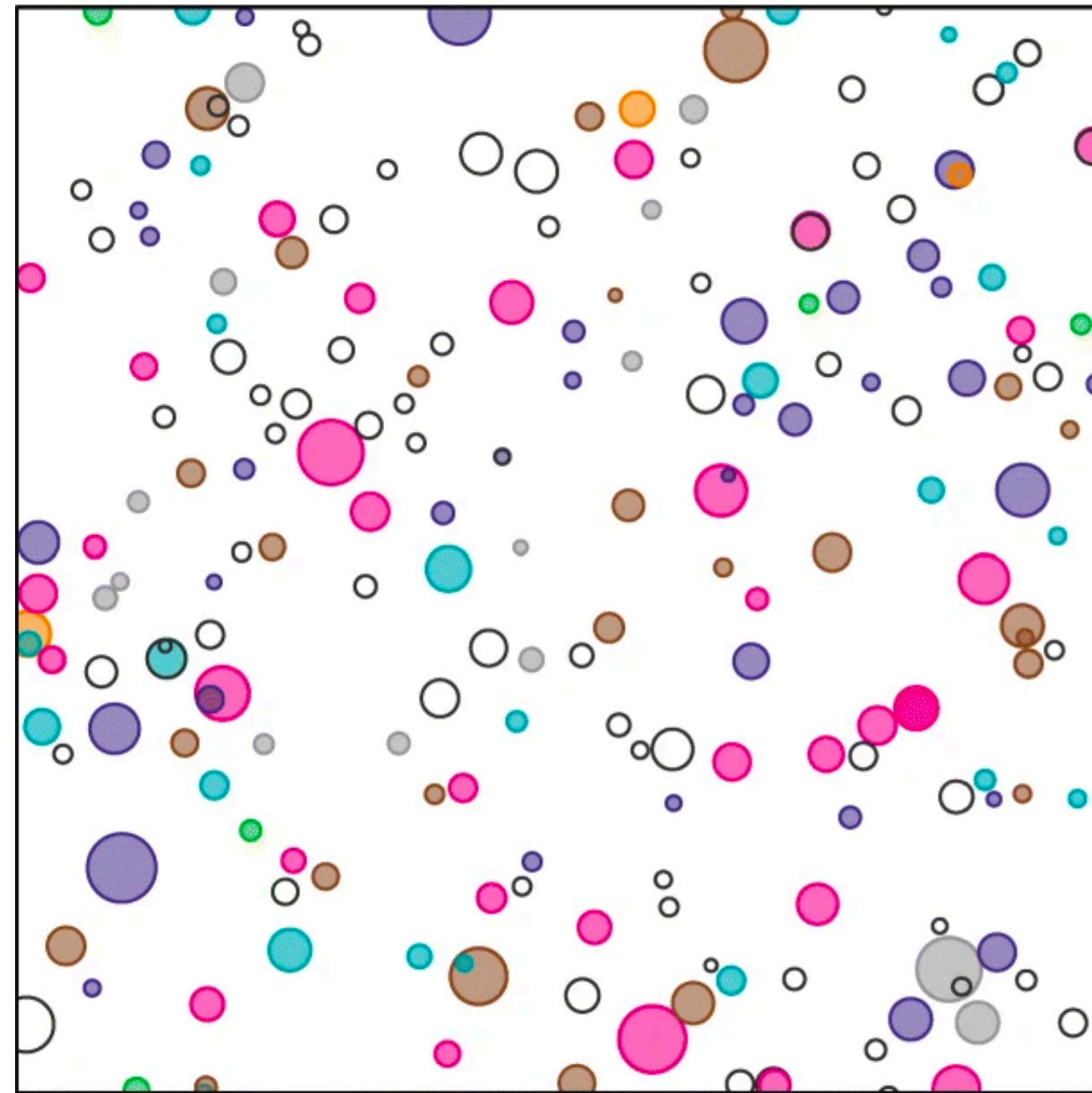
- Identifying Cancer Drivers
- Understanding Neutral Mutagenesis
- Identifying Clonal Hematopoiesis Drivers
- Chemotherapy effect in hematopoiesis
- Cancer Promotion

Driver mutations are necessary

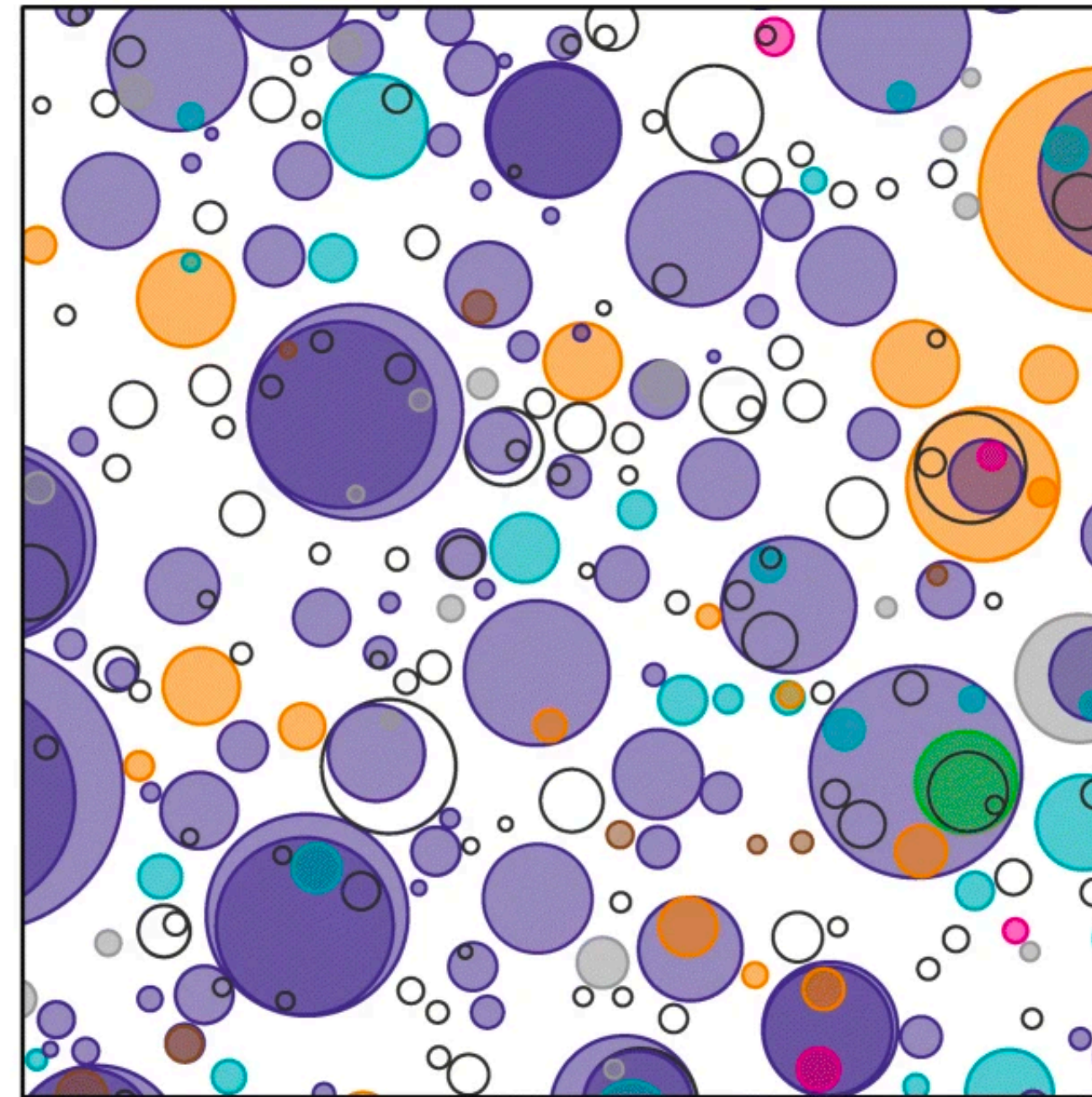


# Driver mutations are necessary but not sufficient

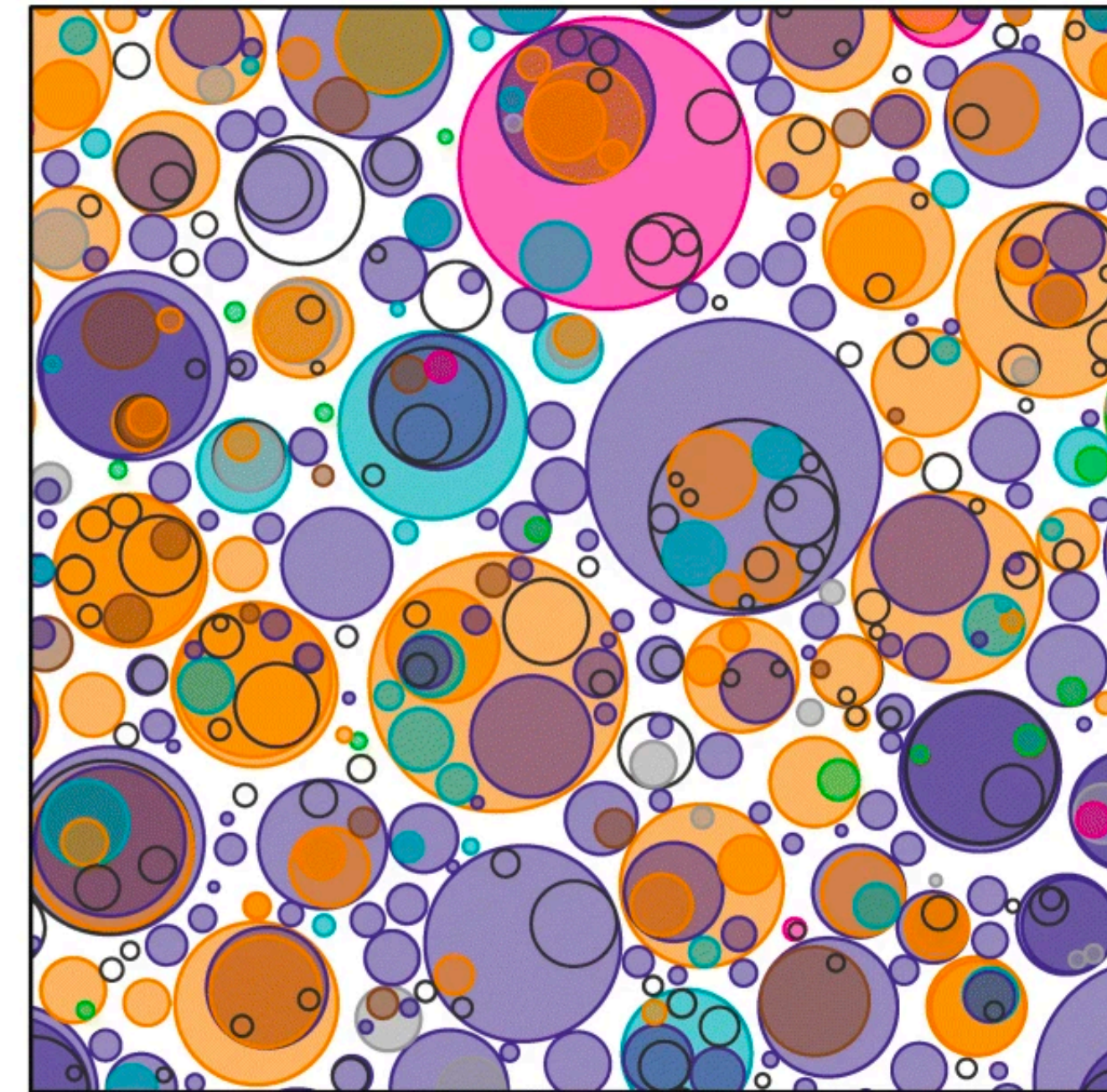
24–27 years old



52–55 years old



72–75 years old



■ *TP53* ■ *NOTCH1* ■ *NOTCH2* ■ *NOTCH3* ■ *FAT1* ■ *ARID1A* ■ Other driver genes □ Other non-driver genes

Mutant cell colonization of healthy esophageal epithelium with age

Martincorena Genome Biology 2019

Martincorena et al., Science 2015

Martincorena et al., Science 2018

Driver mutations and clonal expansions in normal tissue



# Many carcinogens are not mutagens

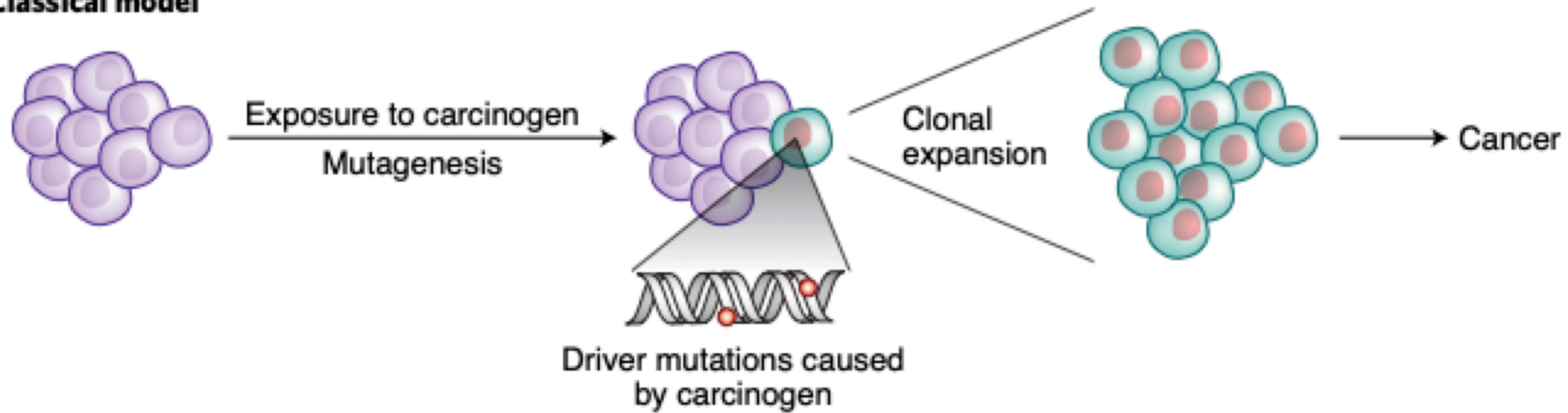


## The mutational signature profile of known and suspected human carcinogens in mice

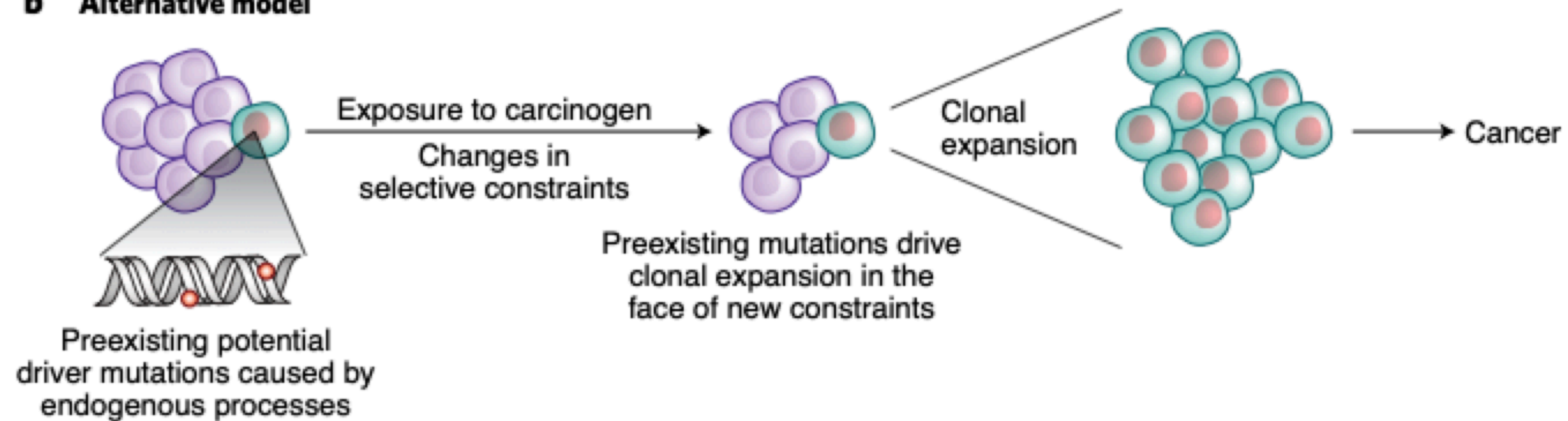
Laura Riva <sup>1,5</sup>, Arun R. Pandiri<sup>2,5</sup>, Yun Rose Li<sup>3,5</sup>, Alastair Droop<sup>1</sup>, James Hewinson<sup>1</sup>, Michael A. Quail<sup>1</sup>, Vivek Iyer<sup>1</sup>, Rebecca Shepherd<sup>1</sup>, Ronald A. Herbert<sup>2</sup>, Peter J. Campbell <sup>1</sup>, Robert C. Sills<sup>2</sup>, Ludmil B. Alexandrov <sup>4</sup>, Allan Balmain <sup>3,6</sup>  and David J. Adams <sup>1,6</sup> 

# Many carcinogens are not mutagens

## a Classical model



## b Alternative model



Abel Gonzalez-Perez





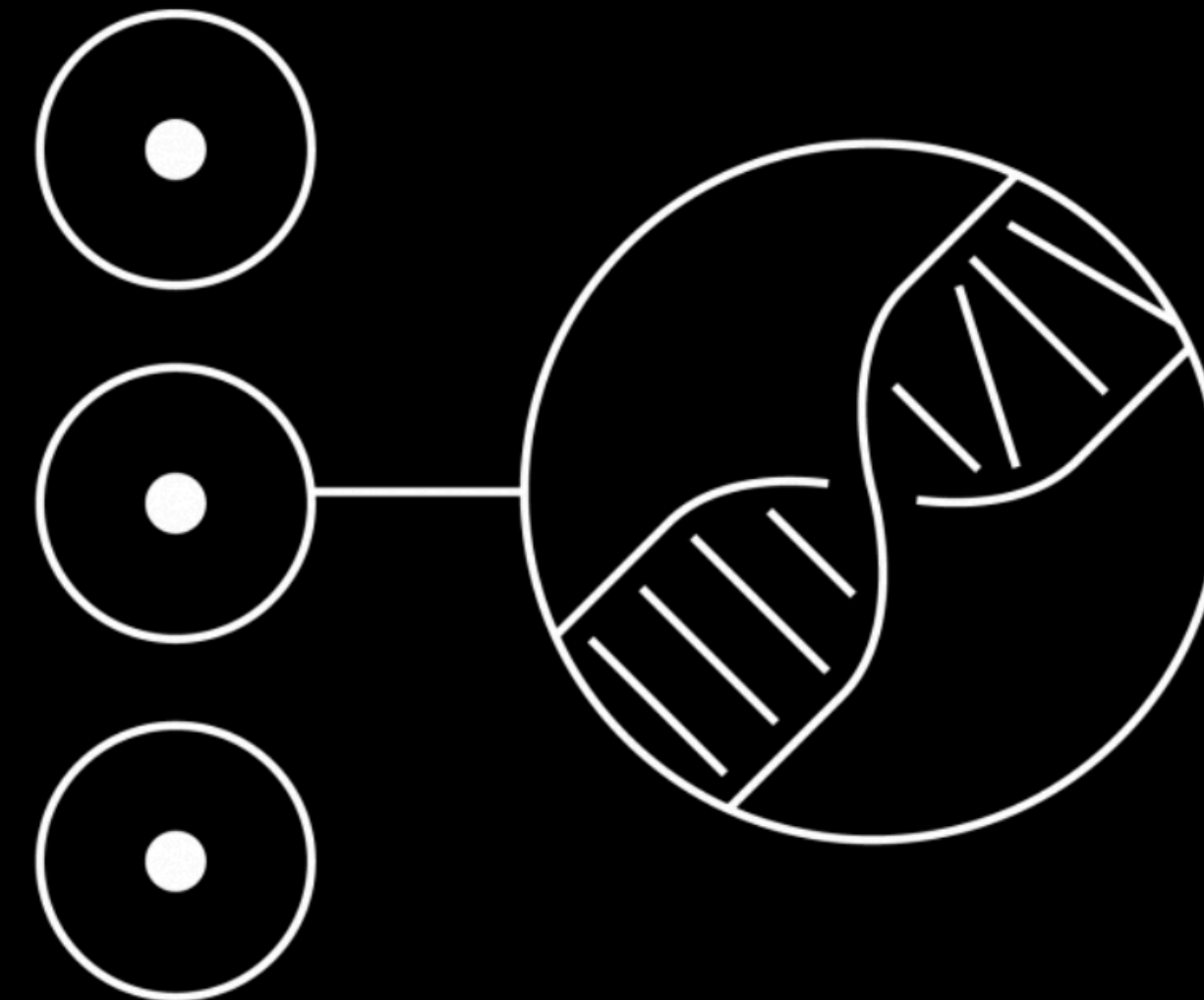
## Driving progress through unprecedented collaboration

Cancer Grand Challenges is a global funding initiative founded by Cancer Research UK and the National Cancer Institute. We set ambitious challenges, providing diverse, global teams with £20m to come together, think differently, with the aim to make the progress against cancer the world urgently needs.

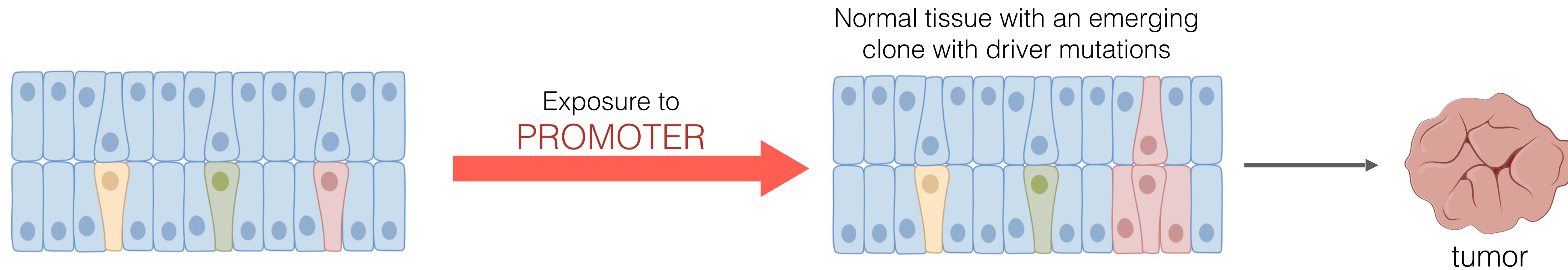
### Normal phenotypes

**CHALLENGE:**

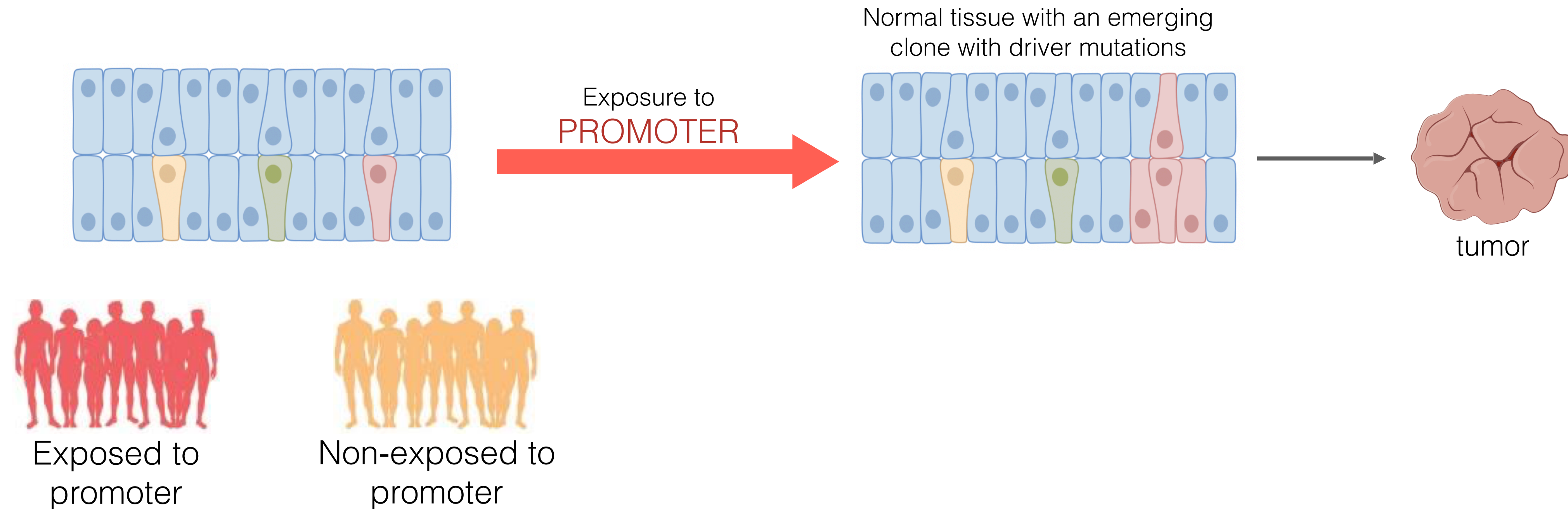
Understand how cells and tissues maintain “normal” phenotypes whilst harbouring oncogenic mutations and how they transition to become a tumour



# How does exposure to promoters change the normal tissue to eventually lead to cancer?



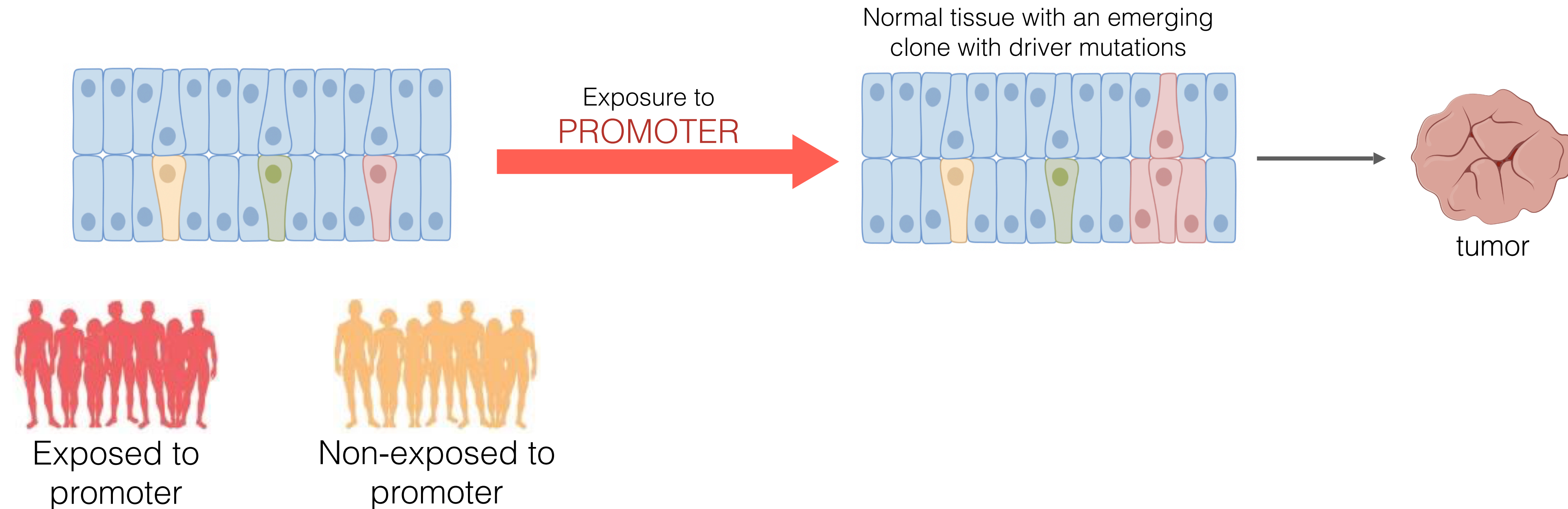
# How does exposure to promoters change the normal tissue to eventually lead to cancer?



Detect emerging clones with driver mutations in normal tissue of individuals exposed and non-exposed to a promoter



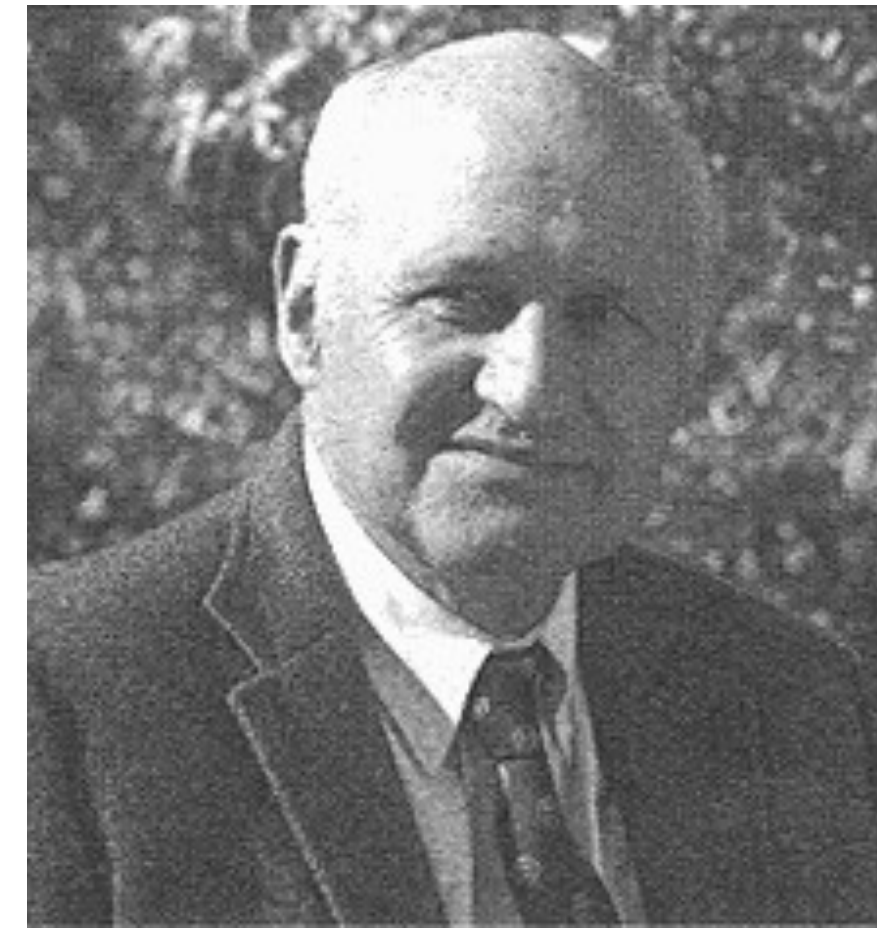
# How does exposure to promoters change the normal tissue to eventually lead to cancer?



Detect emerging clones with driver mutations in normal tissue of individuals exposed and non-exposed to a promoter

- Deep mutagenesis to identify emerging clones
- Single cell profiling with clone genotyping
- Spatial proteomics/transcriptomics with in situ mutation detection





Two stage model of carcinogenesis  
Initiation + Promotion

Berenblum and Shubik, 1947

*“....the initiating process represents a sudden and irreversible change in a small minority of the cells of the treated area, giving rise to isolated “latent tumour cells” .....  
“The presence of these latent tumour cells is only demonstrable by subsequent promoting action, which converts them into morphological tumours.”*



# PROMINENT TEAM

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International Agency  
for Research on Cancer



**Nuria LOPEZ-BIGAS**



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University of California  
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**Luke GILBERT**



University of California  
San Francisco



**Calvin J KUO**



**Marc GUNTER**

International Agency  
for Research on Cancer



**Emma LUNDBERG**



**Chris COUNTER**





# Thanks to:

Those who did the work



Those who generate and share data



And many others

Funding agencies

